# **UNIVERSITY OF BOTSWANA**

## BOTSWANA UNIVERSITY OF AGRICULTURE AND NATURAL RESOURCES



A dissertation submitted in partial fulfillment of the requirements for the award of the Master of Science Degree in Crop Science (Agronomy)

## EFFECT OF COMPOST AMENDMENTS ON SOIL PHYSICO-CHEMICAL PROPERTIES, PLANT GROWTH AND EFFICIENT USE OF WATER, NITROGEN AND PHOSPHORUS IN MAIZE (ZEA MAYS L).

By

## **ORATA TSHOMARELO NDUBO**

May, 2022

## Supervisor : Prof. Utlwang Batlang

## DEPARTMENT OF CROP AND SOIL SCIENCES

FACULTY OF AGRICULTURE

#### CERTIFICATION

Supervisor's name

Date

RAS LITENASTE BATLANG 09/05/2022

Head of Department

Date

Signature TA ....

....

Signature

Pool UTIMANY BATLANG 09/05/2522

i | Page

#### DECLARATION

I solemnly declare that this thesis represents my original work except where otherwise highlighted and has not been submitted for any award for a degree or of degree at any other universities.

ORATA TSHOMARELO NDUBO

...

May 2022

ii | Page

## APPROVAL

#### Supervisor's name

Date

MAY LITUNANE BATLANG

09/05/2022

Head of Department's name

Date

Prof. UTILWARE BATLANG 09/05/2022

Dean of Faculty'name

PKOF. JAMDIMO NGWAKO

Date

10/05/202

Signature AD

Signature

Signature

iii | Page

This work is wholeheartedly dedicated to my late father-in-law (May his soul rest in peace); Mr. Keletshabe Nkosi Ndubo who has encouraged me all the way and whose encouragement has made sure that I give it all it takes to finish that what I have started. I also devote this work to my husband Ndubo Ndubo and my two boys Jaden Abang and Colby Isang who have been a source of inspiration and gave me strength when I thought of giving up.

I would like first to thank the Almighty God for the strength he gave me to pursue this project to the end. I respectfully express my sincere thanks to my research supervisor Prof. Utlwang Batlang of the Department of Crop and Soil Sciences for guiding the progress of the experimental work, for the valuable discussions, suggestions in data analysis and interpretation, thoroughly and consistently guiding the writing of the thesis and for critical comments.

I would also like to thank Ms S. Lucas and the entire farmer's team for allowing me to collect soils for the experiments and conduct experiments on their farms. I would be honoured to convey my heartfelt thanks and sincere gratitude to Mrs. Chipo Nkomazana, Mr Mpho Keabotse and Mr Albert Makgwa of the Department of Agricultural Research for their contributions, assistance, and support throughout my research period. I would have not being able to conduct this thesis alone so I am thankful to Mrs. Cornelia Gwatidzo, Mrs Lesedi Molelekwa (may her soul rest in peace) and Mr T. Mathowa who I have enjoyed working with and learning from them.

I owe a deep sense of gratitude to my husband Ndubo Ndubo, for his keen interest on my work at every stage of my research. His prompt inspiration, timely suggestions with kindness, enthusiasm and dynamism has enabled me to complete my thesis. Special thanks to my two boys (Jaden Abang and Colby Isang) their understanding kept me going at all times.

Finally, I would like to thank the National Environment Fund (NEF) through Botswana University of Agriculture and Natural Resources "BUAN" for awarding me a scholarship to carry out this study. To God be the glory.

Compost is used across the world to sustain soil fertility and crop yield. This investigation consists of two experiments, which were conducted at BUAN greenhouses and farmers field in the Central District of Botswana in the period November 2018 to April 2019 involving maize (Zea mays.L). The experiments were carried out to test the hypotheses that soil amendments with compost as organic fertilizer improved soil physico-chemical properties, plant growth, WUE, NUE and PUE of maize under irrigated and drought stressed conditions. A completely randomized block design was used with the following treatments; (1) Soil + Urea (SU), (2) Soil + Urea + Compost (SUC), (3) Soil + Compost (SC), (4) and Soil (Control) replicated four times. The treatments for greenhouse trial comprised of two levels of irrigation, (W1= stressed at flowering stage, W2= irrigated). Data was collected on soil physico-chemical properties, plant growth, water use efficiency (WUE), nitrogen use efficiency (NUE) and phosphorus use efficiency (PUE). The results showed that compost amendments significantly increased soil pH, EC, CEC, total carbon, total N, available P and cation exchange capacity (CEC). This study also revealed that WUE improved in stressed plants compared to irrigated plants; drought stressed treatments were more water use efficient than the irrigated treatments. PUE and NUE in SC and SUC were maintained higher than in control and SU under both conditions. The highest NUE 24.66kg/kg was exhibited for SUC treatments under irrigated condition as compared to the rest of the treatments. Moreover, SUC and SC recorded the highest PUE 210.35kg/kg and 141.89kg/kg under irrigated condition and 24.71kg/kg and 20.37/kg/kg under drought stressed condition respectively. In conclusion combination of compost and urea fertilizer significantly enhanced NUE, PUE and WUE in maize. These could decrease

the amount of fertilizer and water-use required for the sustainable production in maize under unstressed and stressed environment.

*Key words:* compost, water use efficiency, nitrogen use efficiency, phosphorus use efficiency, drought stress

### **Table of Contents**

CER	TIFICATIONi				
DECLARATIONii					
APPI	APPROVALiii				
DED	ICATIONiv				
ACK	NOWLEDGEMENTSv				
ABSTRACT vi					
TAB	TABLE OF CONTENTSviii				
LIST	° OF TABLES xii				
LIST	OF FIGURES xiii				
LIST	OF SYMBOLS AND ABBREVIATIONxiv				
СНА	PTER ONE				
INTF	RODUCTION				
1.1 G	eneral introduction				
1.2 T	The maize crop and its eological physiology				
1.3 (	Constraints to maize production				
1.4 I	Maize contribution to cereal trends production in Botswana				
1.5 .	Justification of study7				
1.6 (	General objective				
1.7	Specific objectives				
1.8 l	Hypotheses				
CHA	<b>PTER TWO</b>				
LITE	ERATURE REVIEW				
2.1	Nutrient and water use efficiency of maize crop 10				
2.2	Growth responses to drought stress in maize11				
2.3	Physiological responses of maize under drought stress				
2.4	Water use efficiency and its relation to drought resistance in maize				

2.5 Nitrogen use efficiency and its relationship WUE under drought stress conditions
2.6 Phosphorus use efficiency and its relationship with water use efficiency
2.7 Soil nutrient management for improving WUE in crop production2
CHAPTER THREE
MATERIALS AND METHODS
3.1 <b>Description of Experimental sites</b>
3.1.1 Experiment I
3.1.1.1 Soil sampling and analysis
3.1.1.2 Experimental Design and Treatments
3.1.1.3 Planting and cultural practices
3.1.1.4 Data collection
3.1.1.4.1 Precipitation
3.1.1.4.2 Soil analysis
3.1.1.4.3 Plant Sampling and analysis
3.1.1.5 Statistical Data analysis
<b>3.1.2 Experiment II</b>
3.1.2.1 Soil sampling and analysis
3.1.2.2 Experimental design and treatments
3.1.2.3 Planting and cultural practices
3.1.2.4 Data collection
3.1.2.4.1. Soil and moisture content analysis
3.1.2.4.2 Plant growth parameters
3.1.2.4.3 Yield and yield components
3.1.2.4.4. Leaf gas exchange and chlorophyll content measurements
3.1.2.4.5 Determination of Water Use Efficiency
3.1.2.4.6 Plant analysis after harvest
3.1.2.4.7 Determination of nitrogen use efficiency (NUE)
3.1.2.4.8 Determination of phosphorus use efficiency (PUE)
3.1.2.4.9 Statistical Data Analysis
CHAPTER FOUR
RESULTS

4.1 Weather condition	at Lecheng extension areas	
4.2 Preliminary response	se of maize and stover accumulation as influenced by compost	
amendments under rain	n-fed conditioons	40
4.3 Effect of compost	amendments on maize nutrient concentration after harvest	
4.4 Effect of compost and after harvest of ma	amendments on soil chemical properties at 50% days to flower	ring stage 42
4.5 Effect of soil moist	ture content on maize leaf chlorophyll content at vegetative stag	ge 46
4.6 Response of maize drought stress	morphological characteristics as influenced by compost amend	dments and 47
4.6.1 Plant height at re	eproductive stage	47
4.6.2 Main stem diam	neter	
4.6.3 Number of leave	es	
4.6.4 Leaf area $(cm^2)$ .		
4.6.5 Days to 50% Flo	owering	
4.6.6 Days to 90% ph	ysiological maturity	
4.7 Impact of drought components	stress and compost amendments on maize yield contributing	51
4.7.1 Cob dry weight	(g)	
4.7.2 Number of grain	ns per cob	
4.7.3 100 seed weight	t (g)	51
4.8: Accumulation of 1 influenced by compost	maize biomass at vegetative and physiological maturity stages amendments and drought stress	as 54
4.8.1 Biomass at vege	etative stage	54
4.8.2 Shoot biomass a	at maturity	
4.8.3 Root biomass a	ıt maturity	55
4.8.4 Total biomass at	t physiological maturity	55
4.9 Nutrient concent	stration in maize at harvesting stage – Green house experiment.	58
4.10 a). Nitrogen use et	fficiency and phosphorus use ufficiency of maize at maturity st	tage 60
4.10 b). Nitrogen use e	efficiency in maize	60
4.10 c). Phosphorus use	e efficiency of maize under different water regime	60
4.11: Effect of compos	st amendments and drought stress on photosynthetic parameter	s of maize

CHAPTER FIVE
DISCUSSION
5.1 Characteristics of soils at the study site
5.2 Effect of compost amendment on soil chemical and physical properties
5.3 The effect of soil amendment under irrigated and drought stress conditions on morpho- physiological responses of maize
5.3.1 Plant height
5.3.2 Stem diameter
5.3.3 Number of leaves
5.3.4 Leaf area (cm <sup>2</sup> )
5.3.5 Days to 50% flowering
5.3.6 Days to 90% physiological maturity, yield and yield components
5.3 7 Maize biomass production
5.4 Effects of drought stress and compost amendment on chlorophyll content, photosynthetic rates and intrinsic water use efficiency of maize crop
5.4.1 Maize chlorophyll content
5.4.2 Photosynthetic rates and intrinsic water use efficiency
5.5 Effects of compost amendments on nutrient content, nitrogen and phosphorus use efficiency.
5.5.1 Nutrient concentration
5.5.2 Nitrogen use efficiency as influenced by compost amendments
5.5.3 Phosphorus use efficiency in maize as influenced by compost amendments
CHAPTER SIX
CONCLUSION AND RECOMMENDATIONS
6.1. Conclusions
6.2. Recommendations
REFERENCES

### List of Tables

Table 3.1 Soil classification at experiment 1 site 26
Table 3.2 Initial soil and compost physical and chemical properties before planting
Table 3.4 Initial soil and compost physical and chemical propeties used in greenhouseexperiment before planting
Table 4.1 Nutrient concentration in rain-fed field at harvesting stage
Table 4.2. Profile of the medium on soil chemical properties at flowering and after harvest45
Table 4.3 Respose of maize morphological charcteristictics as influenced by compost      amendments and drought stress
Table 4.4 Impact of drought stress and compost amendments on maize yield components53
Table 4.5 Biomass production of maize as influenced by compost application and drought56
Table 4.6 Nutrient concentration in maize at harvesting stage- greenhouse experiment
Table 4.7 Nitrogen use efficiency and phosphorus use efficiency of maize at maturity61
Table 4.8 Photosynthesis, transpiration, internal co2, stomatal conductance and water use      efficiency of maize at flowering stage

## List of Figures

Figure 1.2 Historical maize and sorghum production trends (1979-2015)	6
Figure 3.1 Soils at experimental site and surrounding areas	.26
Figure 3.3 Greenhouse soil treatments layout	.33
Figure 4.1 Rain gauge at Matlhakola and monthly rainfall recorded in Lecheng Agricultural Extension Area in the year 201/2019	39
Figure 4.2 Plant growth and stover accumulation as influenced by compost treaments	40
Figure 4.3. Effect of water stress and compost supplement on maize leaf chlorophyll content	46
Figure 4.4 An overview of maize performance as influenced by compost amendments	57

## LIST OF SYMBOLS AND ABBREVIATION

Α	Photosynthetic rate
ASS	Atomic Absorption Spectrophotometer
ATP	Adenosine Tri Phosphate
В	Boron
BUAN	Botswana University of Agriculture and Natural Resources
С	carbon
Ca	Calcium
CEC	Cation Exchange Capacity
CHL	Chlorophyll
Ci	Internal carbon dioxide gas
CO <sub>2</sub>	Carbon dioxide
E	Transpiration rate
EC	Electrical Conductivity
FAOSTAT	Food and Agricultural Organization Statistics
Fe	Iron
gs	Stomatal conductance
$H_2O_4$	Hydrogen peroxide
$H_2SO_4$	Sulphuric acid
HCl	Hydrochloric acid
ISPAAD	Integrated Support Programme for Arable Agriculture Development
Κ	Potassium
Kg	Kilogram
LSD	Least significant difference

MC	Moisture Content
Mg	Magnesium
Mn	Manganese
Ν	Nitrogen
Na	Sodium
NEF	National Environment Fund
NUE	Nitrogen use efficiency
OC	Organic carbon
Р	Phosphorus
PAR	Photosynthetic Active Radiation
рН	Potential of hydrogen
PUE	Phosphorus use efficiency
S	Sulphur
SAS	Statistical Analysis Software
SC	Soil + Compost
SOM	Soil Organic Matter
SU	Soil + Urea
SUC	Soil + Urea + Compost
TE	Transpiration Efficiency
WAE	Weeks After emergence
WUE	Water Use Efficiency
Zn	Zinc

### CHAPTER ONE

#### **INTRODUCTION**

#### **1.1 General introduction**

Productivity in agriculture and crop growth are influenced by numerous biotic and abiotic stresses such as temperature, drought, pest and diseases, the soil quality (Thalmann and Santelia, 2017). Soil fertility and plant nutrition are critical aspects of cropping system and these include suitable supply of needed nutrients. Hence, the supply of these essential nutrient elements is considered as one of the basic needs to achieve the potential yield. Conversely, the land continues to undergo human-induced soil degradation and loss of productivity. Explicitly, low soil fertility is a threat to smallholder farmers' livelihood in Sub-Saharan Africa (SSA) (Nigussie *et al.*, 2017). According to Havlin *et al.* (2014) as plants are removed from a field or soil sediments are transported offsite, nutrients in the soil are depleted. Natural soil nutrient supply depends on the soil's ability to buffer nutrient loss through crop removal. On the other hand, drought stress also restricts growth and photosynthetic activity (Graça *et al.*, 2010; Jangpromma *et al.*, 2010; Barbosa *et al.*, 2015). Poor management of plant growth resources such as nutrient and water management could leads to low use efficiency of resources due to poor productivity of losses through leaching.

Most soils when put under continuous cultivation decline in physico-chemical properties and leads to low crop yields after a few years of production. Management of soil fertility through use of chemical fertilizers is key to successful production of all agricultural produce. Chemical fertilizers have high nutrient content as compared to organic fertilizers and are rapidly taken up by plants. However, excess use of these chemical fertilizers can result in a number of complications, such as nutrient loss, surface water and groundwater contamination, soil acidification or basification, reductions in useful microbial communities and increased sensitivity to harmful insects (Chen, 2006). Additionally, Eurostat (2015) report stated that abundant use of fertilizers and pesticides also increases the risk that nutrients and pesticides run-off into surface and leach into groundwater. The acidity of the soil reduces crop phosphate intake, increases the concentration of harmful ions in the soil and hinders crop growth (Chandini *et al.*, 2019). The application of chemical fertilizer usually improves crop yield in the short-term (Zhang *et al.*, 2009), but it barely maintains and even decreases SOC and has negative environmental impacts, such as acidification and nutrient loss (Cai *et al.*, 2015).

Integration use of inorganic fertilizers with organic manures is a sustainable strategy for efficient nutrient usage which enhances efficiency of the chemical fertilizers while reducing nutrient losses (Schoebitz and Vidal, 2016). Combination of chemical fertilizer and compost to arable soils can lead to long-term increases in plant essential nutrients such as phosphorus (P), potassium (K), and magnesium (Mg) (Bulluck *et al.*, 2002; Wortman *et al.*, 2012a). The combined use of chemical and organic fertilizers is considered as a good method to sustain high crop yield and enhance soil organic carbon (SOC), but it is still unclear when and to what extent chemical fertilizers could be replaced by organic fertilizers. According to Wang *et al.* (2016) among the types of organic fertilizer applications, manure amendments are favored for increasing SOC stock and supplying nutrients to crops because they have higher SOC sequestration efficiency. Organic fertilizers application is a widely accepted strategy to sustain crop yield and SOC stock, and has significant effects on climate change mitigation (Lu *et al.* 2015) and also soil fertility sustainability (Liu *et al.* 2014).

In semi-arid lands composted organic material can be used as a source of important nutrients for sustainable crop productivity (Amanullah *et al.*, 2015). In Botswana a large proportion of soils are

characterized by high sand content and low organic matter content; hence, their management is very critical. Basically, sandy soils are of low productivity due to poor water holding capacity, low cation exchange capacity and low nutrient holding capacity. The productivity of these soils nevertheless, could be improved by compost amendments to sustain crop production. The use of compost as fertilizer has multiple benefits such as an increase in organic C content and microbial activity (Scotti et al., 2015), a greater concentration of plant nutrients like N, P K and Mg, and a root reinforcement (Donn et al., 2014). Soil structure can also be improved by binding between soil organic matter and clay particles by means of cation bridges and through stimulation of microbial activity and root growth (Farrell and Jones, 2009; Gao et al., 2010). Good soil structure favors air and water transfer in soils, seed germination and root growth and reduction in erosion. In a study Chakraborty et al. (2011) reported that organic amendments, once added to the soil, favors the growth and diversity of microbial communities, highlighting a strong correlation between soil biological fertility and soil organic C content. Studies had provided evidence that the use of compost as organic amendment positively affects soil fertility in terms of biological and enzymatic activities (Thangarajan et al., 2013), in particular under intensive farming systems (Scotti et al., 2015). The use of compost can affect soil microbial diversity, as reported by Zaccardelli et al. (2013a) who showed a clear positive effect on the number of spore-forming bacteria, with an increase directly correlated with the dose of compost. Successful use of compost relies on evaluating the soil to be amended followed by an evaluation of the compost and its properties. In general, organic soil amendments are often promoted as a tool for building soil quality through improved chemical, physical, and biological properties.

#### 1.2 The maize crop and its ecological physiology.

Maize (Zea mays L.) is an important cereal crop grown all over the world under a diverse type of climate. Rainfall of between 500 to 900mm is sufficient to grow a good crop of maize. However, in Botswana maize is grown under marginal rainfall conditions of 200 to 600mm which often exposes the crop to drought stress. Maize is a fast growing crop that requires abundant moisture. A significant reduction in maize yield due to water deficit even at high doses of nitrogen has been reported (Moser *et al.*, 2006). Management of irrigation water and nitrogen is crucial in order to improve maize productivity with reduced pollution risks (Gheysari *et al.*, 2009). Among the major cereal crops, maize is the only monoecious plant bearing unisexual flowers. The male inflorescence or tassel develops from the shoot apical meristem at the top of the plant, whereas the female inflorescence or ear develops from lateral meristems in the axil of leaves. These spatial arrangement of the flowers facilitates both selfing and crossing pollination (Morris, 2002).

Past studies indicate that for better performance, maize needs a pH of 5.8 - 7.0 (Albrecht *et al.*, 2005; Mallarino, 2011; Crouse & Denny, 2015). Kai *et al.* (2012) noted that crop performance is affected indirectly by low soil pH from aluminum and manganese toxicity which resulted from overly acidic conditions of the soil. Good sprouting of this crop is attained at soil temperature of 20-22°C. Optimum range of temperature for better crop growth and yield realization is 25–35°C (Zaidi *et al.*, 2017). Soil moisture of 60-70% field water capacity is most favorable for maize plant. Furthermore, extended low temperature less 5°C severally affects the crop. Being day neutral, maize crop can be cultivated throughout the year which leads to high yield levels in a short period of time.

#### **1.3 Constraints to maize production.**

Previous studies have observed several challenges contributing to reduced maize productivity, such as, combined effects of abiotic and biotic stresses, highly variable environmental conditions, use of inappropriate maize varieties and limited resources (Fanadzo *et al.*, 2009; Jacobson, 2013; Tandzi *et al.*, 2015). Soil moisture retention is low in many of Botswana's soils. This is also intensified by soil caking and layering resulting from inferior tilling technologies (Sustainable Agriculture and Rural Development, 2007).

Nitrogen is a determinant nutrient for plant growth and a good supply for the plant is also important for the uptake of other nutrients including potassium, phosphorous (Bell, 2016). It is also the essential element most frequently scarce in soils around the world. It is a key nutrient for obtaining maximum yield and quality, but also one of the most difficult element to optimize. Nitrogen encourages leaf growth and an inadequate supply means smaller leaves, reduced photosynthesis, and less total yield and protein. If nitrogen supply is too high, then excess leaf will be formed at the expense of grain, starch content will be reduced and yield may also suffer. Too much nitrogen may also delay maturity and result in lodged crops. The choices of suitable forms of fertilizer for the proper growth of the plant are governed by local natural conditions and variations in soil and climate with regard to their suitability for maize cultivation. Excessive N leaching from fields is a leading source for degradation of water resources (Ewing and Runck, 2015) while N fertilizer inputs are also linked to increased nitrous oxide (N<sub>2</sub>O) emissions (USEPA, 2014).

#### 1.4 Maize contribution to cereal trend production in Botswana.

Cereal production in Botswana is based on rain-fed farming and nearly 70% of Batswana are resident in rural areas, where agro-pastoralism is still regarded as the dominant livelihood system (Statistics Botswana, 2012). Bio-physical factors such as low and unreliable rainfall, recurrent droughts, very high summer temperatures and poor sandy soils characterize crop production environment in Botswana. The majority of traditional farming households in the country practice arable production with low factor endowment, which cause few of these farmers to utilize soil fertility enhancing strategies. The hostile environment and lack of fertilizer use lead to low cereal crop productivity of which maize is the most affected. Historical data (1979-2015) (Statistics Botswana, 2015) shows that arable crop sub-sector has the potential to produce sorghum yield exceeding 600kg/ha of cereal grain as was the case in 1995 (Figure 1.2). However, maize productivity hardly exceeds 200 kg/ha and this is evidence that it is vulnerable to unfavorable environment and poor nutrient management.



Figure 1.2: Historical Maize and sorghum production trends (1979-2015): Source (Statistics Botswana)

#### 1.5 Justification of study

Botswana soils are characterized by high sand content and these soils are mostly considered as very delicate with respect to agricultural production due to their very low nutrients and organic matter content (Boul *et al.*, 2003). Most of Batswana farmers are small scale holders who rely on dry-land production thus depending on rainfall as a sole source of water, hence this system of production exposes plants to drought and nutrient stress which results in low productivity. With the least rain water and the low soil nutrient in Botswana, this research focused on how the challenges of drought stress and low soil fertility can be improved by compost to increase productivity with emphasis on some of the key resources namely; water, nitrogen and phosphorus use efficiency.

In agricultural production, water and nutrients plays a major part for plant development, therefore addressing the problem of soil nutrient deficiency farmers are currently issued with free chemical fertilizers by the Ministry of Agricultural Development and Food Security through Integrated Support Programme for Arable Agriculture Development (ISPAAD) to solve the challenges of low agricultural productivity caused by poor soil fertility. However, continuous use of these chemical fertilizers is known to be a source of some greenhouse gases and pollution of underground water. The cost of these chemical fertilizers to the government of Botswana and the potential environmental risk posed by their overuse have awaken the interest in using compost in crop production in order to mitigate climate change effects and with the least rain water and the low soil nutrient in Botswana, this research focused on how the challenges of drought stress and low soil fertility can be improved by compost to intensify desirable use of resources (Water and nutrients) with the aim of maximizing available soil moisture and increasing yields. Combining compost and chemical fertilizers will ensure that the problems associated with the use of either compost or inorganic fertilizers are greatly reduced as the combination of both fertilizers complement each other. The combined application of compost and chemical fertilizers is also widely recognized as a way of increasing yield and improving productivity of the soil (Kassahun and Mekonnen, 2012).

### 1.6 General objective

• To investigate the effect of compost amendments on water, nitrogen and phosphorus use efficiency of maize.

## **1.7 Specific objectives**

- To determine the effect of compost amendments on soil physico-chemical properties under rain-fed, water deficit and irrigated conditions.
- To determine the influence of compost amendments on maize plant growth under water deficit and irrigated conditions.
- To determine the effect of compost amendments on maize WUE under water deficit and full irrigated conditions.
- To determine the effect of compost amendments on NUE and PUE under water deficit and irrigated conditions.

## **1.8 Hypotheses**

- 1) *Null hypothesis:* Amendment of soil with compost will not affect maize WUE under water deficit and irrigated conditions.
- *Alternative hypothesis:* Amendment of soil with compost will affect maize WUE under water deficit and irrigated conditions.

- 2) *Null hypothesis:* Amendment of soil with compost will not affect maize NUE and PUE under water deficit and irrigated conditions.
- *Alternative hypothesis*: Amendment of soil with compost will affect maize NUE and PUE under water deficit and irrigated conditions.

## CHAPTER TWO LITERATURE REVIEW

#### 2.1 Nutrients and water requirements of maize crop.

Efficient use of water and nutrients by crops demands for reviewed or new agricultural crop management practices to sustain agricultural production (Shrestha *et al.*, 2010). The term "nutrient uptake" when applied to crops refers to the process by which plant roots take up nutrients present in soil solution and such nutrients subsequently distributed to aerial portions of the plant. The process is affected mainly by environmental conditions, management practices, the concentration of nutrients and the form in which nutrients are present in the soil. Nutrient use efficiency is described as the ability of a plant to utilize soil available nutrients to result in measurable yield or yield parameters such as dry matter, fruit or grain production (Hati *et al.*, 2006).

Nitrogen and phosphorus are the two most limiting elements of crop production. According to Liu *et al.* (2015) maize plant is considered as a greedily plant to fertilization, particularly to nitrogen when irrigation water is available. The extent of N losses is also dependent on the form of N applied. Nitrate, for instance, is mostly present in soil solution given its low adsorption to soil particles (Marchi *et al.*, 2016). Hence, nitrate can be expected to leach below the root zone in furrow irrigation during periods of excess soil moisture conditions or by preferential flow, given its tendency to be transported by convection. Soil P deficiency is also one of the major factors limiting crop yields worldwide. Although required by plants in a smaller quantity compared with other primary macronutrients, the inadequate supply of P results in severe limitations in plant growth. Phosphorus use efficiency (PUE) on field treated with conventional chemical fertilizers has been reported low ranging between 5 - 40 % for many tropical soils (Syers *et al.*, 2008).

Water is considered as a major factor in nutrient availability to plants as it is the vehicle through which nutrients move through soil to access plant roots for uptake. Increasing global scarcity of water will also impact the way in which N fertilizer is accessed by plants and profoundly compromise crop productivity (Swarbreck et al., 2019). As water is a limiting resource in many regions of the world, the improvement of water-use efficiency (WUE) by crops is critical to maintaining food security. It is clear that improvement of WUE in crops must be accompanied by agronomic strategies to reduce water requirements in agriculture. Du et al, (2015) reported that mild water deficit does not lead to serious losses in yield, even increases production and is beneficial to improve WUE. In water-limited environments, plants should minimize water loss while maximizing carbon uptake to optimize their water-use strategies (Galiano et al., 2011; Adams et al., 2013; Mitchell et al., 2013). For this reason, water use efficiency (WUE) is an important indicator of plant adaptability and sustainability, especially under drought stress. Maize has been found to have high water use efficiency when compared to other crops as well as being highly nutrient efficient because it produces high biomass in linear response to nutrient availability without excessive evapotranspiration (Ogola et al., 2002). Under drought stressed environments, WUEg response to N supply will be closely associated with the timing and intensity of the water and N deficiencies. However, studies found that appropriate moisture regulation increased the WUE of maize and increased its ability to resist drought (Li et al., 2018). One of the most frequently used indices to evaluate the response of crops to a specific climatic condition and water supply is water use efficiency (WUE).

#### 2.2 Growth responses to drought stress in maize.

FAOSTAT (2014) has documented that almost 80% of the world s agricultural production is under rain-fed conditions and it provides 62% of the staple foods. However, the issue of water is crucial

for environmental sustainability of agriculture, because 60% of agriculture is located in semi-arid areas where regular water applications are necessary to complete the growth cycle of crops (Bhattacharya, 2019). According to Hussain et al. (2019), world maize yield and production are projected to decline by 15–20% per year due to heat and drought conditions, with these two factors becoming major threats to this crop. Maize is an extremely water-sensitive crop and most of the maize-grown areas are rain-fed. Maize susceptibility to drought is due to the plant's water requirement for cell elongation and its' inability to delay vegetative growth. Drought is considered as one of the major environmental stresses that limit plant growth and consequently, crop-yield. It causes a broad range of growth, photosynthetic, metabolic, and ultrastructural variations in plants (Cui et al., 2017). Drought affects various morpho-physiological processes including plant biomass, root length, shoot length, photosynthesis, water use efficiency (WUE) and leaf water content (Egilla et al., 2005; Abdul et al., 2008). Research showed that plants respond to water deficit and adapt to drought conditions through various physiological and biochemical changes including phenological modifications (Basu et al., 2016). A study by Anjum et al. (2011a) showed that drought stress in maize led to considerable decline in net photosynthesis (33.22%), transpiration rate (37.84%), stomatal conductance (25.54%), water use efficiency (50.87%), intrinsic water use efficiency (11.58%) and intercellular CO2 (5.86%) as compared to well water (WW) control.

Drought stress may occur during the stage of vegetative or generative transition in the shoot apical meristem depending on the cereal species and on the geographical location of plant cultivation. The appropriate corresponding pattern of inflorescence development and the time of flowering to the temporal variation in water availability is recognized as one of the most important traits conferring adaptation to drought. Maize has different responses to water deficit depending on developmental stages (Cakir, 2004). There are indications that maize is relatively less sensitive to water stress when this occurs during early vegetative growth stages, given the relatively reduced crop evapotranspiration (Steduto *et al.*, 2012). Drought stress that occur during reproductive phase, may lead to embryo abortion and pollen sterility in some cereal species (e.g., rice, maize) (Araus et al., 2011; Fischer et al., 2011), signifying that the effect of drought on yield during this period could not be returned by adding water afterwards therefore, the length of the stress period is also an important factor. Soil water depletion to the wilting percentage for two (2) days during the tasseling or pollination period can result in as much as a 22% decrease in yield, while a six (6) eight (8) days period of depletion can cause a yield reduction of about 50%. At the onset of drought, maize plants of all ages will wilt in the afternoon and recover turgidity at night. The stomata of maize plants when exposed to severe drought for three (3) or four (4) days resume much of their apparent normal behavior after one or two day recovery period. But, severe drought over longer periods of one week or more produces marked changes in stomatal behavior and they never again seem to be able to open fully. Independently from the mechanism by which stomata close, it has been proposed that stomatal regulation avoids excessive drop in leaf water potential by responding to nonlinearities in the relationship between transpiration rate and leaf water potential (Sperry and Love, 2015; Sperry et al., 2016).

Inadequate available soil water declines the metabolic activity of maize, lessens its biomass accumulation, and decreases its photosynthetic rate by reducing the chlorophyll content in leaves, eventually leading to a decrease in maize yield (Zhang *et al.*, 2009; Bu *et al.*, 2010). Drought stress at tasseling does not only hinders the plant's ability to flower and shed pollen, but also can greatly affect the viability of maize pollen, especially when the drought is accompanied by high temperatures as is usually the case. Such stress increases the time required for pollination and

delays silking. The result is that at times all the pollen may be shed before the silks emerge. Water stress limits maize growth and yield mainly due to reductions in its carbon-water balance (Zhang *et al.*, 2018)], which is largely dependent on photosynthesis (Campos *et al.*, 2014).

Plant physiological processes are highly sensitive to water stress, particularly those that are related to the plant organ water state (Zhang *et al.*, 2010). Generally, crop photosynthetic characteristics have been taken as critical indicators of plant growth since they are directly related to net productivity (Huang *et al.*, 2020). Drought stress from booting stage through approximately ten (10) days after anthesis will severely affect yield. At the seedling stage, drought stress is likely to damage secondary root development. In general, the combination of drought and high day time temperatures reduces the photosynthetic efficiency, stomatal conductance, leaf area, water use efficiency (WUE), and yield of plants (Sattar *et al.*, 2020).

#### 2.3 Physiological responses of maize under drought stress.

Water deficit limits crop productivity more severely than other environmental factors apart from soil fertility. The severity of drought is unpredictable as it depends on several factors such as occurrence and distribution of rainfall, evaporative demands and moisture storing capacity of soils (Abdulai, 2005). According to Vadez *et al.* (2011) genotypic variation in crop response to drought depends on agronomic, environmental and genetic factors. The physiological responses of maize plants to drought and heat can be classified into three different mechanisms: escape, avoidance, and tolerance. The photosynthetic and gas exchange responses are the most sensitive to water deficits and maintaining relatively high photosynthetic activity levels may enhance plant drought tolerance. In maize, the combination of drought and heat reduces rate, stomatal conductance, leaf area, and WUE (Sehgal *et al.*, 2017). According to Huo *et al.* (2016) photosynthetic systems are

susceptible to damage during responses to water deficit stress. Zhao *et al.* (2016) also reported that maize crops are extremely sensitive to heat and drought stress. Maize seedlings growing under water stress conditions exhibit several important physiological responses, including decreased cell turgor (Chen *et al.*, 2015; Gall *et al.*, 2015), leaf rolling (Kadioglu *et al.*, 2012), inhibited CO2 exchange and decreased photosynthetic efficiency and chlorophyll contents (Mao *et al.*, 2015; Bunce *et al.*, 2010). It is also believed that CO<sub>2</sub> assimilation by leaves is mainly reduced because of stomata closure in drought stress conditions (Farooq *et al.*, 2009). The reproductive stage is more sensitive to the combination of drought and heat than the vegetative stages (Obata *et al.*, 2015; Sehgal *et al.*, 2017). The reproductive processes most susceptible to the combination of heat and drought stress are pollen and stigma viability, pollen tube growth, early embryo development, flowering and seed filling, and number of kernels (Zandalinas *et al.*, 2017; Lamaoui *et al.*, 2018; Sehgal *et al.*, 2019).

#### 2.4 Water use efficiency and its relation to drought resistance in maize.

Crop plants require sufficient water if they are to grow to their optimum levels and water consumption varies greatly throughout the growing season depending on the environmental conditions they grow. WUE has been defined as the ratio of instantaneous photosynthesis (A) to transpiration (T) (Sun *et al.* 2015), which is mostly controlled by stomata opening and closure (Huang *et al.* 2017a). Intrinsic water use efficiency (WUEi) has also been utilized to reflect the biochemical characteristics of plants based on photosynthesis (A) and stomata conductance (g) (Beer *et al.* 2009; McCarthy *et al.* 2011).

Photosynthesis in C4 plants is more sensitive to drought periods due to stomatal closure and the reduction in the activity of photosynthetic enzymes compared to C3 plants (Ghannoum, 2009; Lipiec et al., 2013). Under thermal stress, photosynthesis in C4 plants shows a greater tolerance than in plants with C3 metabolism, associated with the accumulation of oxaloacetic acid within the bundle sheath cells. Maize is a C4 plant, which confers potentially more efficient use of CO<sub>2</sub>, solar radiation, water and N in photosynthesis than C3 crops. Species with the C4 photosynthetic pathway have evolved biochemical  $CO_2$  concentrating mechanisms that allow Rubisco to function in a high  $CO_2$  environment. The C4 biochemical pathway, in which the first products of photosynthesis are C4 carboxylic acids, and specific bundle sheath anatomy of leaves enable higher rates of photosynthesis than the C3 biochemical pathway. As C4 plants frequently, but not always, have lower stomatal conductance, the Transpiration Efficiency (TE) of C4 species is considerably greater than that of C3 species when directly compared in the same environment. This increases both nitrogen and WUE compared to C3 species. It is well accepted that crops differ in WUE abilities. Several other studies pointed that cereals tend to have higher WUE than oilseed crops (Norton and Waschsmann, 2006; Sadras and Mcdonald, 2012). Water use efficiency (WUE) of maize is approximately double that of C3 crops grown at the same sites. Crops are drought-sensitive at certain growth stages (Wang et al. 2011), whereas drought-tolerant at other phenological stages. Maintenance of water uptake by the development of deep roots into a wet profile will maintain the assimilation rate of leaves and there high WUE.

The soil system is viewed as the storage site for liquid water accessible to the plant through the root system. Soil texture and depth of the soil system determines the total water supply available to the plant. Another very important factor affecting soil water is soil structure which measures the type and degree of aggregation among soil particles defining the texture. Soil aggregates affect the ratio of macro-pores to micro-pores. This ratio affects infiltration and drainage of water and movement of gases in the soil system, therefore changing the soil nutrient status influences water

use efficiency as a result of the nutrient status of the soil that facilitate plant growth and ultimately the amount of biomass produced per unit of water consumed. Application of fertilizers promotes root growth which extracts soil moisture from deeper layers. It is known that proper nutrient levels in the soil will lead to increased crop growth and productivity and as such WUE (Yada, 2011).

#### 2.5 Nitrogen use efficiency and its relationship with WUE under drought stress conditions.

As much as water is a limiting factor in crop production, nutrition also has a greater influence on the final yield. Shortage of N constrains shoot and root growth, which reduces plant water and nutrient uptake capacity (Salvagiotti *et al.*, 2008; Setiyono *et al.*, 2010). Among all the plant nutrients essential for crop growth, nitrogen is the most limiting crop nutrient for most non-legume production systems (Zotarelli *et al.*, 2007) and thus the most limiting in crop production. Nitrogen use efficiency (NUE) is the degree to which N is used by plants, and specifically refers to the efficiency by which crops produce biomass or harvested product from a unit of acquired N (Bell, 2014).

From both a physiological and agronomic point of view, NUE is the result of two main biological processes: N uptake efficiency (NUpE) which corresponds to the amount of N taken up per unit of available N, and N utilization efficiency (NUtE) which corresponds to the increase in biomass or yield per unit of N taken up. During the plant developmental cycle, a number of complex physiological processes are involved in the control of plant NUE notably N uptake, N assimilation and N translocation. In many arid and semi-arid regions, WUE and NUE are often low due to low crop yields, degraded soil fertility and low and erratic water and N inputs (Rockstrom *et al.*, 2010; Sanchez, 2010; Rockstrom and Falkenmark, 2015). It was reported that in many developed and rapidly developing countries, WUE and NUE are also rather low because of over-application and

poor management (Sutton *et al.*, 2013). Cereals in general and maize in particular, need to remobilize the N accumulated in proteins in vegetative tissues and at the same time take up and assimilate N after anthesis, in order to ensure storage protein deposition in the grain.

A study Gebreyesus (2012) reported that soil moisture without soil fertility or fertility without soil moisture is less effective for production increment in the semi- arid areas. Nitrogen and water are directly related as nitrogen requires water to be fully dissolved in the soil for easy uptake by the plant roots. Crops may not be able to use nitrogen (N) efficiently if water is a limiting factor for growth and production. A limited supply of both water and N leads to a distorted crop development and growth or to low crop yields. Effects can be large when the supply of both is limited. Yared *et al.* (2010) also documented that reduction in low soil moisture can reduce nutrient uptake by roots and prompt nutrient deficiency by reducing the flow of nutrients from the soil to the roots, creating restrained transpiration rates and impairing active transport and membrane permeability.

Soil available water and N content are some of the major limiting factors for crop production (Lenssen *et al.*, 2007; Sainju *et al.*, 2009). Precipitation, being the major source of available water for dryland crops, needs to be used efficiently to sustain yield. The increasing use of inorganic fertilizer and neglect of organic fertilizer as a valuable source of nutrients have contributed to nutrient imbalance, low fertilizer use efficiency, deterioration in soil quality, nitrate leaching, soil acidification and carbon (C) loss which seriously limit crop productivity and soil nutrients. Most importantly, the significant increases in WUE under organic fertilizer treatment is not a function of higher water uptake. An integrated approach, combing application of compost with an application of artificial fertilizer is a good strategy for sustainable crop production (Gete *et al.*, 2010). Soheil *et al.* (2012) determined the effects of Municipal Waste Compost (MWC) on soil chemical properties and corn plant responses in pot experiment. They found that the amount of

available N, P and K and micronutrient/heavy-metal concentrations in soil increased as the result of waste compost application Addition of N and P fertilizer enhances root development, which improves the supply of other nutrients and water to the growing parts of the plants, resulting in an increased photosynthetic area and thereby more dry matter accumulation. This could ultimately increase WUE in crops such as maize. Recent studies have demonstrated that there are large differences in maize lines and hybrids in their ability to grow and yield well on soils with low mineral nutrient availability, which depends on both N-uptake efficiency and N-utilization efficiency (Hirel and Gallais, 2011). The effects of fertilizers on wheat yield and water use efficiency (WUE) have been intensively studied. Generally, the application of inorganic fertilizer can increase transpirational leaf area, creating excessive transpiration and water loss from the crop canopy, which in turn cause severe soil water depletion during the wheat-growing season in semiarid regions (Chen *et al.*, 2015). Management of nutrient supply is a strategy to improve WUE.

#### 2.6 Phosphorus use efficiency and its relationship with water use efficiency.

Phosphorus (P) is the most important essential nutrient for cereal production and animals (Wang *et al.*, 2017). It is known to be the second most limiting nutrient in crop production after nitrogen. It is a key element required for normal plant development, but its low mobility in soil results in poor uptake by plants, which consequently inhibits growth and metabolism. It is involved in several key plant functions including energy transfer, photosynthesis, transformation of sugars and starches, nutrient movement within the plant and transfer of genetic characteristics from generation after generation. The majority of soil types, including fertile soils, have low available phosphorus, because the rate of absorption in the rhizosphere exceeds the rate of its replenishment in soil

solution (Suriyagoda *et al.*, 2011). According to Conde *et al.* (2014) low P availability is one of the major factors limiting crop production in acidic soils.

Application of water within a certain range and phosphorus can effectively improve the absorption, transformation, and utilization of fertilizers by crops. Studies showed that appropriate fertilization can reduce the negative effects of soil water deficiency on crop growth and development to a certain degree (Yang, Guo, Wang, Yang, & Yang, 2012) and can also increase the phosphorus concentration in plants (Gu et al., 2018) with increased phosphorus uptake. Schärer et al. (2010) also noted that appropriate management of water and fertilizer can not only increase crop yield and reduce irrigation and phosphorus application but can also reduce total phosphorus and increase available phosphorus in soil. The use of P fertilizer reduces its deficiency in soil, increases the stress-tolerating ability of plants (Cortina et al., 2013) and results in adjustments of physiological, morphological, and biochemical processes that increase plant growth (dos Santos et al., 2004; Jones et al., 2005; Campbell and Sage, 2006; Faustino et al., 2013; Liu et al., 2015). In addition, anions as phosphorus showed an increased solubility subsequently to organic material application (Zaccardelli et al., 2013b; Scotti et al., 2015). P use efficiency (PUE) for cereal production in the world is too low, varying between 15 and 30% (Dhillon et al., 2017). Phosphorus use efficiency in maize fields is critically important, since this nutrient constitutes one of the most limiting factors to production. There are numerous definitions for PUE (White *et al.*, 2005; Hammond et al., 2009; Rose and Wissuwa, 2012). Phosphorus (P) uptake efficiency refers to the plants ability to obtain Pi from the soil, and P utilization efficiency to the capacity for biomass production using the P absorbed (Wang *et al.*, 2010). Increasing PUE can be achieved either by increasing uptake capacity or by optimizing its utilization (Shenoy and Kalagudi, 2005; Parentoni and Junior, 2008). Previous studies suggest that phosphorus contributes for the extension of root
system and P deficiency will increase drought stress (Cramer *et al.*, 2009; Sardans and Penuelas, 2012). Despite the importance of P in plant productivity, relatively few studies have assessed its effects on plant physiological and ecological processes under drought stress (dos Santos *et al.*, 2006; Naeem and Khan, 2009; Fleisher *et al.*, 2012; Jin *et al.*, 2015; Liu *et al.*, 2015).

The combination of water and fertilizer can effectively improve the water-use efficiency and phosphorus-use efficiency of alfalfa (Lenssen *et al.*, 2010), which is beneficial for reducing the loss of agricultural water in the field and the excessive use of phosphate fertilizers. Water and wind erosion are significant factors that contribute to low world PUE and represent an economic and environmental risk. When soil is subjected to erosion, P is also lost, further reducing crop productivity and ultimately, PUE (Schröder *et al.*, 2011). An assessment of soil P loss due to erosion was reported by Liu *et al.* (2008) who suggested that 13, 8, and 3 kg P ha are lost on an annual basis from arable land, overgrazed and normal pastures, respectively. According to Oyeyiola and Omueti. (2016) composted plant residues and animal waste materials mixed with rock phosphate have been demonstrated to enhance P availability and P use efficiency compared to rock phosphate alone on severely acid soils.

#### 2.7 Soil nutrient management for improving water use efficiency in crop production.

The best management practices to pursuit nutrient use efficiency include applying fertilizers according to plant needs and placed correctly to maximize uptake. Fertilizer does not only enhances plant growth but also stimulates root growth to allow water uptake from deeper soil layers, particularly during drought spells. The use appropriate types and quantities of nutrients from mineral and organic sources is an essential practice for improving nutrient efficiency.

Most literature in agricultural fields have reported that the mixed use of chemical fertilizer and organic fertilizers decreases the damage that can be induced by chemical fertilizers and improved crop productivity. Studies also have indicated that the combination of organic and inorganic fertilizers ensure greater synchrony between nutrient release and plant uptake and therefore increase crop yield (Mugwe *et al.*, 2009; Omotayo & Chukwuka, 2009). Poor soil fertility limits the ability of plants to efficiently use water (Bossio *et al.*, 2008). According to Mugwe *et al.* (2009), maize grown in soil enriched with organic materials and inorganic fertilizer had higher grain yield compared to the recommended rate of inorganic fertilizer. The combination is a result of enhanced nutrient use efficiency, improved synchronization of nutrient release and uptake by the crop, as well as reduced acidity and a more balanced supply of nutrients. Moreover, applying organic materials over several seasons results in increased yields, because the tannin and lignin content slows their decomposition and has a long-term effect on nutrient availability.

A number of studies indicated that the presence of organic matter in the soil is fundamental in maintaining the soil fertility and decreasing nutrient losses. Thus, compost is a good organic fertilizer as it contains nutrients as well as organic matter. Organic matter has number of important roles to play in soils, both in their physical structure and as a medium for biological activity. Many recent studies highlight the importance of soil organic matter (OM) with regard to climate change (Adewopo *et al.* 2014; Lin, 2014; Amundson *et al.* 2015; Baveye, 2015). Studies have also shown that compost input could increase the soil water-holding capacity (Fan *et al.*, 2005, and Wang *et al.*, 2011) and successfully match N availability with crop uptake thereby improving yield and WUE.

Application of compost increases soil physical fertility, mainly by improving aggregate stability, decreasing soil bulk density and increasing soil pore volume (Leroy *et al.*, 2007; Olabode *et al.*,

2007; Manivannan *et al.*, 2009). Another study compared three different soil types (two different loamy coarse sands and a coarse sandy loam) with sludge compost application of 50% v/v (Somerville *et al.*, 2018). All three soils had a reduced bulk density at both 3 (15–26% reduced) and 15 (14–25% reduced) months post compost application. Basically, it provides nutrients to the soil, improves its water holding capacity, and helps the soil to maintain good tilth and thereby better aeration for germinating seeds and plant root development (Edwards and Hailu, 2011). According to Van Camp *et al.* (2004) composting helps to optimize nutrient management and the land application of compost may contribute to combat soil organic matter decline and soil erosion.

Many experiments have shown that compost improves the aggregate strength of soils. This means that the soil is more resistant to compaction and roots can penetrate more easily to for reach water and nutrients absorption. Liu *et al.* (2013) found that organic fertilizer increased the soil water-holding capacity by increasing the percentage of macro-aggregates. Furthermore, compost addition increases soil organic matter (SOM) content, which enhances aggregation and stability, thereby ameliorating soil structure (Diacono and Montemurro, 2010). Compost contains organic molecules (chelators) that bind metal cations such as Fe, Cu, Zn and Mn, and maintaining them in a soluble state (Van Schoor, 2009). Chelate formation is important in the soil because it reduces the toxicity of plant nutrients and also minimizes unnecessary losses of nutrients through leaching, thereby making them available exactly when needed. In general compost has the ability to preserve nutrients from leaching away through water. Compost has two main effects on soils, particularly in nutrient poor soils: replenish soil organic matter and supply plant nutrients therefore addition of organic matter in the soil is a well-known practice to increase crop yields. Aziz *et al.* (2010) reported an influence of compost on plant growth. In maize these include; stem length, number of

leaves and leaf length were significantly influenced by the application of compost at different concentrations.

A study on maize (Zea mays L.) in acidic soil by Murmu *et al.* (2013) found that organic manure increases crop productivity, nitrogen utilization efficiency, and soil health compared to chemical fertilizer. Compost contains significant amounts of valuable plant nutrients including N, P, K, Ca, Mg and S as well as a variety of essential trace elements (Agegnehu *et al.*, 2014). Compost helps in retaining soil moisture, slow release of nutrients to crops and can lead to long-term yield increases. Importantly, using compost made from recycled resources is sustainable and can increase soil organic matter and water absorbing and holding capacity.

Cation exchange capacity (CEC) is one of the most important indicators for evaluating soil fertility, more specifically for nutrient retention. Low CEC soils are more susceptible to cation nutrient loss through leaching. This enables the soil to hold nutrients such as potassium, which would otherwise leach beyond rooting depth Mohammed *et al.* (2004); Agegnehu *et al.* (2014); proved that compost amendment resulted in an increase of CEC due to input of stabilized organic matter being rich in functional groups into soil.

## CHAPTER THREE MATERIALS AND METHODS

#### **3.1 Description of Experimental sites**

Two experiments were carried out under open field (Experiment I) and controlled environment in the greenhouse (Experiment II) during the 2018/19 growing season.

#### 3.1.1 Experiment I

This open field experiment was conducted in Matlhakola village in the Central District of Botswana in the period of November-April 2018/19 on a farmer's field. The site is located at latitude 22° 33' 0" S and longitude 27° 8' 0" E. The climate is considered to be a local steppe climate with an average annual rainfall of 427 mm. The average temperature is 20.1 °C. Soils at this experimental site are sandy and considered poor in fertility status. The dominant soils in this area are classified as Orthic Luvisols (Figure 3.1). Classification of soil at experimental plot in Matlhakola site is presented in Table 3.1. The experimental plot was 32m by 100m occupying a total area of 3200m<sup>2</sup> (0.32ha). The area was cultivated to clear it of weed and incorporate crop stubble from previous season crop.



**Figure 3.1:** Soils at experimental site and surrounding areas. Source: Ministry of Agricultural Development and Food Security: Geographical Information System.

Table 3.1 Soi	l classification	at Experiment	I site.
---------------	------------------	---------------	---------

Field Locatio	n	Symbol		Soil description FA	10
				Cl	assification
(Matlhakola)	•	G7-10a	•	Dark grayish brown massive	Orthic
				coarse sandy loam to sandy clay	luvisols
				loam.	
			•	Dark brown sandy clay loam to	
				clay.	
	Field Locatio (Matlhakola)	Field Location (Matlhakola) •	Field Location     Symbol       (Matlhakola)     • G7-10a	Field Location     Symbol     S       (Matlhakola)     • G7-10a     •	Field Location       Symbol       Soil description       FA         (Matlhakola)       • G7-10a       • Dark grayish brown massive coarse sandy loam to sandy clay loam.       • Dark brown sandy clay loam to sandy clay loam.         • Dark brown sandy clay loam to clay.       • Dark brown sandy clay loam to clay.       • Dark brown sandy clay loam to clay.

Source: Ministry of Agricultural Development and Food Security, Botswana Government.

#### **3.1.1.1 Soil sampling and analysis**

Prior to construction of the sunken seedbeds and planting, soil samples were collected at 30 cm depth (the required plough depth in sandy soils) in the 0.32 ha plot. Soil samples were collected systematically in a grid form (Tan, 2005). A composite sample of approximately 500g was then taken to the Soil Science Laboratory at Botswana University of Agriculture and Natural Resources, where it was further prepared and analyzed for pH, EC, CEC, exchangeable bases, organic carbon, available phosphorus and total nitrogen. Each analysis was done following established standard protocols and procedures. Results of soil analysis before planting classified the soil as sandy (Table 3.2). Textural class, key fertility and nutrient status are presented in Table 3.2. The soils have very low cation exchange capacity (CEC), organic carbon and acidic pH of 5.85. Analysis of the compost shows that it had higher fertility status than the experimental soil (Table 3.2).

Table 3.2: Initial Soil and Compost physical and chemical properties before planting.

Parameters	Soil	Compost
рН	5.99	7.29
EC (mS/cm)	0.077	1.920
CEC (cmol (+)/kg	3.78	12.10
Total Carbon (%)	1.009	1.120
Total Nitrogen (%)	0.009	0.020
Available P (g/kg)	0.014	0.072
Ca (cmol (+)/kg	1.391	21.275
Mg (cmol (+)/kg	0.614	0.472
K (cmol (+)/kg	0.515	4.321
Na (cmol (+)/kg	0.1190	5.1991
Bulk density (g/cm <sup>3</sup> )	1.43	-
Soil textural class	sand	-

#### **3.1.1.2 Experimental Design and Treatments**

On the 0.32 ha plot, sunken seedbeds were constructed each measuring 1mx1m and 0.3m deep. Sunken beds were constructed as a way of promoting conservation tillage and generally designed to reduce soil erosion and also improves water infiltration, water storage and thus yield potential and improve WUE. The spacing between each sunken bed was 1m\* 1m. A completely randomized block design was used to arrange the growth patterns for the different treatments. The following treatments were used: (1)Soil + Urea (SU), (2) Soil + Urea + Compost (SUC), (3) Soil + Compost (SC), (4) Soil (Control). Soil and compost at appropriate ratios of 2:3 by volume were uniformly distributed among the relevant treatments and mixed thoroughly. For control treatments, plots were also excavated and the same soil was replaced without amendments. Each treatment, including the control, was replicated eight times resulting in a total of 32 sunken beds.

#### 3.1.1.3 Planting and cultural practices

Seed of maize (Zea mays L.) variety Kalahari Early Pearl (KEP) from Seed Multiplication Unit (SMU) in the Department of Agricultural Research (DAR) were planted in the sunken seedbed. Plant spacing for each seedbed was 25cm by 25cm inter- and intra-row spacing and 5cm deep at the rate of 3 seeds/hill. Two (2) weeks after emergence (WAE) the seedlings were thinned to one per hill giving a plant population of 12 plants/sunken bed. At this stage, 50g of urea/ha/sunken bed was applied to the relevant treatments to balance N to the equivalence of 46kg N/ ha Urea. The quantity of urea application rate was calculated based on soil total N results (Table 3.2). Weeding was manual by hand hoeing and this started two (2) WAE to maintain the plots free of weed until sampling was carried out. It was necessary to maintain plots free of weed at all times because infestations differed depending on whether compost was used. The experiment was maintained as rain-fed from planting to sampling.

#### **3.1.1.4 Data collection**

#### 3.1.1.4.1 Precipitation

Rainfall was measured with a rain gauge placed in the middle of the field at the experimental site and the accumulated monthly rainfall for the whole growing season was calculated.

#### 3.1.1.4.2 Soil analysis

Soil samples were obtained before planting and after plant harvest. Soil pH was measured in 0.01 N calcium chloride solution using 1:2 soil to calcium chloride ratio. Soil EC was measured using portable EC meter in 1:2.5 soil to distilled water ratio as described by Reeuwijk. (2002). The CEC was determined by measuring ammonium concentration after the soil was extracted with ammonium acetate and then distilled and titrated with 0.01 hydrochloric acid as described by Reeuwijk. (2002). Organic carbon was analyzed by modified spectrophotometric Walkley and Black method as described by Souza *et al.* (2016). Available phosphorus in soil was extracted according to Bray and Kurtz method as described by Reeuwijk. (2002). Nitrogen percentage was determined by the micro–kjeldahl procedure which involve digestion in sulphuric acid–selenium mixture and hydrogen peroxide. The digest was distilled and ammonium was trapped into boric acid and the titrated with hydrochloric acid as described by Reeuwijk. (2002). Percentage soil nitrogen was then calculated using the formula by Estefan *et al.* (2013) below:

N% = [D \* 1.4007 \* (Va - Vb) \* N] Ws

Where:

Vs (ml) = volume of the acid used in the titration of the sample

Vo (ml) = volume of the acid used for blank titration

 $\mathbf{N} =$ molarity 0.01 for HCL

1.4007 (mg) = constant related to the molecular weight of the N

Ws(g) = Weight of the soil sample

#### **3.1.1.4.3 Plant Sampling and analysis**

Ten plants per plot were sampled in the middle row and transported to BUAN Plant Analysis Laboratory where they were dried at 72 °C until they reached content dry weight after 48 hours. The dried samples were weighed and ground into a fine powder and placed into air tight bottles according to the treatments and replications. The weight biomass was expressed as above ground biomass. The nutrient concentration of N, P, K, Ca, Na and Mg were determined by the micro-Kjeldahl procedure which involve the digestion in a concentrated H<sub>2</sub>SO<sub>4</sub> and H<sub>2</sub>O<sub>2</sub>. The plant digest were analyzed using Atomic Absorption Spectrophotometer (AAS) to find the concentration of Na, K, Ca, and Mg. Available P in plants was analyzed by modified spectrophotometric Walkley and Black method as described by Souza *et al.* (2016). The determination of total N in plants, the digest were distilled and titrated with 0.01 HCL and then total % N was calculated using the formula by Estefan *et al.* (2013) below:

N% = [D \* 1.4007 \* (Va - Vb) \* N] Ws

Where:

 $\mathbf{D}$  = dilution factor

**1.4007** (mg) = constant related to the molecular weight of N

Va (ml) = volume of the acid used in the titration of the sample

**Vb** (**ml**) = volume of the acid used for blank titration

 $\mathbf{N} =$ molarity 0.01 for HCL

Ws (g) = Weight of sample

#### **3.1.1.5 Statistical analysis**

Data collected was subjected to analysis of variance (ANOVA) using the Statistical Analysis System (SAS) version 9.2. Where a significant F-test was observed, treatment means were separated using the Least Significant Difference (LSD) at risk level of P < 0.05.

#### **3.1.2 Experiment II**

A pot experiment was conducted at BUAN campus in a greenhouse located at Sebele Content Farm, 12 Kilometers north of the Gaborone City. The site is situated at latitude (24°35′20′′S) and longitude (25°56′20′′E). Its estimated terrain elevation above sea level is 993 meters, in the South Eastern part of Botswana, which is a semi-arid climatic zone.

#### 3.1.2.1 Soil sampling and analysis

Pre- planting physico-chemical properties of the trial soil and compost are shown in Table 3.4. Before setting up the experiment, a soil sample of approximately 500 grams was collected from the field experimental plot for analysis of the following parameters; pH, EC, CEC, total N, available P, soil total carbon, and the exchangeable bases (K, Mg, Na and Ca). Each analysis was prepared according to established standard protocols and procedures. After setting up the green house experimental trial, samples were also collected from all the relevant treatments for analysis of total N and available P to be used in plants for the determination of NUE and PUE.

 Table 3.4: Initial Soil and Compost physical and chemical properties used in greenhouse

 experiment before planting.

Parameters	Soil	Compost
рН	5.85	7.30
EC (mS/cm)	0.04	4.76
CEC (cmol (+)/kg	3.32	23.7
Total Carbon (%)	0.22	1.22
Total Nitrogen (%)	0.002	0.02
Available P (g/kg)	0.01	1.36
Ca (cmol (+)/kg	1.548	32.190
Mg (cmol (+)/kg	0.601	5.847
K (cmol (+)/kg	0.076	24.024
Na (cmol (+)/kg	Non-detectable	5.716
Bulk density (g/cm <sup>3</sup> )	1.35	-
Soil textural class	sandy	-

#### **3.1.2.2 Experimental design and treatments**

The experimental design was 2\*4 factorial arranged as a Randomized Complete Block Design (RCBD). The treatments were; (1) Soil + Urea (SU) (2) Soil + Urea fertilizer +Compost (SUC), (3) Soil + Compost (SC), (4) Soil (Control). Ammonium (NH<sub>4</sub><sup>+</sup>) and nitrate (NO<sub>3</sub><sup>-</sup>) are the inorganic forms of plant available nitrogen in soils. Based on soil analysis results as presented in Table 3.4 above 8.5g of urea was applied to the relevant treatments to balance N to the equivalence of 46kg N/ ha Urea. The two factors were level of irrigation and soil amendment treatment. The

treatments comprised of two levels of irrigation, (W1= stressed at flowering stage, W2= irrigated). The crops received full irrigation until flowering stage, then irrigation was withheld from the other half of the maize crop whereby plants were subjected to progressive drought stress for approximately two weeks because maize water requirements tends to increase at periods of flowering and yield formation. Again during flowering period the uptake of nitrogen, phosphorus and potassium reaches a peak du Plessis. (2003). Water stress was imposed by withholding watering and allowing the soil moisture to be depleted naturally until the plants showed visible signs of wilting early in the morning. Once stress was established, plants were returned to the normal watering routine immediately. Well-watered plants were irrigated to field capacity throughout the experiment.

Thirty- two (32) cylindrical plastic pots of approximately (50L) 35 cm in diameter and 60 cm high with perforated bases were used. Sixteen pots (16) were filled with 72 kg of soil and the other sixteen pots (16) were filled with soil and compost at appropriate ratios of 2:3 in relevant treatments in the greenhouse set at 30°c then separated into two sets.

#### 3.1.2.3 Planting and cultural practices

Locally released maize (Zea mays L.) variety of Kalahari Early Pearl was obtained from Seed Multiplication Unit at the Department of Agricultural Research, Sebele. A total of five (5) seeds were sown per pot at a depth of 5cm and one week after emergence (WAE) seedlings were thinned to two per pot. Watering was done on alternate days using tap water. At the beginning of booting stage, there was an outbreak of sugarcane aphids (*Melanaphis sacchari*). Dectome insecticide was used to control aphids.



Figure 3.3 Greenhouse soil treatments layout.

#### 3.1.2.4 Data collection

#### 3.1.2.4.1. Soil and moisture content analysis

Soil pH, electrical conductivity, cation exchangeable capacity, organic carbon, soil available P, soil total nitrogen, and exchangeable bases (Na, Ca, K and Mg), were determined following the procedures described in experiment I section; 3.1.1.4.2. Soil moisture content was also monitored using MPKIT soil moisture sensor (ICT international, Armidale NSW Australia).

#### **3.1.2.4.2** Plant growth parameters

The following components were measured from the randomly selected plants.

a). Days to 50% emergence: Assessment of plant emergence percentage was done at 10 days after planting by counting the number of seedling that has emerged. The outcome was expressed as percentage of the total expected from each treatment.

**b). Number of leaves:** Only healthy and fully matured or opened leaves were counted from the selected plants at flowering stage.

c). Plant height determination (*cm*): Plant height at vegetative stage and at early flowering stage were obtained. Measurements were done by using a tape measure from the soil surface to the top of end of the flag at the beginning of booting.

**d**). **Stem diameter**: The thickness of the stem was measured using a caliper to the nearest 1mm. The measurements were taken from the lower part of the main stem.

e). Days to 50% tasseling: Number of days from sowing to the date on which 50% plants produced tassels was recorded.

**f**). **Days to 50% Silking:** Days to silking was also be counted from the date of sowing to the date on which 50% plants produced silk.

**g**). Days to 90% physiological maturity: Appearance of black layer in seeds was used as criteria for physiological maturity, and was calculated as difference between date of physiological maturity and date of emergence.

**h**). Leaf area: To determine leaf area, the length and the greatest width of selected leaves was measured with a ruler.

#### 3.1.2.4.3 Yield and yield components

a). Weight of cob (g): This was computed as the average weight from the randomly sampled plants using an electronic balance.

**b).** Grains per cob (g): The number of grains per each cob were recorded.

c). 100 seed weight (g): The 100 seeds were counted using the electronic seed counter and then weighed to the nearest 1mg.

d). Biomass yield: At harvest, roots were separated from the shoots and gently removed from the soil mass. Both the roots and shoots samples were obtained from all the pots. Samples were then oven dried at 72°C for 48 hrs. Weights of the dry roots and dry shoots were measured using a sensitive balance at resolution  $\pm$  0.00g. Biomass yield was calculated as follows:

Biomass yield = (DW/FW) \* 100

#### 3.1.2.4.4. Leaf gas exchange and chlorophyll content measurements

#### a). Gas exchange

Photosynthetic rate (*A*), Stomatal conductance (*gs*), Transpiration rate (*E*) and internal  $CO_2$  concentration (*Ci*) were measured using a portable photosynthesis system (LI- 6400/LI6400XT model). Readings were taken on the 2 fresh photo flag leaves.

#### **b).** Chlorophyll content

On daily basis the Minolta SPAD-502 plus, Konica Minoita meter was used to monitor chlorophyll concentration estimates on the fourth or fifth leaf down from the top of the plant. An average of three readings in leaf of each plant was used. The technique instantly measures the chlorophyll content of leaves by simply clamping the meter over the leaf and obtaining a chlorophyll content reading.

#### **3.1.2.4.5 Determination of Water Use Efficiency**

Photosynthetic water use efficiency was calculated by dividing leaf photosynthesis by leaf transpiration (Wilson *et al.*, 2012).

 $PWUE = Pn/Tr (\mu molCO_2 . m^{-2} S^{-1} / \mu molH_2 O. m^{-2} S^{-1})$ 

PWUEL is the leaf level water use efficiency in, µmolCO<sub>2</sub>/µmolH<sub>2</sub>O

Where;

**Pn** is the photosynthesis rate in  $\mu$ molCO<sub>2</sub>·m<sup>-2</sup>·s<sup>-1</sup>

**Tr** is the transpiration rate in  $\mu$ molH<sub>2</sub>O·m<sup>-2</sup>·s<sup>-1</sup>.

#### 3.1.2.4.6 Plant analysis after harvest

The nutrient concentration of N, P, K, Ca, Na and Mg were determined by the micro-Kjeldahl procedure which involve the digestion in a concentrated H<sub>2</sub>SO<sub>4</sub> and H<sub>2</sub>O<sub>2</sub>. The plant digest were analyzed using Atomic Absorption Spectrophotometer (AAS) to find the concentration of Na, K, Ca, and Mg. Available P in plants was analyzed by modified spectrophotometric Walkley and Black method as described by Souza *et al.* (2016). The determination of total N in plants, the digest were distilled and titrated with 0.01 HCL and then total % N was calculated using the formula by Estefan *et al.* (2013) elaborated in experiment I section: 3.1.1.4.3.

#### **3.1.2.4.7 Determination of Nitrogen Use Efficiency (NUE)**

It was calculated from biomass yield at sampling. The Micro Kjeldahl method (AOAC, 2011) was used for the determination of nitrogen (N) concentration and the calculation of nitrogen use efficiency (NUE) was done following the formula by Moll *et al.* (1982) below.

$$NUE(kg/kg) = \frac{Nt}{N \text{ soil}} X \frac{Bw}{Nt}$$

Where:

N soil = N supply from the soil plus the added N

**Nt** = total plant N at maturity

**Bw** = Aboveground Biomass weight at sampling

#### 3.1.2.4.8 Determination of phosphorus use efficiency (PUE)

It was calculated from the biomass yield at maturity using the formula by (Baligar et al., 2001).

 $PUE (kg/kg) = \frac{Bw(Total P in the plant (kg / plant))}{Available P}$ 

Where:

Available P = P available in the soil in kg / plant.

Bw = Aboveground Biomass weight at sampling

#### 3.1.2.4.9 Statistical Analysis

Data collected was subjected to analysis of variance (**AVONA**) using the Statistical Analysis System (SAS) version 9.2. Where a significant F-test with observed, treatment means were separated using the Least Significant Difference (LSD) at risk level of P<0.05. The P $\leq$ 0.05 was used to derive the significant difference unless stated otherwise.

### CHAPTER FOUR RESULTS

#### 4.1 Weather conditions at Lecheng extension areas

The extension area is composed of three villages, namely: Lecheng, Malaka and Matlhakola. The area received rainfall distribution as shown in (Figure 4.1) for the 2018/2019 growing season. Rains started very late around December 2018 and it is during this month when planting was implemented. Low rainfall was experienced in March 2019 when plants were at flowering stage. Generally, 2018/2019 growing season was a very dry year which resulted in poor plant stand and crop yields.



**Figure 4.1:** Rain gauge at Matlhakola (Figure 1a) and monthly rainfall recorded at Lecheng Agricultural Extension Area in 2018/2019 Figure 2b.

4.2 Preliminary response of maize and stover accumulation as influenced by compost amendments under rain-fed conditions at Lecheng Extension area during 2018/19.

In this rain-fed study it was observed that low rainfall experienced had strong effect on plant growth and biomass accumulation. However, treatments with compost amendments (SC and SUC) improved plant growth and stover production (Figure 4.2).



**Figure 4.2:** Plant growth and stover accumulated as influenced by compost treatment. Control (Soil), SU (Soil+ Urea fertilizer), SC (Soil+ Compost), SUC (Soil+ Compost +Urea fertilizer).

#### 4.3 Effect of compost amendments on maize nutrient concentration after harvest

The results presented in Table 4.1 shows maize nutrient content at harvest as influenced by compost amendments, under rain-fed condition. From Table 4.1, it was observed that there was no significant variation among the treatments with regard to N, Ca and Na level of concentration in maize. However, significant treatment differences were observed in maize plants with regard to P, K and Mg nutrient contents. Soil amended with compost resulted in higher levels of P, K and Mg in maize than those in the control treatments. It is worth noting that compared to controls, SU, SC and SUC treated plants had high nitrogen content although this is not statistically significant. Compared to the controls maize grown on soil amended with compost (SC and SUC) and urea fertilizer (SU) had high total P content. With regard to K and Mg nutrient concentration, a similar observation was made among the treatments where the highest and lower values were recorded in SC and control treatments respectively.

	Ν	Р	K %/plant	Ca	Mg	Na
parameters			nutrient content			
Control	0.1872 <sup>a</sup>	0.00054 <sup>b</sup>	2.68717 <sup>b</sup>	0.19853 <sup>a</sup>	0.15389 <sup>b</sup>	$0.00947^{a}$
SU SC SUC	$0.2015^{a}$ $0.2057^{a}$ $0.2202^{a}$	$\begin{array}{c} 0.00375^{a} \\ 0.00380^{a} \\ 0.00411^{a} \end{array}$	$\begin{array}{c} 2.16450^{b} \\ 6.26570^{a} \\ 4.82460^{a} \end{array}$	$\begin{array}{c} 0.19489^{a} \\ 0.25467^{a} \\ 0.21240^{a} \end{array}$	$0.17284^{b}$ $0.23069^{a}$ $0.18305^{b}$	$\begin{array}{c} 0.00894^{a} \\ 0.00788^{a} \\ 0.00762^{a} \end{array}$
LSD0.05	0.0665	0.0007	1.9277	0.0741	0.0442	0.0022

Table 4.1 Nutrient concentration in rain-fed maize at harvesting stage.

<sup>a</sup>Means followed by the same letters in the same column are not significant at P<0.05; The following acrynoms and words stand for: LSD - Least significant difference, Control - Soil, SU - Soil +Urea fertilizer, SC - Soil +Compost and SUC - Soil + Compost +Urea fertilizer.

# 4.4 Effect of compost amendments on soil chemical properties at 50% days to flowering stage and after harvest of maize crop.

Data in Table.4.2 shows the pH, EC, CEC, total carbon, exchangeable cations, total N, and available P concentration in soil at mid-growth stage and after harvest of maize as affected by compost amendments, under irrigation and water stressed conditions.

At flowering stage the pH values obtained with SU, SC and SUC were significantly higher than the control (Table 4.2). It is clear that application of compost amendments and urea fertilizer significantly increased soil pH values when compared to control. The results shows that SC treatment (6.54 and 6.56), SUC (6.50 and 6.52) and SU (6.26 and 6.25) had the highest effect in increasing pH values under irrigated as well as drought stressed experiments respectively. Control had the lowest pH values (5.77 and 5.85) under both irrigated and drought stress conditions respectively, which implies that addition of urea and amendment of soil with compost increased growth medium pH.

The results of EC revealed a significant change among treatments. The obtained results show that the application of compost under both conditions increased the EC values as compared to control treatments. Statistically the highest EC values were SC treatments (1.41dS/m and 1.78dS/m) and SUC treatments (1.39dS/m and 1.59dS/m), whereas the least values were recorded in SU treatments (0.11dS/m and 0.08dS/m) and control treatment (0.05dS/m and 0.07dS/m). Generally, control treatments had the lowest EC values under irrigated as well as water stressed conditions, which means amendment of soil with compost increased EC of the growth medium.

The results indicated that the values of CEC were significantly higher in soil amended with compost than the control and SU treatments. The highest CEC values were found in SC and SUC

treatments (9.49 and 9.42cmol (+) /kg and 11.2 and 9.19cmol (+)/kg) respectively under both condition while the lowest CEC values were shown in SU (2.29 and 2.45cmol (+)/kg) and control treatments ranging between 2.19 and 2.42cmol (+)/kg under both conditions. Therefore amendment of soil with compost (SC and SUC) increased media CEC.

As shown in Table 4.2 compost amendments significantly increased total carbon compared to the control. The results revealed that the highest percent values were recorded in SUC (0.93 and 0.59) followed by SC treatments (0.50 and 0.50) under both conditions. The least total carbon values were obtained in SU (0.23 and 0.27) and control treatments (0.26 and 0.23) under irrigated as well as drought stress conditions.

The results also showed that total N and available P concentration in soil significantly responded to compost amendments and urea (P<0.05). The soils treated with compost and urea had more total N compared to soil without compost (control). Statistically there was no significant difference between the treatment means with respect to soil available P under both conditions. Compared to the control and SU treatments, application of compost amendments alone or in combination with urea increased the exchangeable Ca and K contents under irrigated as well as non- irrigated conditions. The highest Ca values were observed in SC treatment (1046.9 and 617.1cmol (+)/ kg) and SUC (907.8 and 374.1 cmol (+)/ kg). The least Ca value was obtained with SU (32.4cmol (+)/ kg) and control (37.1 cmol (+)/ kg) under drought stressed condition. Similar to exchangeable Ca, soil exchangeable K and Mg exhibited a significant increasing trend with compost amendment. The highest K values were obtained with SC (392.2 and 549.1cmol (+)/ kg) and SUC (318.8 and 582.8cmol (+)/ kg) in comparison with SU (30.8 and 29.9cmol (+)/ kg) and control (17.1 and 31.7cmol (+)/ kg) under both irrigated and water stressed conditions. As for Mg concentration,

SUC treatment had the greatest values as compared to the rest of the treatments. The concentration of exchangeable Na was non-detectable under both conditions and similar results were revealed after harvest period.

Data was also collected at the end of the trial, which was at the time of harvest. Data shows that soil pH, EC, CEC, total carbon, total N, available P and exchangeable cations significantly responded to compost amendments with the exception of Na (Table 4.2). The highest pH values were obtained in SC, SU and SUC treatments. Generally the amendments of the acidic soil pH (5.85) with compost raised pH values. Similar trend was observed with EC results in both experiments. CEC as one of the major soil quality indexes was highly significant (P<0.0001). The highest CEC values were recorded in SC and SUC treatments; (12.4cmol (+)/kg and 9.42cmol (+)/kg), (10.9cmol (+)/kg and 11.2cmol (+)/kg) respectively. The lowest CEC values were shown in control (2.79 and 2.19cmol (+)/kg). As indicated total carbon was significantly greater in treatments incorporated with compost and urea fertilizer as compared to control treatments. With regard to total N and available P, the results revealed that there was significant increase with SU, SC and SUC treatments as compared to control treatments. Application of compost (SC) amendments alone or in combination with urea (SUC) showed marked increase in exchangeable Ca under irrigated as well as non-irrigated conditions over control and SU treatments (Table 4.2). The highest Mg concentration was noted with SUC treatments (308.2 and 599.1 cmol (+)/kg) as compared to the rest of the treatments. Statistically, for K soil content the highest values were reflected in SC (279.8 and 250.7cmol (+)/kg) followed by SUC (193.4 and 159.6cmol (+)/kg) treatments while the least values were recorded in SU (27.9 and 29.2cmol (+)/kg).and control (29.8 and 27.2 cmol (+)/kg under both conditions.

Treatments	Contro	l	SU		SC		SUC	
Parameters	Ι	D	Ι	D	Ι	D	Ι	D
Flowering stage								
pH (cacl <sub>2</sub> )	5.77 <sup>b</sup>	5.85 <sup>b</sup>	6.26 <sup>ab</sup>	6.25 <sup>a</sup>	6.54 <sup>a</sup>	6.56 <sup>a</sup>	6.50 <sup>a</sup>	6.52 <sup>a</sup>
EC (dS/m)	$0.05^{b}$	0.07 <sup>c</sup>	0.11 <sup>b</sup>	$0.08^{\circ}$	1.41 <sup>a</sup>	$1.78^{a}$	1.39 <sup>a</sup>	1.59 <sup>b</sup>
CEC(cmol(+)/kg	2.42 <sup>b</sup>	2.19 <sup>b</sup>	2.29 <sup>b</sup>	2.45 <sup>b</sup>	9.49 <sup>a</sup>	9.42 <sup>a</sup>	9.19 <sup>a</sup>	11.2 <sup>a</sup>
Total carbon (%)	0.26 <sup>b</sup>	0.23 <sup>c</sup>	0.23 <sup>c</sup>	0.27 <sup>b</sup>	$0.50^{ab}$	$0.50^{b}$	0.93 <sup>a</sup>	0.59 <sup>a</sup>
Total N (%)	0.001 <sup>b</sup>	0.001 <sup>b</sup>	0.004 <sup>a</sup>	0.003 <sup>a</sup>	0.003 <sup>a</sup>	0.003 <sup>a</sup>	0.003 <sup>a</sup>	$0.004^{a}$
Available P(g/kg)	0.02 <sup>a</sup>	0.02 <sup>a</sup>	0.03 <sup>a</sup>	0.02 <sup>a</sup>	0.03 <sup>a</sup>	0.03 <sup>a</sup>	0.03 <sup>a</sup>	0.03 <sup>a</sup>
Ca(cmol(+)/kg	72.1 <sup>b</sup>	37.1 <sup>c</sup>	63.5 <sup>b</sup>	32.4 <sup>c</sup>	1046.9 <sup>a</sup>	617.1 <sup>a</sup>	907.8 <sup>a</sup>	374.1 <sup>ab</sup>
Mg(cmol(+)/kg	15.8 <sup>c</sup>	12.4 <sup>b</sup>	10.1 <sup>c</sup>	6.5 <sup>b</sup>	191.0 <sup>b</sup>	39.3 <sup>b</sup>	334.9 <sup>a</sup>	352.9 <sup>a</sup>
K(cmol(+)/kg	17.1 <sup>b</sup>	31.7 <sup>b</sup>	30.8 <sup>b</sup>	29.9 <sup>b</sup>	392.2ª	549.1 <sup>a</sup>	318.8 <sup>a</sup>	582.8 <sup>a</sup>
Na(cmol/kg)	ND	ND	ND	ND	ND	ND	ND	ND
<u>End of trial</u>	1			,			,	
$pH(cacl_2)$	6.45 <sup>b</sup>	5.95 <sup>c</sup>	7.36 <sup>a</sup>	6.45 <sup>b</sup>	7.25 <sup>a</sup>	7.23 <sup>a</sup>	6.69 <sup>b</sup>	7.04 <sup>a</sup>
EC (dS/m)	0.07 <sup>b</sup>	$0.06^{c}$	0.08 <sup>b</sup>	0.11 <sup>b</sup>	1.79 <sup>a</sup>	1.39 <sup>a</sup>	1.61 <sup>a</sup>	1.75 <sup>b</sup>
CEC(cmol(+)/kg	2.79 <sup>b</sup>	2.19 <sup>b</sup>	3.27 <sup>b</sup>	2.46 <sup>b</sup>	$12.40^{a}$	9.42 <sup>a</sup>	10.9 <sup>a</sup>	11.2 <sup>a</sup>
Total carbon (%)	$0.27^{b}$	0.16 <sup>c</sup>	$0.45^{a}$	$0.67^{a}$	0.49 <sup>a</sup>	$0.50^{a}$	$0.52^{a}$	0.44 <sup>b</sup>
Total N (%)	$0.002^{b}$	$0.002^{b}$	0.010 <sup>a</sup>	0.003 <sup>a</sup>	$0.002^{a}$	$0.002^{a}$	$0.002^{a}$	$0.002^{a}$
Available P(g/kg)	$0.02^{b}$	$0.02^{c}$	$0.04^{a}$	0.04 <sup>a</sup>	0.01 <sup>a</sup>	0.01 <sup>a</sup>	$0.04^{a}$	0.03 <sup>b</sup>
Ca(cmol(+)/kg	69.9 <sup>b</sup>	29.8 <sup>b</sup>	59.8 <sup>b</sup>	23.6 <sup>b</sup>	460.3 <sup>a</sup>	295.3 <sup>a</sup>	438.4 <sup>a</sup>	$204.3^{ab}$
Mg(cmol(+)/kg	16.9 <sup>b</sup>	11.2 <sup>c</sup>	46.4 <sup>b</sup>	47.8 <sup>b</sup>	129.2 <sup>b</sup>	276.9 <sup>b</sup>	308.2 <sup>a</sup>	599.1 <sup>a</sup>
K(cmol(+)/kg	29.8 <sup>c</sup>	27.2 <sup>c</sup>	27.9 <sup>c</sup>	29.2 <sup>c</sup>	279.8 <sup>a</sup>	250.7 <sup>a</sup>	159.6 <sup>b</sup>	193.4 <sup>b</sup>
Na(cmol/kg)	ND	ND	ND	ND	ND	ND	ND	ND

Table 4.2: Profile of the medium on soil chemical properties at flowering and after harvest in the green house.

<sup>a</sup>Means followed by the same letters in the same column are not significant at P< 0.05. I and D represent irrigated and drought stressed conditions respectively, ND represent not detectable. Control (Soil), SU (Soil + Urea fertilizer), SC (Soil + Compost), SUC (Soil + Compost + Urea fertilizer).

#### 4.5 Effect of soil moisture content on maize leaf chlorophyll content at vegetative stage

Results of ANOVA showed significant differences among treatments with regard to moisture content and chlorophyll content in both conditions (P< 0.05) (Figure 4.3). Comparatively there was no variation within drought stressed and irrigated treatments with respect to chlorophyll content although statistically SUC had the highest chlorophyll with 50.33 followed by SC and SU treatments with 34.90 and 34.75 SPAD values respectively whereas the lowest chlorophyll content was obtained in control under irrigated and drought stressed treatments (Figure 4.3a). In this study it was observed that maize plants subjected to reduced soil moisture had lower chlorophyll content as compared to irrigated condition. As summarized in Figure 4.3b significant differences were observed for soil moisture content subjected to compost amendments in which the greatest amount was noted with SC and SUC treatments under both conditions while the least amount was obtained with SU and Control treatments. It is clear that adding compost to sandy soil has a significant effect in retaining the moisture under both conditions.



**Figure 4.3:** Effect of water stress and compost supplement on maize (a) leaf chlorophyll content and (b) soil moisture content at vegetative stage. Means followed by different letter are significantly different (P<0.05). Error bars indicate standard error of the means of plant chlorophyll (a) and soil moisture (b) contents, respectively. The following symbols mean: Control -Soil, SU - Soil +Urea fertilizer, SC- Soil +Compost and SUC - Soil+ Compost +Urea fertilizer).

4.6 Response of maize morphological characteristics as influenced by compost amendments and drought stress.

#### 4.6.1 Plant height at reproductive stage

The results for plant height are presented in Table 4.3. ANOVA results shows that there was significant water and fertilizer amendment treatments and their interactions. Plant height variations of maize at reproductive stage indicated that plant height differed due to compost application under drought stress and irrigated conditions. The highest measurements for plant height were obtained from maize crops planted in SC and SUC while the shortest heights were noted from the control and SU treatments under both irrigated and drought stressed conditions. Under irrigated conditions SC and SUC treatment led to higher plants than control and SU treatments. Similar results were obtained under drought stress conditions whereby SC and SUC treated plants were taller. The results show that amendment of soil with compost improved plant growth under irrigated and drought stressed conditions.

#### 4.6.2 Main stem diameter

The results for main stem diameter are presented in Table 4.3. ANOVA results shows there was significant water and fertilizer treatment. However, there was no significant interactions between water and fertilizer amendment for main stem diameter response. Under irrigated conditions SC and SUC treatment led to thicker plant stems than control and SU treatments. Similar results were obtained under drought stress conditions whereby SC and SUC treatments had thicker stems. The thickest stem diameter was observed in maize plants sown in SC (7.5) and SUC (8.4cm) under irrigated conditions and in SC (6.7cm) and SUC (6.8cm) under drought stressed conditions. These results were significantly different from control and SU treatment under both conditions.

The results show that amendment of soil with compost improved plant growth under irrigated and drought stressed conditions.

#### 4.6.3 Number of leaves

The effects of water and fertilizer amendments and their interactions were non-significant at P<0.05 on the number of leaves (Table 4.3). However, there is tendency for the number of leaves to increase in SC and SUC treatment under both irrigated and drought stress conditions.

#### **4.6.4** Leaf area (cm<sup>2</sup>)

Treatments variation on leaf area is described in Table 4.3, it showed the influence of irrigation deficit and compost application on maize leaf area. The effects of water and fertilizer amendments were significant while their interactions were not. The average leaf area of maize plants under irrigated and drought stress conditions was 442.9cm<sup>2</sup> and 310.2cm<sup>2</sup> respectively (Table 4.3). Under irrigated conditions, SC and SUC treatments increased leaf area and these were 478.8cm<sup>2</sup> and 562.5cm<sup>2</sup> respectively. Under drought stress conditions only SUC led to increased leaf area of 466.4cm<sup>2</sup> over the other treatments. Generally, the results show that amendment of soil with compost improved plant growth under irrigated and drought stressed conditions.

#### 4.6.5 Days to 50% Flowering

ANOVA for days to tasseling shows that water and fertilizer amendment treatments and their interactions were highly significant (Table 4.3). While data seems to be inconsistent among other treatments, it is apparent that SUC delayed tasseling by about 2 days compared to other treatments (controls and SU treatments), while SC delayed this response by a similar magnitude under both irrigated and drought stressed treatment. The results of ANOVA for days to silking shows water and fertilizer amendments and their interactions were significant.

The treatments SC and SUC delayed days to silking under both irrigated and drought stressed treatments compared to controls. These were 60 and 59 days under SC and SUC respectively compared to 56 days of controls.

#### 4.6.6 Days to 90% physiological maturity

ANOVA for days to physiological maturity shows that water and fertilizer amendment treatments and their interactions were highly significant (Table 4.3). Under irrigated conditions, SU, SC and SUC delayed maize to reach physiological maturity. Days to reach maturity were SU (119 days), SC (119 days) and SUC (120 days) compared to 118 days of controls. Under drought stress SU and SC delayed days to maturity with 121 days and 119 days respectively while SUC treatments reached maturity at the same time with Controls at 117 days each.

Treatment	Plant height(cm)	Stem diameter (cm)	Number of leaves	Leaf area (cm <sup>2</sup> )	Days to 50% tasseling	Days to 50% silking	Days to 90% maturity			
	Irrigated trial									
Control	93.0 <sup>b</sup>	4.3 <sup>c</sup>	9.8 <sup>b</sup>	269.9 <sup>b</sup>	57 <sup>b</sup>	56 <sup>c</sup>	118 <sup>c</sup>			
SU	95.0 <sup>b</sup>	5.7 <sup>b</sup>	11.3ª	460.3 <sup>ab</sup>	57 <sup>b</sup>	56 <sup>c</sup>	119 <sup>b</sup>			
SC	221.5ª	7.5 <sup>a</sup>	12.0ª	478.8 <sup>a</sup>	56 <sup>c</sup>	59 <sup>b</sup>	119 <sup>b</sup>			
SUC	222.0ª	8.4 <sup>a</sup>	12.8ª	562.5ª	59 <sup>a</sup>	60 <sup>a</sup>	120 <sup>a</sup>			
Mean	145.9	10.1	11.5	442.9	57	58	119			
LSD <sub>0.05</sub>	18.64	0.98	2.26	195.23	0	0	0			
			Drought	stressed						
Control	72.0 <sup>b</sup>	3.9 <sup>b</sup>	9.5 <sup>bc</sup>	214.4 <sup>b</sup>	56 °	58 <sup>b</sup>	117°			
SU	98.5 <sup>b</sup>	4.4 <sup>b</sup>	9.0°	272.9 <sup>b</sup>	57 <sup>b</sup>	57°	121 <sup>a</sup>			
SC	166.5 <sup>a</sup>	6.7 <sup>a</sup>	11.3ª	287.0 <sup>b</sup>	55 <sup>d</sup>	57°	119 <sup>b</sup>			
SUC	152.8ª	6.8 <sup>a</sup>	10.8 <sup>b</sup>	466.4 <sup>a</sup>	59 <sup>a</sup>	59 <sup>a</sup>	117°			
Mean	109.9	5.5	10.2	310.2	57	58	118			
LSD <sub>0.05</sub>	44.8	1.09	1.71	169.31	0	0	0			
ANOVA										
Water	$0.0001^{****}$	0.0001****	0.097 <sup>ns</sup>	$0.001^{***}$	$0.0001^{****}$	0.0001****	$0.000^{1****}$			
Fertilizer F * W	$0.0001^{****}$ $0.01^{*}$	0.0001 <sup>****</sup> 0.280 <sup>ns</sup>	0.146 <sup>ns</sup> 0.749 <sup>ns</sup>	0.001 <sup>***</sup> 0.503 <sup>ns</sup>	$0.0001^{****}$ $0.0001^{****}$	$0.0001^{****}$ $0.0001^{****}$	$0.0001^{****}$ $0.0001^{****}$			

 Table 4.3: Response of maize morphological characteristics as influenced by compost amendments and drought

<sup>a</sup>Means followed by the same letters in the same column are not significant at P<0.05,; \*,\*\*,\*\*\*,\*\*\*\* denote significant difference at

P<0.05, P<0.01, P<0.001 and respectively; and ns= not significant. Control (Soil), SU (Soil+ Urea fertilizer), SC (Soil

+Compost), SUC (Soil+ Compost +Urea fertilizer).

# 4.7 Impact of drought stress and compost amendments on maize yield contributing components

#### 4.7.1 Cob dry weight (g)

ANOVA for cob dry weight shows that water and fertilizer amendment treatments and their interactions were highly significant (Table 4.4). Under irrigated conditions, the SC and SUC treatments increased cob dry weight. The weights were SC (230.3g) and SUC (140.5g). Under drought stress, the SU, SC and SUC increased cob weights by compared to controls. The weights were SU (91.8g), SC (86.7g) and SUC (127.6g), while that of the controls were 25.7g. The results show that amendment of the soil with compost (SC and SUC) increased maize cob weight under irrigated and drought stressed conditions.

#### 4.7.2 Number of grains per cob

ANOVA for number of grains per cob shows that water and fertilizer amendment treatments and their interactions were significant (Table 4.4). Under irrigated conditions, SU SC and SUC increased number of grains/cob compared to controls. These were 43 (SU), 188 (SC) and 127 (SUC) grains per cob while controls did produce grain at the time of harvest. Under drought stress conditions, SU, SC and SUC treatments increased the number of grains/cob compared to the controls. These were 30 grains/cob (SU) grains/cob, 92 (SC) grains/cob, 82 grains/cob (SUC) while those for controls were 8 grains per cob. The results shows that compost amendment increased the number of grains per cob in maize under both irrigated and drought stress conditions.

#### 4.7.3 100 seed weight (g)

ANOVA for 100 seed weight shows that there was no significant water effect, while fertilizer amendment treatment was significant (Table 4.4). The interaction between water and fertilizer

amendments was however insignificant. Under irrigated conditions SU, SC and SUC increased the 100 seed weight over the control. There were 8.3g (SU), 55.3g (SC) and 68.3g (SUC) compared to controls where there was no grain. Under drought stress conditions, SC and SUC increased 100 seed weight compared to SU and controls where there were no grain production in these treatments. The results show that soil amendment with compost (SC and SUC) increased 100 seed weight in maize.

Treatments	Cob weight(g)	Grains/ cob	100 seed weight(g)				
Irrigated trial							
Control	13.9 <sup>c</sup>	$0^{c}$	$0^{\mathrm{b}}$				
SU	67.2 <sup>c</sup>	43.3 <sup>bc</sup>	8.3 <sup>b</sup>				
SC	230.3 <sup>a</sup>	188 <sup>a</sup>	55.3ª				
SUC	140.5 <sup>b</sup>	127 <sup>ab</sup>	68.3 <sup>a</sup>				
Mean	112.9	89.6	33.1				
LSD0.05	72.8	102.93	30.59				
	Drought	stressed					
Control	25.7 <sup>b</sup>	8.8 <sup>c</sup>	$0.0^{b}$				
SU	91.8 <sup>a</sup>	30.0 <sup>b</sup>	$0.0^{b}$				
SC	86.7 <sup>ab</sup>	92.3 <sup>a</sup>	41.5 <sup>a</sup>				
SUC	127.6 <sup>a</sup>	82.3 <sup>a</sup>	50.3 <sup>a</sup>				
Mean	83.1	53.4	23.1				
LSD0.05	63.52	92.44	38.39				
	ANO	VA					
Water	$0.047^{*}$	0.09 <sup>ns</sup>	0.209 <sup>ns</sup>				
Fertilizer	0.0001****	0.001***	$0.0001^{****}$				
F * W	0.001 <sup>***</sup> 0.337 <sup>ns</sup>		0.856 <sup>ns</sup>				

Table 4.4: Impact of drought stress and compost amendments on maize yield components.

<sup>a</sup>Means followed by the same letters in the same column are not significant at P< 0.05,; \*,\*\*, \*\*\*, \*\*\*\*, \*\*\*\* denote significant differences at, and P< 0.05, P < 0.01, P< 0.001 and P< 0.0001 respectively; and ns- not significant difference at P<0.05. LSD: Least significant difference. Control (Soil), SU (Soil +Urea fertilizer), SC (Soil +Compost), SUC (Soil +Compost +Urea fertilizer).

4.8: Accumulation of maize biomass at vegetative and physiological maturity stages as influenced by compost amendments and drought stress.

#### **4.8.1** Biomass at vegetative stage

There was no significant interactions between drought and fertilizer amendments (Table 4.5). Fertilizer amendment treatments resulted in significant difference at P<0.001. Within the treatments under irrigated condition, the difference were significant with maximum dry matter of 18.08g yielded from maize plants sown in combination of SUC as compared to other treatments. Under drought stress treatment SC and SUC treatments had significantly higher vegetative biomass of 12.94 and 10.84g/plant respectively (Table 4.5). The results shows compost amendments (SC and SUC) compared to Control and SU increased biomass production in maize under both irrigated and drought stressed conditions.

#### 4.8.2 Shoot biomass at maturity

There was no significant interaction between drought and fertilizer amendment treatments (Table 4.5), while fertilizer amendment treatments indicated significant difference at P<0.0001. Under irrigated conditions, SC and SUC produced significantly higher biomass of 116.80 and 179.33g/plant respectively. Similar results were obtained under drought stress where SC produced 139.99g/plant and SUC resulted in 177.80 g/plant. In general, the highest figures of shoot biomass weight were recorded with combination of SUC under irrigated and water stress conditions while the lowest figures were noted in SU and Control treatments under both conditions (Table 4.5).

#### **4.8.3 Root biomass at maturity**

Concerning the effect of drought and the interaction of water \* fertilizer amendment data presented in Table 4.5 shows that there was no significant difference on root biomass at maturity stage though fertilizer addition induced significant difference at P<0.001.

Total root biomass varied among all the treatments except in SC and SUC treatment increased root biomass under irrigated and drought stressed conditions. It is indicated from this Table 4.5 that SC and combination SUC under both conditions yielded highest root dry weights (121.76 and 50.41g/plant) and (82.46 and 57.74g/plant) respectively, statistically greater than weights of root biomass in SU and control maize plants. Therefore, compost amendment (SC and SUC) treatments increased the root biomass compared to SU and the control.

#### 4.8.4 Total biomass at physiological maturity

The results shows that drought and its interaction with fertilizer amendments were not significant on biomass production (Table 4.5). Fertilizer amendment treatments were highly significantly different at P<0.0001 (Table 4.5). Under irrigated conditions, SC and SUC produced 238.64 and 261.73g/plant respectively, while SU and Control produced 33.8 and 57.69g/plant respectively. Under drought stress, SC and SUC produced 190.40g/plant and 235.54g/plant respectively, while SU and Control treatment produced 23.50 and 65.35g/plant respectively. Therefore, incorporation of compost in the soil (SC and SUC) resulted in higher biomass under both irrigated and drought stress conditions compared to SU and control treatments.

Treatment	Biomass at vegetative stage (g)	Shoot biomass (g)	Root biomass (g )	Biomass at maturity (g)
		Irrigated trial	-	-
Control	2.91°	26.70°	7.10 <sup>c</sup>	33.80 <sup>b</sup>
SU	6.68 <sup>bc</sup>	43.53°	14.16 <sup>bc</sup>	57.69 <sup>b</sup>
SC SUC	$16.48^{ab}$ 18.08 <sup>a</sup>	116.8 <sup>b</sup> 179.33ª	$121.76^{a}$ $82.46^{ab}$	238.64ª 261.73ª
Mean	11	91.59	56.4	238.5
LSD0.05	11.22	53.69	71.02	73.41
		Drought Stressed		
Control	4.63 <sup>b</sup>	15.51 <sup>b</sup>	7.79 <sup>b</sup>	23.50 <sup>b</sup>
SU	4.24 <sup>b</sup>	46.25 <sup>b</sup>	19.10 <sup>b</sup>	65.35 <sup>b</sup>
SC	12.94 <sup>a</sup>	139.99 <sup>a</sup>	50.41 <sup>a</sup>	$190.40^{a}$
SUC	10.84 <sup>ab</sup>	177.80 <sup>a</sup>	57.74 <sup>a</sup>	235.54 <sup>a</sup>
Mean	8.9	94.9	33.8	127.5
LSD <sub>0.05</sub>	7.19	60.92	28.95	74.2
		ANOVA		
Water	0.223 <sup>ns</sup>	0.821 <sup>ns</sup>	0.09 <sup>ns</sup>	0.431 <sup>ns</sup>
Fertilizer	$0.001^{***}$	$0.0001^{****}$	$0.001^{***}$	0.0001****
F * W	0.590ns	0.857 <sup>ns</sup>	$0.170^{ns}$	0.871 <sup>ns</sup>

Table 4.5: Biomass production of maize as influenced by compost application and drought

<sup>a</sup>Means followed by the same letters in the same column are not significant at P< 0.05,; \*,\*\*, \*\*\*, \*\*\*\* denote significant differences at P< 0.05, P < 0.01, P< 0.001 and P< 0.0001 respectively; and ns- no significant difference at P<0.05. LSD is least significant difference. Control -Soil without amendments, SU - Soil +Urea fertilizer, SC - Soil +Compost, SUC -Soil +Compost +Urea fertilizer.


(c)

**Figure 4.4:** An overview of maize performance as influenced by water regime and soil compost amendments: (a) Maize growth under irrigated and drought stressed conditions, (b) Root biomass (c) grain yield of maize under soil amended using compost and urea fertilizer.

#### 4.9 Nutrient concentration in maize at harvesting stage – Green house experiment

Data in Table 4.6 shows maize nutrient content at harvest as influenced by compost amendments under irrigated and drought stressed conditions. There was no significant interactions between drought stress \* fertilizer amendments on N concentration. On the other hand, the effect of fertilizer amendments and drought stress on N content reflects significant different at P<0.0001. The greatest N concentration was recorded in SU treatment under both irrigated and drought stressed conditions.

Concerning phosphorus the results shows that there was no significant interaction between drought stress \* fertilizer amendment on P content whereas fertilizer amendment had a significant effect (P<0.0001). Drought stress also had a significant effect on maize P content at P<0.05. Among the treatments control recorded the greatest P concentration as compared to the rest of the treatments under both irrigated and drought stressed conditions. The results on potassium (Table 4.6) demonstrated that water stress and fertilizer amendments and its interactions were all significant at P< 0.0001. The highest K content was obtained from compost amended soil (SC and SUC).

parameters	Ν		Р		Κ		Ca		Mg		Na	
	<		% plant nutrient content				$\rightarrow$					
Treatments	Ι	D	Ι	D	Ι	D	Ι	D	Ι	D	Ι	D
Control SU SC	$0.0923^{b}$ $0.1809^{a}$ $0.0971^{b}$	$0.1025^{d}$ $0.1938^{a}$ $0.1250^{c}$	0.00135 <sup>a</sup> 0.00020 <sup>c</sup> 0.00043 b	$\begin{array}{c} 0.0021^{a} \\ 0.0002^{b} \\ 0.0005^{b} \end{array}$	$\begin{array}{c} 1.0938^{b} \\ 0.8491^{b} \\ 2.0155^{ab} \end{array}$	$0.9611^{b}$ $0.8854^{b}$ $6.4081^{a}$	$\begin{array}{c} 0.1427^{c} \\ 0.2494^{bc} \\ 0.5181^{a} \end{array}$	$\begin{array}{c} 0.1416^{c} \\ 0.2187^{bc} \\ 0.3423^{a} \end{array}$	$0.1104^{b}$ $0.1885^{a}$ $0.1863^{a}$	$0.1479^{a}$ $0.1506^{a}$ $0.1567^{a}$	$\begin{array}{c} 0.0343^{ab} \\ 0.0408^{a} \\ 0.0305^{ab} \end{array}$	$\begin{array}{c} 0.0226^{b} \\ 0.0221^{b} \\ 0.0215^{b} \end{array}$
SUC LSD(0.05) ANOVA	0.0980 <sup>b</sup> 0.05	0.1545 <sup>b</sup> 0.02	0.00024 <sup>c</sup> 0.21	0.0008 <sup>b</sup> 0.0007	2.9192 <sup>a</sup> 0.05	6.8012ª 0.69	0.4269 <sup>ab</sup> 1.247	0.2823 <sup>ab</sup> 0.11	0.1689ª 0.01	$0.1258^{a}$ 0.05	0.0266 <sup>b</sup> 0.0002	0.0321 <sup>a</sup> 0.0064
Fertilizer Water F*W	0.0001**** 0.0001**** 0.3762 <sup>ns</sup>		0.0001 <sup>****</sup> 0.01* 0.0860 <sup>ns</sup>		0.0001**** 0.0001**** 0.0001****		0.0001**** 0.0283* 0.301 <sup>ns</sup>		$0.028^{*}$ $0.095^{ns}$ $0.043^{*}$		$0.0441^{*}$ $0.001^{***}$ $0.010^{*}$	

Table 4.6: Nutrient concentration in maize at harvesting stage – Green house experiment

<sup>a</sup>Means followed by the same letters in the same column are not significant at P<0.05; \*,\*\*,\*\*\*,\*\*\*\* denote significant difference at P<0.05, P<0.01, P<0.001 and P<0.0001 respectively; and ns=not significant. I and D represent Irrigated and drought stress conditions respectively. LSD: Least significant difference. Control (Soil), SU (Soil + Urea fertilizer), SC (Soil+ Compost), SUC (Soil + Compost + Urea fertilizer).

## 4.10 a). Nitrogen and phosphorus use efficiency of maize at maturity stage.

#### 4.10 b). Nitrogen use efficiency in maize

Drought stress and fertilizer amendment interactions were not significantly different (p>0.05) on nitrogen use efficiency (NUE) at maturity stage whereas fertilizer amendment treatments had significant effect (P<0.0001) on NUE in maize at physiological maturity (Table 4.7). The highest NUE was detected on maize plants grown in SUC (24.66g/kg and 14.17g/kg) as compared to other treatments under both conditions. The lowest NUE was noted in SU (2.25g/kg), SC (9.22g/kg) and control treatments (2.46g/kg) under irrigated conditions.

# **4.10 c).** Phosphorus use efficiency of maize under different water regime and compost amended soil.

Data for phosphorus use efficiency (PUE) of maize is given in Table 4.7. PUE was also influenced by compost amendment and irrigation deficit conditions. The interactive effects of drought stress and fertilizer amendment were highly significant at P<0.001. The effects of drought stress and fertilizer amendment were also highly significant at P< 0.0001. The highest PUE was noted in SUC treatment (210.35kg/kg) and SC (141.89 kg/kg) under irrigated condition and the lowest was recorded in control and SU (34.11 kg/kg and 17.08 kg/kg) treatments respectively under irrigated condition (Table 4.7). A similar behavior was observed in maize plants under drought stressed condition. Therefore, amendment of soil with compost (SC and SUC) treatment increased PUE under both irrigated and drought stressed conditions.

	Nitrogen use effic	iency (NUE) kg/kg)	Phosphorus use eff	iciency (PUE		
TREATMENTS	Ι	D	Ι	D		
Control	2.46 <sup>b</sup>	2.79 <sup>b</sup>	17.08 <sup>b</sup>	2.40 <sup>b</sup>		
SU	2.25 <sup>b</sup>	4.31 <sup>b</sup>	34.11 <sup>b</sup>	6.65 <sup>b</sup>		
SC	9.22 <sup>b</sup>	7.88 <sup>ab</sup>	141.89 <sup>a</sup>	20.37 <sup>a</sup>		
SUC	24.66 <sup>a</sup>	14.17 <sup>a</sup>	210.35 <sup>a</sup>	24.71 <sup>a</sup>		
MEAN	8.74	7.29	100.86	13.53		
LSD(0.05)	10.68	6.69	81.9	10.42		
		ANOVA				
Fertilizer	0.0	001****	$0.0001^{****}$			
Water	0.	265 <sup>ns</sup>	0.0001****			
Fertilizer* water	0.	169 <sup>ns</sup>	0.001***			

Table 4.7: Nitrogen Use Efficiency and Phosphorus Use Efficiency of maize at maturity

<sup>a</sup>Means followed by the same letters in the same column are not significant at P<0.05,; \*,\*\*,\*\*\*,\*\*\*\* denote significant difference at P<0.05, P<0.01, P<0.001 and P<0.0001 respectively; and ns=not significant. I and D represent Irrigated and drought stress conditions respectively. LSD: Least significant difference.

# **4.11:** Effect of compost amendments and drought stress on photosynthetic parameters of maize.

Fertilizer amendments application and its interaction with drought stress significantly (P<0.0001) affected the rate of photosynthesis in maize (Table 4.8) while water treatment had no significant effect on the rate of photosynthesis in maize. A non-significant difference was observed among the treatments with regard to photosynthetic rate of maize under irrigated condition. Under drought stressed condition control exhibited the lower photosynthetic rates compared to other treatments. Amendment of soil with compost (SC and SUC) led to increased photosynthesis in maize under drought stress, but not under irrigated conditions.

With regard to transpiration, the interactions of fertilizer amendment and drought stress had significant effect at P<0.05 on transpiration rates (Table 4.8). Under both conditions maize plants sown in Control and SU treatments exhibited significantly higher rates of transpiration than in SC and combination of SUC (Table 4.8). Transpiration rate was significantly (P<0.0001) affected by fertilizer amendment while drought stress also significantly (P<0.001) affected the rate of transpiration in maize plants. Amendment of soil with compost (SC and SUC) significantly reduced transpiration under both irrigated and drought stressed conditions.

The results also show that fertilizer amendment and drought stress had no significant differences in the internal  $CO_2$  of maize plants although their interactions were significant (P<0.05) on internal  $CO_2$  concentration. Amendment of soil with compost (SC and SUC) led to reduction in internal  $CO_2$  concentration under drought stress condition, but not under irrigated conditions. As Table 4.8 indicates, drought stress, fertilizer amendments and interaction of fertilizer amendment and drought stress had no significant effect on stomatal conductance. There was no significant differences in stomatal conductance among all the treatments under both conditions.

The results on WUE shows significant difference between drought stress treatments and interaction of water and fertilizer amendments at (P< 0.05). WUE was significantly (P<0.001) affected by fertilizer amendments. As indicated in Table 4.8, there was no significant difference among irrigated treatments with regard to WUE. Maize plants under water deficit condition had similar trend of WUE compared to their corresponding irrigated treatments except that control exhibited the lowest value (1.99 g/g<sup>-</sup> plant). Therefore, amendment of soil with compost (SC and SUC) led to increased WUE and these were not significantly different from soil amended with urea fertilizer (SU).

Photosynthesis (A) µmol CO <sub>2</sub> m <sup>-2</sup> s <sup>-1</sup>		Transpiration (E) mmol H <sub>2</sub> O m <sup>-2</sup> s <sup>-1</sup>		Internal CO <sub>2</sub> (Ci) µmol m <sup>-2</sup> s <sup>-1</sup>		Stomatal conductance (g) µmol m <sup>-2</sup> s <sup>-1</sup>		Water Use efficiency (WUE) (µmol	
•								CO <sub>2</sub> /mmol H <sub>2</sub> O)	
Ι	D	Ι	D	Ι	D	Ι	D	Ι	D
$25.74^{a} \\ 26.14^{a} \\ 26.98^{a} \\ 23.47^{a} \\ 5.93$	14.84 <sup>b</sup> 29.13 <sup>a</sup> 27.25 <sup>a</sup> 28.96 <sup>a</sup> 4.54	9.60 <sup>a</sup> 9.00 <sup>a</sup> 1.24 <sup>b</sup> 1.41 <sup>b</sup> 1.43	7.43 <sup>a</sup> 7.29 <sup>a</sup> 1.35 <sup>b</sup> 1.25 <sup>b</sup> 0.55	339.58 <sup>a</sup> 354.40 <sup>a</sup> 333.29 <sup>a</sup> 342.84 <sup>a</sup> 29.98	361.49 <sup>a</sup> 328.99 <sup>bc</sup> 339.40 <sup>b</sup> 327.38 <sup>c</sup> 11.81	1.28 <sup>a</sup> 1.27 <sup>a</sup> 1.24 <sup>a</sup> 1.41 <sup>a</sup> 0.49	$     1.10^{a} \\     1.13^{a} \\     1.35^{a} \\     1.25^{a} \\     0.27 $	2.70 <sup>a</sup> 3.01 <sup>a</sup> 2.98 <sup>a</sup> 2.62 <sup>a</sup> 0.72	$   \begin{array}{r}     1.99^{b} \\     4.00^{a} \\     3.68^{a} \\     3.63^{a} \\     0.78   \end{array} $
				ANG	)VA				
er 0.0001****		$0.0001^{****}$		0.125 <sup>ns</sup>		0.321 <sup>ns</sup>		0.001***	
0.632 <sup>ns</sup>		0.001***		0.510 <sup>ns</sup>		0.658 <sup>ns</sup>		$0.01^*$	
0.0001****		0.02*		0.01*		0.654 <sup>ns</sup>		$0.01^{*}$	
	Photosyn           μmol CO           I           25.74 <sup>a</sup> 26.14 <sup>a</sup> 26.98 <sup>a</sup> 23.47 <sup>a</sup> 5.93           r         0.0001           0.632 <sup>ns</sup> 0.0001	I       D         25.74 <sup>a</sup> 14.84 <sup>b</sup> 26.14 <sup>a</sup> 29.13 <sup>a</sup> 26.98 <sup>a</sup> 27.25 <sup>a</sup> 23.47 <sup>a</sup> 28.96 <sup>a</sup> 5.93       4.54         r       0.0001****         0.632 <sup>ns</sup> 0.0001*****	Image: Photosynthesis (A) $\mu$ mol CO <sub>2</sub> m <sup>-2</sup> s <sup>-1</sup> Transpiration mmol H <sub>2</sub> O         I       D       I         25.74 <sup>a</sup> 14.84 <sup>b</sup> 9.60 <sup>a</sup> 26.98 <sup>a</sup> 27.25 <sup>a</sup> 1.24 <sup>b</sup> 23.47 <sup>a</sup> 28.96 <sup>a</sup> 1.41 <sup>b</sup> 5.93       4.54       1.43         r       0.0001*****       0.0001*****         0.632 <sup>ns</sup> 0.001****       0.02*	Photosynthesis (A) µmol CO2 m² s¹Transpiration (E) mmol H2O m² s¹IDID25.74a14.84b9.60a7.43a26.14a29.13a9.00a7.29a26.98a27.25a1.24b1.35b23.47a28.96a1.41b1.25b5.934.541.430.55r0.0001****0.0001****0.632ns0.001****0.002*	Photosynthesis (A) µmol CO2 m <sup>-2</sup> s <sup>-1</sup> Transpiration (E) mmol H2O m <sup>-2</sup> s <sup>-1</sup> Internal C m <sup>-2</sup> s <sup>-1</sup> IDIDI25.74a14.84b9.60a7.43a339.58a26.14a29.13a9.00a7.29a354.40a26.98a27.25a1.24b1.35b333.29a23.47a28.96a1.41b1.25b342.84a5.934.541.430.5529.98r0.0001****0.0001****0.125 <sup>ns</sup> 0.632 <sup>ns</sup> 0.001****0.001*	Photosynthesis (A) µmol CO2 m² s¹Transpiration (E) mmol H2O m² s¹Internal CO2 (Ci) µmol m² s¹IDID25.74a14.84b9.60a7.43a339.58a361.49a26.14a29.13a9.00a7.29a354.40a328.99bc26.98a27.25a1.24b1.35b333.29a339.40b23.47a28.96a1.41b1.25b342.84a327.38c5.934.541.430.5529.9811.81r0.0001****0.0001****0.510ns0.510ns0.0001****0.02*0.01*0.01*	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\begin{array}{ c c c c c c c c c c c c c c c c c c c$	Photosynthesis (A) µmol CO2 m² s¹Transpiration (E) mmol H2O m² s¹Internal CO2 (Ci) µmol m² s¹Stomatal conductance (g) µmol m² s¹Water Us (WUE) (µ CO2/mmolIDIDIDIDI25.74a14.84b9.60a7.43a339.58a361.49a1.28a1.10a2.70a26.14a29.13a9.00a7.29a354.40a328.99bc1.27a1.13a3.01a26.98a27.25a1.24b1.35b333.29a339.40b1.24a1.35a2.98a23.47a28.96a1.41b1.25b342.84a327.38c1.41a1.25a2.62a5.934.541.430.5529.9811.810.490.270.72ANOVA0.632ns0.001****0.510ns0.658ns0.658ns0.0001****0.02*0.01*0.654ns0.654ns

4.8: Photosynthesis, Transpiration, internal CO<sub>2</sub>, Stomatal conductance and WUE of maize at flowering stage.

<sup>a</sup>Means followed by the same letters in the same column are not significant at P<0.05,; \*,\*\*,\*\*\*,\*\*\*\* denote significant difference at P<0.05, P<0.01, P<0.001 and P<0.0001 respectively; and ns=not significant. I and D represent Irrigated and drought stress conditions respectively. LSD: Least significant difference.

# CHAPTER FIVE DISCUSSION

#### 5.1 Characteristics of soils at the study site

Soils at the experimental study site was classified as orthic-luvisol and this soil types are coarse sandy loam. Sandy soils generally have low CEC hence less ability to hold and retain nutrients. These soils are moderately deep to very deep with low nutrient due to very severe leaching and low organic matter content (Simon and Czacko, 2014). The results from the initial soil analysis showed low soil fertility levels of the experimental site. This was indicated by the low levels of basic cations (Ca, K, Mg, and Na). Water stress and low soil nutrient content adversely affects plant growth which ultimately decreases the final yield. Hence soils amended with compost (SC and SUC) alleviate the negative effects of drought stress and low soil fertility on plants. This shows substantial effect of compost amendment on utmost parameters investigated during the study period.

## 5.2 Effect of compost amendment on soil chemical and physical properties.

The results of this experiments show a positive effect of compost application on soil pH, EC, CEC, total C, total N, available P and exchangeable cations (Ca, Mg and K). The significant increase of soil pH in the study work was predominantly due to the initial high pH values of the compost added. Addition of compost is essential to provide necessary nutrients for crops and improving soil physico-chemical properties (Meena *et al.*, 2015). It has been observed that the addition of soil amendments (organic and inorganic) has great potential for increasing soil pH (3.2–7), reducing the solubility of trace metals by more than 80%, and stabilizing the soil (Pardo *et al.*, 2017). The results are in line with the work of Liu *et al.* (2019) on remediation effectiveness of

vermicompost for a potentially toxic metal contaminated tropical acidic soil, where vermicompost amendment increased soil pH by 0.7 to 1.5 units. Mostly the amendment of acidic soil with compost increase soil pH values to levels that are more suitable for most crops growth. A study by Brar *et al.* (2015) showed that integrated use of inorganic fertilizer along with organic fertilizer (100% NPK + FYM) improved soil pH which resulted in higher maize and wheat yields. These findings confirmed the effectiveness of compost in enhancing these properties Ch'ng *et al.* (2015) and Sanusi *et al.* (2018), in which the treatments with organic amendments significantly increased soil pH. According to Agegnehu *et al.* (2016) composts have a liming effect because of their richness in alkaline or base cations such as Ca, Mg, Na, and K which were liberated from organic matter through mineralization. The pH change is a key factor for regulating the solubility and availability of nutrients in the soil.

Electrical conductivity (EC) is a soil parameter that indirectly determines the total concentration of soluble salts and also a direct salinity measurement. The results in this experiment revealed that the overall EC was lower in SU and control soil treatments under both conditions as compared to compost treated soils (SC and SUC). The EC values of soil treated with compost ranged from 1.39 to 1.78 (ds/m) in both conditions compared with control. The increase in the soil EC values can be explained by the inputs of nutrients and salts contained in the compost. Integration of composts into soil increases the salt content as well as soil EC, especially if high rates of compost are applied (Angelova *et al.*, 2013).

The results of the study indicated elevation of CEC with compost treated soils (SC and SUC) as compared to non- compost soils, this could also be attributed to the fact that compost have a higher cation exchange capacity and that can therefore increase CEC. Liu *et al.* (2012) also indicated that compost amendment results in an increase of CEC due to input from stabilized OM being rich

in functional groups such as carboxylic and phenolic acid groups being released into the soil exchange sites. Agegnehu *et al.*, (2016b) suggested that enhanced CEC increased soil fertility by increasing nutrient availability through nutrient retention in soil instead of leaching through soil profile out of the rooting zone.

The results also exhibited a higher total carbon content in soils treated with compost (SC and SUC) than the control and urea treatments. The increase of soil total carbon with addition of compost amendments may be associated by high organic matter content of compost. The results concur with the findings of Bouajila and Sanaa (2011) who reported that the application of compost from manure and household waste resulted in a significant increase in organic carbon with the compost treatment being the most efficient. Their results showed that the application of household waste compost and manure improved the organic carbon. Trupiano *et al.* (2017) also found that application of compost and biochar, alone or in combination, increased soil TOC content than that in the unamended soils, which is indicative that biochar and/or compost applications to soils can enhance C accumulation and sequestration.

This study revealed that compost amendments showed a slight effect on soil available P, as compared to control treatments, which may be partially attributed to the fact that compost released phosphorus and organic acids from decomposition of organic matter. A study by Mensah and Frimpong (2018) found out that the application of compost significantly increased the soil available P. The structural improvements made to soil by increasing the quantity of SOM can improve plant P availability by allowing for greater root access to soil P stores (Schröder *et al.*, 2011). Similarly, Mao *et al.* (2008) and Olowolafe (2008) also observed a higher phosphorus

content of the soil where cattle manure or municipal waste was used for a period of 5 years on maize compared to inorganic fertilized soil.

The results for exchangeable cations, such as potassium (K), calcium (Ca) and magnesium (Mg), significantly increased with the compost amendment (SC and SUC) treatments. This implies that compost was very beneficial for plants as a source of exchangeable cations. The findings are in line with Adugna (2016) who expressed that the mineralization of compost would release many nutrients into the soil so that the nutrients would be greatly increased.

# 5.3 The effect of soil amendment under irrigated and drought stress conditions on morphophysiological responses of maize.

#### 5.3.1 Plant height

The application of compost in this study had a significantly positive effect on the growth and yield attributes of maize, when applied together with urea fertilizer. The data obtained from the study revealed that plant height was significantly affected by water stress and compost application. Although there was variation between treatments, increase in plant height after planting indicated that water availability and uptake of nutrients significantly increased plant height under irrigated as well as non-irrigated experiments. Xu and Mou (2016) reported that increased growth in-terms of length and biomass accumulation due to compost treatment may be ascribed to increased photosynthesis.

Regarding compost application, the results revealed that the highest measurements were obtained from compost soil and/or in combination of compost soil + urea while the shortest heights were noted from non-compost treatments. This study agrees with the work of Mahmud *et al.* (2016) who reported that the combined application of compost and chemical fertilizer on rice plants had a great

influence on plant height which might have been due to the presence of major nutrients from the organic fertilizer combined with the instantaneous readily soluble nutrients from the inorganic fertilizers. Manish et al. (2017) also reported that the tallest plant height was observed on the plots treated by vermicompost and cattle manure, whereas the shortest was in the control without this treatments. The application of compost increased plant height in both conditions meaning that compost has the ability to mitigate drought effects on plant height. A similar result was also reported by Mahmud et al. (2016), stating that incorporation of vermicompost to the soil influenced plant growth, especially plant height compared to control treatment (non-fertilizer application). Plant height plays a major role in the final yield of maize crops. The increase in plant height with compost addition can be attributed to that compost promotes plant growth, increases the number and length of the internodes which results in progressive increase in plant height. The findings are also in agreement with the work of Aziz et al. (2010) and Ogbonna et al. (2012), who both reported a significantly taller plants and larger leaves with compost application. According to Okoroafor *et al.* (2013) the application of organic manure highly increased plant height, number of maize leaves, stem girth, number of cob and weight of fresh maize at harvest. Similar results were reported by Coulibaly et al. (2019) who found that the tallest growth parameters of maize were obtained with compost made from pig's manure. Generally, it was observed that treatments that received both compost and urea fertilizer produced taller plants compared to plants in control treatments.

Concerning drought stress effect, this study showed that compost amendment into the soil increased water availability to the plants under stressed condition and accelerated recovery of plants after drought stress. It was observed that low water availability suppressed maize plant

height during the experimental study. The results depicted a varying range of plant height under well-watered and stressed conditions. Well-watered maize plants were taller, yet again taller plants were also observed under drought stressed condition on plants treated with SC and SUC, and the results can be associated to the fact that the incorporation of compost added organic sources into the soil which improved soil water holding capacity. This agrees with the work by Logsdon et al. (2017) who examined the water content of lawns with compost incorporation. Similar finding was stated by Mahmood et al. (2017), who reported that organic manure application decreased soil bulk density and enhanced soil porosity and water holding capacity. Compost is known to have a high water holding capacity and can provide water to plants over time (Crogger, 2005). Drought stressed plants had significantly reduced plant height as compared to well-watered treatments. So it is clear that plant height is negatively affected by drought stress. According to Sikuku et al. (2012) the depression of plant height could also have resulted from a reduction in plant photosynthetic efficiency. Studies have shown that the decrease in transpiration rate under drought stress significantly decreases plant height and dry matter content (Ramegowda et al., 2014). Drought stress induced reduction in plant height was also observed by Khan et al. (2014) in soybean.

## 5.3.2 Stem diameter

Stem diameter of maize is an important measure, which determines its strength and ability to resist to lodging. It was observed that under reduced water supply the stem becomes thinner and furthermore the increase in stem diameter due to compost amendments can be elucidated by the fact that compost promoted plant growth. The results revealed that main stem diameter of maize planted in SC and combination of SUC tend to have thicker stems as compared to those planted in control and SU treatments under both treatments. Control and SU plants resulted in the lowest stem diameter, which can be described as a direct effect of nutrient availability. Results of this trial are in line with several earlier work of (Ravi *et al.*, 2012; Lone *et al.*, 2013). The increase in stem diameter indicated improved growth of maize plants after the addition of composts to the soil. Organic manures have been said to improve soil fertility by activating soil microbial biomass, which in turn leads to development in crops (Ayuso *et al.*, 1996) and this may have been responsible for the observed increase in stem diameter resulting from nutrient application. Comparable results were obtained by Haouvanga *et al.* (2017) who stated that adding compost on soil significantly increased the number of leaves and the stem diameter of Moringa oleifera. Previous studies also stated that nitrogen significantly increased cereal leaf area. Positive effects of compost may be attributed to providing available source of carbon and nitrogen for soil microorganisms which enhances soil structure, reduces soil erosion, lowers the temperature at the soil surface and helps increase water holding capacity of the soil.

#### **5.3.3 Number of leaves**

In this study the number of leaves per plant was significantly affected by water treatments which caused a reduction in leaf number under drought stressed condition. Again in the present experiment averages of 12 and 10 numbers of leaves were formed in maize under irrigated and drought conditions respectively. A study by Lamm *et al.* (2005) found that deficit irrigation reduced total number of leaves. With regard to amendment of compost (SC and SUC) the number of leaves per plant improved significantly when compared with the control treatments. The results of the experiment agreed with the work of Adamu *et al.* (2015), who also reported that application of full farm yard manure, N and P produced the highest leaves per plant, while the control had the

lowest leaves. However, Sagar and Sharma. (2015) reported that application of farm yard manure and nitrogen did not significantly influence the number of leaves per plant. The results of the study (Manyuchi *et al.*, 2013) reported that vermicompost application increases the number of leaves of maize plants. The number of leaves on a plant determines the photosynthetic activity of a plant which influences growth and yield of the crop.

## 5.3.4 Leaf area (cm<sup>2</sup>)

Combination of compost with urea fertilizer in the trial showed great potential to increase the leaf area of individual plant compared to that of control under both conditions. This explained that different nutrient release from urea and compost improved the chemical and physical properties of the soil thereby increasing the growth and yield parameters of maize. Reduction in leaf area is morphological parameters for measuring drought stress experienced by the plant (Ku *et al.* 2013). Thus decline of leaf area significantly affects maize productivity, due to its low plasticity. Moreover, reduction in leaf area is a mechanism used by plants to avoid higher rate of transpiration and reduce surfaces for radiation due to water deficit (Hayatu, 2014). In general drought stress significantly reduced the total leaf area. Khan *et al.* (2014) in soybean and Samson and Helmut (2007) in cowpea reported earlier that water deficit stress reduced significantly the total leaf area. The results of this study agrees with those found by Muhammad and Jan (2016) who revealed that compost amendment enhanced maize crop yield and yield components.

#### 5.3.5 Days to 50% flowering

During the study period it was observed that maize plants grown under combination of SUC delayed tasseling, while under other treatments tassel emergence was earlier. Delayed tasseling due to compost treatment could be associated with vigorous and prolongs vegetative growth as a

result of higher nutrient availability. The results were supported by Li and Cai (2003) who concluded that tasseling in maize was delayed when compost was applied. Dolan *et al.* (2006) also found that compost incorporation had delayed tasseling, which might be due to more fertilizer availability and improved soil condition and fertility. Imran *et al.* (2015) stated that increasing nitrogen level consistently increased days to 50% tasseling due to prolonging the vegetative growth period. Another observation made was that at 11<sup>th</sup> week after emergence some plants from all treatments had tasseled while maize plants in control and urea treatments were already shedding pollen. According to Carvaco *et al.* (2003) in maize, tasseling normally occurs 2 to 3 days before silk emergence and this varies between genotypes. Various studies (Zamir, 1998; Modarres *et al.* (1998); Gozubenli, 2001) reported that variation in tasseling and silking period of maize hybrid is due to its genetic makeup.

Furthermore the results indicated that days to silking was delayed by compost application and drought stress. Maize planted in SC and SUC treatments had prolonged silking stage ranging between 59- 60 days as compared to control treatments. This could be explained that N content in compost lengthened the vegetative growth period. These results are in line with the findings of Dolan *et al.*, (2006) who reported that compost application had delay silking in maize.

## 5.3.6 Days to 90% physiological maturity, yield and yield components.

With respect to maturity, the number of days required for 90% maturity was influenced by the interaction of water deficit and compost. Application of compost (SUC) caused delayed (120 days) physiological maturity under irrigation condition and early (117 days) to physiological maturity were observed under control treatment under drought stress. In accordance with Bekele *et al.* (2018), physiological maturity in maize was significantly prolonged by applications of lime, vermicompost, and chemical P fertilizer. The result of this study does not agree with the findings

of Hegde and Dwivedi (1993) who found that integration of organic manure with inorganic fertilizers were observed to hasten maturity period of the potato crop. This could be described that crop response to fertilizer application depends on the morpho-physiological characteristics, species, cultivar and the rate of application.

#### 5.3 7 Maize biomass production.

Water deficiency imposes limitations on production of biomass by plants. In this study, these limitations were primarily observed in plants subjected to water stress and that were not supplemented with compost. The results demonstrated that the shoots, roots and total biomass yield of maize was negatively affected by water-deficit stress and fertilizer amendments, while these characteristics were significantly increased by the application of compost under both conditions. This results agrees with the work of Nazarideljou and Heidari (2014) who reported that reduction in growth and productivity is a common response of many crop to water deficit. The findings are also in agreement with the results from El-Mageed *et al.* (2018), who reported a significant increase in sorghum shoot biomass after compost addition to a sandy loam, in both water stressed and unstressed conditions. Recently, Zhang *et al.* (2020) demonstrated increased growth and biomass production in cotton due to organic fertilizer through modification of root length, volume and surface area. Kibunja *et al.* (2010) also observed that total biomass of maize was higher in treatment combination of organic and inorganic fertilizer.

Plants that were grown under non-stressed water condition had the highest shoot, root and total biomass yield compared with those that were planted under stressed condition. Generally, the results in this study were consistent with a published data by Abbas *et al.* (2018), which report a decrease in wheat growth and biomass under water stress conditions. The reduction in yield of plants irrigated at four days interval indicates that these plants were subjected to water deficit stress

and yield decreasing may be explained by effect of water deficit stress (Bouazzama et al., 2012; Dhakar et al., 2018). The parameters examined exhibited similar trends, where the highest values were attained from plants with compost application and full irrigation as comparable with control under both experiments. Compost application is one of the important practical measures to enhance seed yield under water stress condition as reported by EL Sabagh et al. (2015b). The results obtained from this experiment could be associated to the fact that addition of compost significantly alleviated the negative impacts of the drought stress. Such trend was consistent with the trend observed for other plant growth parameters. Irrigation water applied at the beginning of the intensive vegetative growth stage increased the process of biomass accumulation. The adverse effect of drought on dry matter accumulation appeared to be significant during tasseling stage. Similar result was also observed by Alghabari and Isham, (2018) that drought stress affected barley yield through impaired grain development and grain filling duration. Serious decreases have been recorded for control and SU plants under both conditions. According to Amanullah et al. (2015), application of compost along with N was found to be the best combination to yield and yield components of maize. Gholami and Zahedi (2019) stated that the reduction of yield, yield component and quality under drought stress could be due to numerous reasons including decrease of photosynthesis efficiency, leaf area, net assimilation rate, and reduction of water and mineral absorption by the root which ultimately decline developmental and vegetative growth.

# 5.4 Effects of water regime and compost amendment on chlorophyll content, photosynthetic rates and intrinsic water use efficiency of maize crop.

# 5.4.1 Maize chlorophyll content

In the present study, the application of compost alone or together with urea fertilizer and adequate soil moisture content were very effective in helping maize plants to decrease the detrimental effects

of drought stress on leaf chlorophyll content. Soil amended with compost fertilizer (SC and SUC) significantly high chlorophyll content of maize leaves and soil water content under both conditions. In general chlorophyll meter readings of maize leaves responded positively to compost amendment (SC and SUC) in both conditions. This could be ascribed to the fact that compost was able to retain soil moisture. This agrees with the work of EL Sabagh et al. (2016a) where it was observed that chlorophyll content of soybean plant decreased significantly under high levels of water deficit conditions. Leaf chlorophyll content is influenced by soil and environmental factors. According to Hosseinzadeh et al. (2018) chlorophyll content can be tolerance index to water stress in plants. In addition, the results also showed that with increased drought stress, leaf chlorophyll content decreases and applying compost the leaf chlorophyll content increases. Our results are in line with, Ndiso et al. (2017) and Tembe et al. (2017) who both reported that drought stress significantly reduced chlorophyll content in cowpeas and tomato, respectively. It was observed from the experiment that compost amendment prolonged the green leaves which basically allows for a longer period of leaf photosynthesis. Overall, the majority of chlorophyll lost from plant leaves subjected to drought stress is lost from the mesophyll cells. Huerta-Pujol et al. (2010) went on to explain that the reasons for this preferential loss could be attributed to the fact that the mesophyll cells are farther removed from the vascular supply of water than the bundle sheath cells and hence develop greater cellular water deficits which lead to a greater loss of chlorophyll. Chlorophyll content of maize subjected to the compost treatment was significantly higher when comparable with control treatments. This significant increase maybe due to the improvement of the nutritional condition of soil especially N, which reflected on the growth of the plants.

#### 5.4.2 Photosynthetic rates and intrinsic water use efficiency.

In this study, maize under drought stress more especially the control without compost application had considerable reduced photosynthetic rate as compared to irrigated or well watered treatments. During water stress, stomatal closure leads to decreased leaf conductance, photosynthesis and transpiration. Due to the sensitive response of leaf conductance to reduced leaf water potential, the more conservative use of water results in higher WUE in water-deficient plants, which may be a mechanism for improving resource use efficiency (Liu et al., 2016). Photosynthesis is one of the most important physico-chemical processes of higher plants that is directly linked to plant biomass production; however, it is very sensitive to drought stress (Yang et al., 2014). A study by Adugna (2016) proved that plant's photosynthesis rates improved with the availability of soil moisture due to the application of soil organic amendments such as biochar and compost. Stomata closure is an initial response of plants to drought stress (Pirasteh-Anosheh et al., 2016). When roots are exposed to water stress generate the chemical signals such as ABA that send response in the stomatal. Controlling water loss through stomatal closure has been considered as an early response of plants to water stress (Yan et al., 2016, Harb et al., 2010). In this study, under drought condition water use was significantly low resulting in high WUE. Severe decrease in terms of water use was recorded in control treatments indicating that WUE was affected by water stress and WUE decreased with an increase in water stress. This agreed with many previous studies who have found that WUE of various plant species is improved under water stress (Ye et al., 2013). It has been suggested that the drought-induced suppression of photosynthesis could be generally attributed to stomatal limitation and/or non-stomatal/metabolic limitation (Zhang. et al., 2013).

WUE describes the intrinsic trade-off between carbon fixation and water loss, because water evaporates from the interstitial tissues of leaves whenever stomata open for CO2 acquisition for photosynthesis (Bramley *et al.* 2013). It represents the ultimate performance of crop yield and water consumption, and it determines the water saving capacity and water productivity of crops. According to Mashilo *et al.* (2017) WUE is an important physiological adaptation mechanism that can improve crop productivity under conditions of water scarcity.

Furthermore, regarding compost application the results of this study revealed that maize crops sown in compost applied soil and combination of urea + compost were more water use efficient under both conditions. It is evident that applying compost significantly increases WUE and this could be attributed to the subsequent observed higher biomass yield. Consequently, the incorporation of compost treatment resulted increase in WUE compared with non-compost treatments.

# 5.5 Effects of compost amendments on nutrient content, nitrogen and phosphorus use efficiency of maize.

#### **5.5.1 Nutrient content**

This study showed that nutrient content of maize plants under irrigated and drought stress conditions were improved by compost amendment of soil. On the effects of treatments on the nutrient accumulation in the maize plant tissue at final harvest, maize grown on SC, SU and SUC had the highest content of Nitrogen, Potassium and Phosphorous as compared to control treatments and this could be attributed to increased availability of essential plant nutrients content in the soil. Sánchez *et al.* (2017) also reported the highest yield and tissue content of K and P where compost consisting of chicken manure was applied. Moreover, Zhang *et al.* (2016) documented that separate or combined application of bio-char and compost had a significant influence on plant N, P and K content in comparison with inorganic amendments. According to Carroll (2011) the amount of any micronutrient absorbed depends on the plant's response to the nutrient,

bioavailability of the nutrient and concentration of the nutrient around the root's surface of the plant.

# 5.5.2 Nitrogen use efficiency as influenced by compost amendments

The study revealed that enhancing sandy soils with compost significantly affected the agronomic NUE of maize under full irrigation and drought stressed conditions. NUE was significantly higher at treatments where compost was applied with urea fertilizer than where urea was applied without compost and control. Increased NUE as a result of combining compost with urea fertilizer was most likely attributed to the contribution of compost in alleviating other crop growth constraints. According to Souri and Hatamian (2019) it is well known that nutrients uptake and the water available to plant roots are closely related. Li et al. (2015) reported that under severe water stress, the photosynthetic capacity of the ear leaf decreased as did the dry matter production capacity, which resulted in yield decreases and limited plant N uptake, which in turn seriously affected N utilization in the plant. The application of organic manures to the soils causes increased in SOM, increased water holding capacity and aggregation stability, resulting in nutrient leaching reductions and improving the nutrient use efficiency (Baligar et al., 2001). However, the results of this study does not agree with the findings of Djaman et al. (2013), who found that excess of irrigation favors the residual loss of NO<sub>3</sub><sup>-</sup>, through either leaching or denitrification, due to its high mobility in soil under high moisture conditions, causing a reduction in the efficiency of N use for production. This is supported by Gholamhoseini et al. (2013) findings that higher N responses in maize yield under favorable soil water conditions, with an increase in reduced water regime under semiarid conditions. Plants take up N in the form of NH4<sup>+</sup>, a result of mineralization, and  $NO3^{-}$ , a result of nitrification. As mentioned, compost as an organic source when incorporated

into soil results in stabilization of nutrients against volatilization and leaching hence continual nutrient availability to the plants.

#### 5.5.3 Phosphorus use efficiency in maize as influenced by compost amendments.

This study indicated that PUE increased with available water and was generally greater for compost treatments, especially under irrigation condition as comparable to stressed condition. Increased PUE during the study period could be linked mainly to water availability to the maize crop grown under unstressed experiment. This work agrees with Qin et al. (2005), who detailed that the diffusion process, as a result of which phosphorus is carried towards the root, occurs faster in a moist rather than in a dry environment. This result indicated that optimizing water and compost application improved the PUE maize. Reductions in phosphorus use was observed under water deficit condition. With regard to compost addition, phosphorous use efficiency showed a remarkable result for the combined application of SC and SUC and these findings were also studied by Ademba et al. (2015) who reported that integrated use of phosphate fertilizers (inorganic) and manure( organic) applications significantly improved maize yield and PUE. Limitation of grain crop productivity by phosphorus (P) is widespread and will probably increase in the future. Improving the efficiency of phosphorus (P) fertilizer use for crop growth requires enhanced P acquisition by plants from the soil (P-acquisition efficiency) and enhanced use of P in processes that lead to faster growth and greater allocation of biomass to the harvestable parts (P-use efficiency (PUE). Phosphorus use efficiency in plants is a complex trait that is controlled by both P uptake, or P acquisition efficiency, and P utilization efficiency Mendes et al., 2014; Manschadi et al., 2014).

# **CHAPTER SIX**

#### **CONCLUSION AND RECOMMENDATIONS**

#### **6.1 Conclusions**

**6.1.1** The study verified that compost applied alone or in combination with chemical fertilizers offers potential to enhance soil quality and improve crop yield. Among all treatments, SC and SUC treatments showed potential as a soil conditioner, which directly improved soil physicochemical features such as soil pH, EC, CEC, TC, TN, available P and exchangeable cations under any condition.

**6.1.2** Hypothesis that compost amendment under drought stressed condition is the key factor to retain soil moisture and increase the productivity of maize crop has been confirmed by this study. Compost efficacy on morph-physiological indices of maize under both well watered as well as stressed condition has been observed in this study.

**6.1.3** Among all the treatments studied, SUC (soil + compost +urea) treatments greatly increased nitrogen use efficiency of maize (NUE) as compared to SU (soil +urea), SC (soil + compost) and control treatments recorded lower value under water deficit and well watered conditions.

**6.1.4** From the above obtained results, SC (soil compost) and SUC (soil +compost + urea) had the highest phosphorus use efficiency (PUE) whereas SU (soil + urea) and control treatments recorded the lowest values in both conditions. It can then be concluded that compost amendment has effect on PUE.

**6.1.5** The current study proved that compost amendments is the best approach to overcome drought stress effects on maize crop. The results revealed that soil + compost (SC), soil+ urea (SU) and

soil +compost +urea (SUC) and control treatments had the highest values under irrigated condition although under drought stressed condition control treatments exhibited the lowest value. Finally it is concluded that amending soil with compost is a good strategy to promote water use efficiency (WUE) and therefore, it is considered useful under stressed conditions.

## **6.2. Recommendations**

- **6.2.1** Further studies should be on soil-crop modelling as a tool for understanding the collaborative effects between water and nutrient use in yield, WUE and NUE, and for improved managing approaches.
- **6.2.2** There is a need to evaluate the response of different varieties of maize to confirm genotypic variation in water use efficiency and nutrient use efficiency under rain-fed environment.
- **6.2.3** Further research studies should include compost in evolving nutrient response curves, for various cereals crops under rain-fed conditions.
- **6.2.4** Investigations should be done on the prolonged existence of compost in various soil textural class after one application under rain –fed condition.

- Abbas, T., Rizwan, M., Ali, S., Adrees, M., Mahmood, A., Zia-Ur-Rehman, M., Ibrahim, M., Arshad, M., Qayyum, M.F. (2018). Biochar application increased the growth and yield and reduced cadmium in drought stressed wheat grown in an aged contaminated soil. *Ecotoxicology and Environmental Safety*. 148: 825–833.
- Abdulai, A.L. (2005). Morphological and physiological responses of sorghum (Sorghum bicolor
   L. Moench) to different patterns of drought. MSc Thesis. Rheinische Friedrich Wilhems-University, Ghana.
- Adams, H. D., Germino, M. J., Breshears, D. D., Barron-Gafford, G. A., Guardiola-Claramonte, M., Zou, C. B., Huxman, T. E. (2013). Nonstructural leaf carbohydrate dynamics of Pinus edulis during drought-induced tree mortality reveal role for carbon metabolism in mortality mechanism. *New Phytologist*. 197:1142–1151.
- Adamu, U. K., Mrema Jerome, P., Msaky, J. J. (2015). Growth response of maize (Zea mays L.) to different rates of nitrogen, phosphorus and farm yard manure in Morogoro Urban district, Tanzania. *American Journal of Experimental Agriculture*. 9:1–12.
- Ademba, J.S., Kwach, J.K., Esilaba, A.O., Ngari, S.M. (2015). The effects of phosphate fertilizers and manure on maize yields in South Western Kenya, Egypt. *East African Agricultural and Forestry Journal.* 81: 1-12.
- Adewopo, J.B., VanZomeren, C., Bhomia, R.K., Almaraz, M., Bacon, A.R., Eggleston E., Judy J.D, Lewis, R.W., Lusk, M., Miller ,B., Moorberg, C., Snyder, E.H., Tiedeman, M. (2014)

Top-ranked priority research questions for soil science in the 21 century. *Soil Science Society of American Journal*. 78:337–347.

- Adugna, G. (2016) A review on impact of compost on soil properties, water use and crop productivity. Academic Research Journal of Agricultural Science and Research. 4: 93– 104.
- Agegnehu, G., Nelson, P. N., and Bird, M. I, (2016) Crop yield, plant nutrient uptake and soil physicochemical properties under organic soil amendments and nitrogen fertilization on Nitisols. *Soil and Tillage Research*.160: 1–13.
- Agegnehu, G., Nelson, P.N., Bird, M.I., (2016b). The effects of biochar, compost and their mixture and nitrogen fertilizer on yield and nitrogen use efficiency of barley grown on a nitisol in the highlands of Ethiopia. *Science of the Total Environment*. 569: 869–879.
- Agegnehu, G., van Beek, C., Bird, M. (2014). Influence of integrated soil fertility management in wheat and productivity and soil chemical properties in the highland tropical environment. *Journal of Soil Science and Plant Nutrition*. 14: 532- 545.
- Albrecht, G., Ketterings, Q. M., Beckman, J. (2005). Soil pH for Field Crops: Agronomy Fact Sheet Series. New York. Retrieved from <u>http://nmsp.css.cornell.edu</u>
- Alghabari, F., Ishan, M.Z. (2018). Effect of drought stress on growth, grain filling duration, yield and quality attributes of barley (*Houdium vulgare l.*). *Bangladesh Journal of Botany*. 47: 421-428.

- Amanullah, H., Stewart, B. A. (2015). Analysis of growth response of cool season cereals "wheat vs. rye" grown in organic and inorganic soils. *Emirates Journal of Food and Agriculture*. 27: 430–440.
- Amundson, R., Berhe, A.A., Hopmans, J.W., Olson, C., Sztein, A.E., Sparks, D.L. (2015). Soil and human security in the 21<sup>st</sup> century. *Science*. 348:647–653.
- Angelova, V.R., Akova, V.I., Artinova, N.S., Ivanov, K.I. (2013). The effect of organic amendments on soil chemical characteristics. *Bulgarian Journal of Agricultural Science*. 19:958–971
- Anjum, S.A., Wang, L.C., Farooq, M., Hussain, M., Xue, L.L., Zou, C.M. (2011a). Brassinolide application improves the drought tolerance in maize through modulation of enzymatic antioxidants and Leaf gas exchange. *Journal of Agronomy and Crop Science*. 197:177-185.
- AOAC, (2011). Official Methods of Analysis. Association of Official Agricultural Chemists, (18<sup>th</sup> ed). Gaithersburg, MD.
- Araus, J.L., Sanchez, C., Edmeades, G.O. (2011). Phenotyping maize for adaptation to drought.In: Monneveux P, Ribaut JM, editors. Drought phenotyping in crops: from theory to practice CGIAR Generation Challenge Program; pp. 263–283.
- Ayuso, M.A., Pascal, J.A., Garcia, C., Hernandez, B. (1996). Evaluation of urban wastes for agricultural use. *Soil Plant Nutrition*. 42: 105-111.
- Aziz, T., Ullah, S., Sattar, A., Nasim, M., Faroog, M., Khan, M.M. (2010). Nutrient availability and maize (Zea mays L.) growth in soil amended with organic manures. *International Journal of Agriculture and Biology*. 12: 621-624.

- Baligar, V.C., Fageria, N.K., and He, Z.L. 2001. Nutrient use efficiency in plants. Journal of Communications in soil science and plant analysis. 32: 921-950.
- Barbosa, A. M., Guidorizi, K. A., Catuchi, T. A., Marques, T. A., Ribeiro, R. V., Souza, G. M. (2015). Biomass and bioenergy partitioning of sugarcane plants under water deficit. *Acta Physiologiae Plantarum*. 37:142-157.
- Basu, S., Ramegowda, V., Kumar, A., Pereira, A. (2016.) Plant adaptation to plant stress. Crop,Soil, and Environmental Sciences, University of Arkansas, Fayetteville, Arkansas, 72701,USA.
- Baveye P.C (2015). Grand challenges in the research on soil processes. *Front Environmental Science* 3:1–5.
- Beer, C., Ciais, P., Reichstein, M., Baldocchi, D. (2009). Temporal and among-site variability of inherent water use efficiency at the ecosystem level. Glob Biogeochem Cycles. 23:GB2018.
- Bekele, A., Kibret, K., Bedadi, B., Balemi, T., Yli-Halla, M. (2018). Effects of lime, vermicompost and chemical P fertilizer on yield of maize in Ebantu District, Western highlands of Ethiopia. *African Journal of Agricultural Research*. 13:477–489.
- Bell, C. (2016). The importance of nitrogen for plant health and productivity. *Growcentia:* Mammoth.
- Bell, M. (2014). Use of enhanced efficiency fertilizers to increase fertilizer nitrogen us efficiency in sugarcane. (Bell, M. J., Ed) A review of nitrogen use efficiency in sugarcane. Australia.

- Bhattacharya, A. (2019). Changing Climate and Resource Use Efficiency in Plants reviews the efficiencies for resource use by crop plants under different climatic conditions. pp 111-180.
- Bossio, D., Noble, A., Molden, D., Nangia, V. (2008). Land degradation and water productivity in agricultural landscapes. In Conserving land, protecting water, ed., Bossio, D.; Geheb, K. Wallingford, UK: CABI; Colombo, Sri Lanka: International Water Management Institute (IWMI); Colombo, Sri Lanka: CGIAR Challenge Program on Water and Food, pp.20-32. (Comprehensive Assessment of Water Management in Agriculture Series 6).
- Bouajila, K., Sanaa, M. (2011). Effects of Organic Amendments on Soil Physico-chemical and Biological Properties. *Journal of Materials and Environmental Science*. 2:485-490.
- Bouazzama, B., Xanthoulis, D., Bouaziz, A., Ruelle, P., Mailhol, J.C. (2012). Effect of water stress on growth, water consumption and yield of silage maize under flood irrigation in a semiarid climate of Tadla (Morocco). *Biotechnology, Agronomy, Society and Environment*.16: 468-477.
- Boul, S.W., Southard, R.J., Graham, R.C., McDaniel, P.A. (2003). Soil genesis and classification, (5<sup>th</sup> ed). Iowa State Press, Ames, Iowa. pp 494.
- Bramley, H., Turner, N.C., Siddique, K.H.M. (2013) Water use efficiency. In: Kole C (ed). Genomics and breeding for climate-resilient crops. 2: 225-268.
- Brar, B.S., Singh, J., Singh, G., Kaur, G. (2015). Effects of Long Term Application of Inorganic and Organic Fertilizers on Soil Organic Carbon and Physical Properties in Maize-Wheat Rotation. *Agronomy:* 5: 220-238.

- Bu, L., Zhang, R., Chang, Y., Xue, J., Han, M (2010). Response of photosynthetic characteristics to water stress of maize leaf in seeding. *Acta Ecological Sinica Journal*. 30: 1184-1191.
- Bulluck, L.R., Brosius, M., Evanylo, G.K., Ristaino, J.B. (2002). Organic and synthetic fertility amendments influence soil microbial, physical and chemical properties on organic and conventional farms. *Applied Soil Ecology*. 19:147-160.
- Bunce, J.A. (2010). Leaf transpiration efficiency of some drought-resistant maize lines. Crop Science. 50:1409-1415.
- Cai, Z. J., Wang, B. R., Xu, M. G., Zhang, H. M., He, X. H., Zhang, L., Gao, S. D. (2015). Intensified soil acidification from chemical N fertilization and prevention by manure in an 18-year field experiment in the red soil of southern China. *Journal of Soils and Sediments*. 15: 260–270.
- Campbell, C. D., Sage, R. F. (2006). Interactions between the effects of atmospheric CO<sub>2</sub> content and P nutrition on photosynthesis in white lupin (*Lupinus albus L.*). *Plant Cell and Environment*. 29: 844-853.
- Campos, H., Trejo, C., Peña-Valdivia, C.B., García-Nava, R., Conde-Martínez, F.V., Cruz-Ortega,
   M. (2014). Stomatal and non-stomatal limitations of bell pepper (Capsicum annuum L.)
   plants under water stress and re-watering: Delayed restoration of photosynthesis during
   recovery. *Environmental and Experimental Botany*. 98: 56–64.

Carroll, S. (2011). Absorption of water and mineral salts in plants.

- Carvaco J., Andrieu B., Otegui M.E. (2003). Silk elongation in maize: relationship with flower development and pollination. *Crop Science Journal*. 43:914-920.
- Ch'ng H.Y., Ahmed ,O.H, Majid N.M.A. (2015) Improving phosphorus availability, nutrient uptake and dry matter production of Zea mays L. on a tropical acid soil using poultry manure biochar and pineapple leaves compost. *Experimental Agriculture* 52: 447–465.
- Chakraborty, A., Chakrabarti, K., Chakraborty, A., Ghosh, S. (2011). Effect of long-term fertilizers and manure application on microbial biomass and microbial activity of a tropical agricultural soil. *Biology and. Fertility of Soils*. 47: 227-233.
- Chandini, K. R., Kumar, R., Prakash, O. (2019). The impact of chemical fertilizers on our environment and ecosystem. *Research trends in environmental Sciences*. 2: 69-86.
- Chen, D., Wang, S., Cao, B., Cao, D., Leng, G., Li, H., Yin, L., Shan, L., Deng, X. (2015) Genotypic variation in growth and physiological response to drought stress and re-watering reveals the critical role of recovery in drought adaptation in maize seedlings. *Frontiers in Plant Science*.6:1241-1256.
- Chen, J.H. (2006). The combined use of chemical and organic fertilizers and/or biologicalfertilizer for crop growth and soil fertility. *Proceedings of International Workshop on Sustained Management of the Soil-Rhizosphere System for Efficient Crop Production and Fertilizer Use. pp* 5 - 7.
- Chen, Y., Liu, T., Tian, X., Wang, X., Li, M., Wang, S. (2015). Effects of plastic film combined with straw mulch on grain yield and water use efficiency of winter wheat in Loess Plateau. *Field Crops Research*. 172: 53–58.

- Conde, L.D., Chen, Z., Chen, H., Liao, H. (2014). Effects of phosphorus availability on plant growth and soil nutrient status in the rice/soybean rotation system on newly cultivated acidic soils. *American Journal of Agricultural and Forestry*. 2: 309-316.
- Cortina, J., Vilagrosa, A., Trubat, R. (2013). The role of nutrients for improving seedling quality in drylands. *New Forests*. 44: 719-732.
- Coulibaly, S.S., Kouassi, K.I., Koffi, K.K., Zoro, B. I. A. (2019). Effect of compost from different animal manures on maize (*Zea mays*) growth. *Journal of Experimental Biology and Agricultural Sciences*.7: 178-185.
- Cramer, M. D., Hawkins, H. J., Verboom, G. A. (2009). The importance of nutritional regulation of plant water flux. *Oecologia*.161:15-24.
- Crogger, C.G. (2005). Potential compost benefits for restoration of soils disturbed by urban development. *Compost Science and Utilization*. 13: 243-251.

Crouse, K., Denny, G. (2015). Soil pH and Fertilizers. Plant and Soil Sciences. 372: 7-8.

- Cui, G., Zhao, X., Liu, S., Sun, F., Zhang, C., Xi, Y. (2017). Beneficial effects of melatonin in overcoming drought stress in wheat seedlings. *Plant Physiology and Biochemistry*. 118: 138-149.
- Du Plessis, J. (2003). Maize production, Department of Agriculture, Republic of South Africa. pp 1-35.

- Dhakar, R., Chandran, M.A.S., Nagar, S., Kumari, V.V., Subbarao, A.V.M., Bal, S.K., Kumar,
   P.V. (2018). Field crop response to water deficit stress: Assessment through crop models.
   Advances in Crop Environment Interaction.11: 287-315.
- Dhillon, J., Torres, G., Driver, E., Figueiredo, B., Raun, W. R. (2017). World phosphorus use efficiency in cereal crops. *Agronomy Journal*. 109:1670-1677.
- Diacono, M., Montemurro, F. (2010). Long-term effects of organic amendments on soil fertility. *A review Agronomy for Sustainable Development*. 30: 401–422.
- Djaman, K., Irmak, S., Martin, D. L., Ferguson, R. B., Bernards, M. L. (2013). Plant nutrient uptake and soil nutrient dynamics under full and limited irrigation and rain-fed maize production. *Agronomy Journal*.105: 527-538.
- Dolan, M.S., Saleem, M.F., Cheema, M.A., Hammad, H.M. (2006). Effect of poultry manure level on the productivity of spring maize. *Journal of Animal and plant Sciences*.19: 122-125.
- Donn, S., Wheatley, R. E., McKenzie, B. M., Loades, K. W., Hallett, P. D. (2014). Improved soil fertility from compost amendment increases root growth and reinforcement of surface soilon slope. *Ecological Engineering*. 71: 458-465.
- Dos Santos, M. G., Ribeiro, R. V., de Oliveira, R. F., Machado, E. C., Pimentel, C. (2006). The role of inorganic phosphate on photosynthesis recovery of common bean after a mild water deficit. *Plant Science*. 170: 659–664.
- Dos Santos, M. G., Ribeiro, R. V., Oliveira, R. F., Pimentel, C. (2004). Gas exchange and yield response to foliar phosphorus application in Phaseolus vulgaris L. under drought. *Brazilian Journal of Plant Physiology*. 16: 171–179.

- Du, T.S., Kang, S.Z., Zhang, J.H., Davies, W.J. (2015). Deficit irrigation and sustainable waterresource strategies in agriculture for China's food security. *Journal of Experimental Botany*. 66: 2253–2269.
- Edwards, S., Hailu, A. (2011). How to make compost and use. In: Ching. L. L., Edwards, S., and Nadia, H. S. (Ed), *Climate Change and Food Systems Resilience in Sub-Saharan Africa*. FAO, Italy. pp 379-436.
- Egilla, J.N., Davies F.T., Boutton, T.W. (2005). Drought stress influences leaf water content, photosynthesis, and water use efficiency of Hibiscus rosa-sinensis at three potassium concentrations. *Photosynthetica*. 43:135-140.
- EL Sabagh A., Sorour S., Omar A., Islam M.S., Ueda A., Saneoka H., Barutçular C., (2015b)
   Soybean (Glycine max L.) growth enhancement under water stress conditions.
   *International Conference on Chemistry Agriculture and Biological Science (CABS-2015) Sept. 4-5, Istanbul* (Turkey).
- EL Sabagh, A., Sorour, S., Morsi A., Islam, M.S., Ueda, A., Barutcular, C., Arioglu, H., Saneoka, H. (2016a). Role of osmoprotectants and compost application in improving water stress tolerance in soybean (*Glycine max L.*). *International Journal of Current Research*. 8: 25949-25954.
- El-Mageed, T.A.A., El-Samnoudi, I.M., Ibrahim, A.E.A.M., El Tawwab, A.R.A. (2018). Compost and mulching modulates morphological, physiological responses and water use efficiency in (sorghum bicolor L. Moench) under low moisture regime. *Agricultural Water Management*. 208: 431-439.
- Estefan, G., Sommer, R., Ryan, J (2013). Methods of Soil, Plants and Water Analysis: A manual for the West Asia and North Africa region (3<sup>rd</sup> edition). Beirut: International Center for Agricultural Research in the dry Areas (ICARDA).
- Eurostat (2015). Statistics Explained (http://ec.europa.eu/eurostat/statistics-explained/). (accessed March 2015).
- Ewing, P. M., Runck, B. C. (2015). Optimizing nitrogen rates in the mid-western United States for maximum ecosystem value. *Ecological. Society.* 20:18-32.
- Fageria, N. K., Baligar, V. C. (2005). Enhancing nitrogen use efficiency in crop plants. Advances in Agronomy. 88: 97-185.
- Fan, T.L., Stewart, B., Yong, W., Luo, J.J., and Zhou, G.Y. (2005). Long-term fertilization effects on grain yield, water-use efficiency and soil fertility in the dry land of Loess Plateau in China. *Agriculture, Ecosystems and Environment*. 106: 313-329.
- Fanadzo, M., Chiduza, C., Mnkeni, P.N.S. (2009). Comparative response of direct seeded and transplanted maize (Zea mays L.) to nitrogen fertilization at Zanyokwe irrigation scheme, Eastern Cape, South Africa. *African Journal for Agriculture*. 4:689-694.
- FAOSTAT. (2014). Statistical database of the Food and Agriculture of the United Nations.
- Farooq, M., Wahid, A., Kobayashi, N., Fujita, D., Basra, S. M. A. (2009). Plant drought stress: Effects, mechanisms and management. *Agronomy for Sustainable Development*. 29: 185-212.
- Farrell, M., Jones, D.L. (2009). Critical evaluation of municipal solid waste composting and potential compost markets. *Bio resource Technology*. 100: 4301-4310.

- Faustino, L. I., Bulfe, N. M. L., Pinazo, M. A., Monteoliva, S. E., Graciano, C. (2013). Dry weight partitioning and hydraulic traits in young *Pinus taeda* trees fertilized with nitrogen and phosphorus in a subtropical area. *Tree Physiology*. 33: 241–251.
- Fischer, K.S., Fukai, S., Kumar, A., Leung, H., Jongdee, B. (2011). Phenotyping rice for adaptation to drought. In: Monneveux P, Ribaut JM, editors. Drought phenotyping in crops: from theory to practice: *CGIAR Generation Challenge Program*; pp. 215–243.
- Fleisher, D. H., Wang, Q., Timlin, D. J., Chun, J. A., Reddy, V. R. (2012). Response of potato gas exchange and productivity to phosphorus deficiency and carbon dioxide enrichment. *Crop Science*. 52: 1803–1815.
- Galiano, L., Martínez-Vilalta, J., Lloret, F. (2011). Carbon reserves and canopy defoliation determine the recovery of Scots pine 4 yr after a drought episode. *New Phytologist*. 190:750–759.
- Gall, H., Philippe F., Domon, J., Gillet, F., Pelloux, J., Rayon, C. (2015). Cell Wall metabolism in response to abiotic stress. *Plants*. 4:112–66.
- Gao, M., Liang, F., Yu, A., Li, B., Yang, L. (2010). Evaluation of stability and maturity during forced- aeration composting of chicken manure and sawdust at different C/N ratios. *Chemosphere*. 78: 614 - 619.
- Gebreyesus, B.T. (2012). Effect of tillage and fertilizer practices on sorghum production in Abergelle area, North Ethiopia. *Momona Ethiopian Journal of Science*. 4: 52-69.

Gete, Z., Getachew, A., Dejene, A., Shahid, R. (2010). A Report on Fertilizer and Soil Fertility Potential in Ethiopia: Constraints and opportunities for enhancing the system. IFPR.

Ghannoum, O. (2009). C4 photosynthesis and water stress. Annals of Botany.103: 635–644.

- Gheysari, M., Mirlatifi, S.M., Bannayan, M., Homaee, M., Hoogenboom, G. (2009). Interaction of water and nitrogen on maize grown for silage. *Agricultural Water Management*. 96:809-821.
- Gholamhoseini, M., Agha-Alikhani, M., Sanavy, S. A. M. M., Mirlatifi, S. M. (2013). Interactions of irrigation, weed and nitrogen on corn yield, nitrogen use efficiency and nitrate leaching. *Agricultural Water Management*. 126: 9-18.
- Gholami, R., Zahedi, S. M. (2019). Identifying superior drought-tolerant olive genotypes and their biochemical and some physiological responses to various irrigation levels. *Journal of Plant Nutrition*. 42: 2057-2069.
- Gozubenli, H., Ulger, A.C., Sener, O. (2001). The effect of different nitrogen doses on grain yield and yield related characters of some maize genotypes grown as second crop. *Journal of Agricultural Faculty. C. U.* 16:39-48.
- Graça, J. P., Rodrigues, F. A., Farias, J. R. B., Oliveira, M. C. N., Hoffmann-Campo, C. B., Zingaretti, S. M. (2010). Hydric and nitrogen dose. *African Journal of Agricultural Research*. 11: 1475–1485.
- Gu, Y.J., Han, C.L., Fan, J.W., Shi, X.P., Kong, M., Shi, X.Y., Li, F.M. (2018). Alfalfa forage yield, soil water and P availability in response to plastic film mulch and P fertilization in a semiarid environment. *Field Crops Research*. 215: 94-103.

- Hammond, J. P., Broadley, M. R., White, P. J., King, G. J., Bowen, H. C., Hayden, R. Meacham, M.C., Mead, A., Overs, T., Spracklen, W.P., Greenwood, D.J. (2009). Shoot yield drives phosphorus use efficiency in *Brassica oleracea* and correlates with root architecture traits. *Journal of Experimental Botany*. 60: 1953–1968.
- Haouvanga, L.C., Alberta, N., Martin, Y., Mbaiguinam, M. (2017). Growth Response of *Moringa* oleifera L.as Affected by Various Amounts of Compost under Greenhouse Conditions.
   Annals of Agricultural Sciences. 62:221-226.
- Harb, A., Krishnan, A., Ambavaram, M.M., Pereira, A. (2010). Molecular and physiological analysis of drought stress in Arabidopsis reveals early responses leading to acclimation in plant growth. *Plant Physiology*. 154: 1254–1271.
- Hati, K.M., Mandal, K.G., Misra, A.K., Ghosh, P.K., Bandyopadhyay, K.K. (2006). Effect of inorganic fertilizer and farmyard manure on soil physical properties, root distribution, and water-use efficiency of soybean in Vertisols of central India. *Bioresource Technology*. 97: 2182-2188.
- Havlin, J.L., Tisdale, S.L., Nelson, W.L., Beaton, J.D. (2014). Soil Fertility and Nutrient Management: An Introduction to Nutrient Management, 8th ed.; Pearson: Upper Saddle River, NJ, USA, pp. 516.
- Hayatu M., Muhammad S.Y., Abdu H.U. (2014). Effect of water stress on the leaf relative water content and yield of some Cowpea (Vigna Unguiculata (L) Walp.) genotype. *International Journal of Science and Technology Research*. 3: 148-152.

- He, J., Wen, R.Tian, S., Su, Y., He, X., Su, Y., Cheng, W., Huang, K., Zhang, S. (2017). Effects of drought stress and re-watering on growth and yield of various maize varieties at tasseling stage. *Journal of South African Agriculture*.
- Hegde, D.M., Dwivedi, B.S. (1993). Integrated nutrient supply and management as a strategy to meet nutrient demand. *Fertilizer Research*. 38:49-59.
- Hirel, B., Gallais, A. (2011). Nitrogen use efficiency—physiological, molecular and genetic investigations towards crop improvement. In: Prioul JL Thévenot C Molnar T, eds. Advances in maize: 3. Essential reviews in experimental biology. pp 285 –310.
- Hosseinzadeh S.R., Amiri H., Ismaili A. (2018). Evaluation of photosynthesis, physiological, and biochemical responses of chickpea (*Cicer arietinum* L. cv. Pirouz) under water deficit stress and use of vermicompost fertilizer. *Journal of Integrative Agriculture*. 17:2426-2437.
- Huang, G., Li, Y., Mu, X., Zhao, H., Cao, Y. (2017a). Water-use efficiency in response to simulated increasing precipitation in a temperate desert ecosystem of Xinjiang, *China Journal of Arid Land.* 9:823–836.
- Huang, Z., Liu, Y., Tian, F.P., Wu, G.L. (2020). Soil water availability threshold indicator was determined by using plant physiological responses under drought conditions. *Ecological Indicators*. 118: 106740-106751.
- Huerta-Pujol, O., Soliva M., Martínez-Farré, F. X., Valero, J., López, M. (2010). "Bulk density determination as a simple and complementary tool in composting process control," *Bioresource Technology*. 10: 995–1001.

- Huo, Y., Wang, M., Wei, Y., Xia, Z. (2016). Over expression of the maize psbA gene enhances drought tolerance through regulating antioxidant system, photosynthetic capability, and stress defence gene expression in tobacco. *Frontiers in Plant Sciences*.6:1223-1233.
- Hussain, H. A., Men, S., Hussain, S., Chen, Y., Ali, S., Zhang, S., *et al.* (2019). Interactive effects of drought and heat stresses on morpho–physiological attributes, yield, nutrient uptake and oxidative status in maize hybrids. *Scientific. Reports.* 9: 1–12.
- ICT International Pty Ltd. MPM 160 Moisture Probe Meter operation manual. Armidale NSW 2350 Australia. Accessed from www.ictinternational.com.au on 20/02/2019.
- Imran, S., Arif, M., Khan, A., Khan, M.A., Shah, W., et al. (2015) Effect of nitrogen levels and plant population on yield and yield components of maize. Advances in Crop Science and Technology. 1-7.
- Jacobson, K. (2013). From betterment to Bt maize. Agricultural development and the introduction of genetically modified maize to South African smallholders. Doctoral Thesis No. 2013:28.
  Faculty of Natural resources and Agricultural Sciences. Sweedish University of Agricultural Sciences, Uppsala.
- Jangpromma, N., Songrsi, P., Thammasiririak, S., Jaisil, P. (2010). Rapid assessment of chlorophyll content in sugarcane using a SPAD chlorophyll meter across different water stress conditions. *Asian Journal of Plant Science*. 9: 368–374.
- Jin, J., Lauricella, D., Armstrong, R., Sale, P., Tang, C. (2015). Phosphorus application and elevated CO2 enhance drought tolerance in field pea grown in a phosphorus-deficient vertisol. *Annals of Botany*. 116: 975-985.

- Jones, C. A., Jacobsen, J. S., Wraithl, J. M. (2005). Response of malt barley to phosphorus fertilization under drought conditions. *Journal of Plant Nutrition*. 28: 1605–1617.
- Kadioglu, A., Terzi, R., Saruhan, N., Saglam, A. (2012). Current advances in the investigation of leaf rolling caused by biotic and abiotic stress factors. *Plant Science*. 182:42–48.
- Kai, L. Z., Ngoh, G., Peng, H., Min, G. M., Ken, T. Y., Green, L., Student, D. (2012). Investigating The Effect of Soil pH on the Germination of Avicennia alba Seedlings.
- Kant, S., Bi, Y. M., Rothstein, S.J. (2011). Understanding plant response to nitrogen limitation for the improvement of crop nitrogen use efficiency. *Journal of Experimental Botany*. 62:1499-1509.
- Kassahun, D., Mekonnen, L.S. (2012). Integrated agronomic crop managements to improve teff productivity under terminal drought, water stress. Ismail Md. Mofizur R (eds.), In Tech, improve teff- productivity-under-terminal-drought.
- Khan, M. S. A., Karim, M. A., Haque, M. M. (2014). Genotypic differences in growth and ions accumulation in soybean under NaCl salinity and water stress conditions. Bangladesh *Agronomy Journal*. 17: 47-58.
- Kibunja, C. N., Mwaura, F.B., Mugendi, D.N., Kitonyo, E. M., Salema, M.P. (2010). Nitrogen (N) use efficiency under continuous maize-bean cropping system in semi-humid highlands of Kenya. *East African Agricultural and Forestry Journal*. 76: 115-120.
- Ku, Y. S., Yeung, W. K. A., Yung, Y. L., Li, M. W., Wen, C. Q., Liu, X., Lam, H. M. (2013).Drought Stress and Tolerance in Soybean.

- Lamaoui, M., Jemo, M., Datla, R., Bekkaoui, F. (2018). Heat and drought stresses in crops and approaches for their mitigation. *Frontiers in Chemistry*. 6:26-40.
- Lamm, F.R., Manges, H.L., Stone, L.R., Khan, A.H., Rogers, D.H (2005). Water requirement of subsurface drip-irrigated corn in northwest Kansas. Transactions of ASAE 38: 441-448.
- Lenssen, A. W., Cash, S. D., Hatfield, P. G., Sainju, U.M., Grey., W. R, Blodgett, S. L., Johnson, G. D. (2010). Yield, quality, and water and nitrogen use of durum and annual forages in two-year rotations. *Agronomy Journal*. 102: 1261–1268.
- Lenssen, A.W., Johnson, G.D., Carlson, G.R. (2007). Cropping sequence and tillage system influences annual crop production and water use in semiarid Montana. *Field Crops Research*. 100: 32-43.
- Leroy, B.L.M.M., Bommele, L., Reheul, D., Moens, M., De Neve, S. (2007). The application of vegetable, fruit and garden waste (VFG) compost in addition to cattle slurry in a silage maize monoculture. Effects on soil fauna and yield. *European Journal of Soil Biology*. 43: 91-100.
- Li G.H., Zhao B., Dong S.T., Liu P., Zhang J.W, He Z.J. (2015). Effects of coupling controlled release urea with water on yield and photosynthetic characteristics in summer maize. *Acta Agronomica Sinica*. 41: 1406–1415.
- Li, H., Han, Y., Cai, Z. (2003). Nitrogen mineralization in paddy soil of the Taihu region of China under anerobic condition, dynamics and model fitting. *Geoderma*.115:161-175.
- Li, X., Kang, S., Zhang, X., Li, F., Lu, H. (2018). Deficit irrigation provokes more pronounced responses of maize photosynthesis and water productivity to elevated CO2. *Agricultural Water Management*.195: 71–83

- Lin, H. (2014). A new worldview of soils. Soil Science Society of American Journal. 78:1831– 1844.
- Lipiec, J., Doussan, C., Nosalewicz, A., Kondracka, K. (2013). Effect of drought and heat stresses on plant growth and yield: a review. *International Agrophysics*. 27: 463–477.
- Liu J., Schulz, H., Brandl, S., Miehtke, H., Huwe, B., Glaser, B. (2012). "Short-term effect of biochar and compost on soil fertility and water status of a Dystric Cambisol in NE Germany under field conditions," *Journal of Plant Nutrition, Soil Science*.175: 1–10.
- Liu, B., Wua, C.H., Pana, P., Fua,Y., Heb ,Z., Wua, L., Li ,Q. (2019). Remediation effectiveness of vermicompost for a potentially toxic metal contaminated tropical acidic soil in China. *Ecotoxicology, Environment and Safety*. 182:109394-109409.
- Liu, B., Yue, Y.M., Li, R., Shen, W. J., Wang, K.L. (2014). Plant leaf chlorophyll content retrieval based on a field imaging spectroscopy system. *Sensors*. 14: 19910-19925.
- Liu, C.A., Li, F.M., Zhou, L.M., Zhang, R.H., Lin, S.L., Wang, L.J. (2013). Effect of organic manure and fertilizer on soil water and crop yields in newly-built terraces with loess soils in a semi-arid environment. *Agricultural Water Management*. 117: 123-132.
- Liu, E.K., Mei, X.R., Yan, C.R., Gong, D.Z., Zhang, Y.Q. (2016). Effects of water stress on photosynthetic characteristics, dry matter translocation and WUE in two winter wheat genotypes. *Agricultural Water Management*. 167: 75-85.
- Liu, S.L., Huang, D.Y., Chen, A.L., Wei, W.X., Brookes, P.C., Li, Y., Wu, J. S. (2014). Differential responses of crop yields and soil organic carbon stock to fertilization and rice straw incorporation in three cropping systems in the subtropics. *Agriculture, Ecosystems and Environment.* 184:51–58.

- Liu, Z.X., Liu, P., Jail, X.C., Cheng, Y., and Dong, S.T. (2015). Effects of irrigation and fertilization on soil microbial properties in summer maize field. *Journal of Applied Ecology*. 26: 113-121.
- Liu, Y., Villalba, G., Ayres, R.U., Schroder, H. (2008). Global phosphorus flows and environmental impacts from a consumption perspective. *Journal of Indian Ecology*. 12: 229– 247.
- Logsdon, S.D., Sauer, P.A., Shipitalo, M.J. (2017). Compost improves urban soil and water quality *Journal of water Resources and Protection:* 9: 345-357.
- Lone A.A., Allai B., Nehvi, F. (2013). Growth, yield and economics of baby corn (Zea mays L.) as influenced by Integrated Nutrient Management (INM) practices. *African journal of agricultural research*. 8: 4537–4540.
- Lu, Y, L., Chadwick, D., Norse, D., Powlsond, D., Shi, W. M. (2015). Sustainable intensification of China's agriculture: The key role of nutrient management and climate change mitigation and adaptation. *Agriculture, Ecosystems & Environment.* 209: 1–4.
- Mahmood, F., Khan, I., Ashraf. U., Shahzad, T., Hussain. S., Shahid, M. (2017). Effects of organic and inorganic manures on maize and their residual impact on soil physico-chemical properties. *Journal of Soil Science and Plant Nutrition*. 17:22–32.
- Mahmud A.J., Shamsuddoha A.T.M, Haque N. (2016). Effect of organic and inorganic fertilizer on the growth and yield of rice (Oryza sativa L.). *Nature and Science*. 14:45–54

Maize.net. (2018). Accessed on 30/03/2019 from http://www.maize.net.

- Mallarino, A. P. (2011). Corn and soybean response to soil pH level and liming. *Integrated Crop Management Conference*. 93–102.
- Manish K.P., Prakash M., Salikram G. (2017). Growth Attributing Traits of Maize Affected by Different Nutrient Management in Lamjung Nepal. *International Journal of Applied Sciences and Biotechnology*. 5:98-101
- Manivannan, S., Balamurugan, M., Parthasarathi, K., Gunasekaran, G., Ranganathan, L.S. (2009).
   Effect of vermicompost on soil fertility and crop productivity-beans (*Phaseolus vulgaris*).
   *Journal of Environmental Biology*. 30: 275 -281.
- Manschadi, A.M., Hans-Peter, K., Vollman, J., Eitzinger, J., Wenzel, W. (2014). Developing phosphorus-efficient crop varieties: An interdisciplinary research framework. *Field Crops Research*. 162: 87–98.
- Manyuchi M.M., Kadzngura I., Phiri., Muredzi P. (2013). Effect of vermicompost, vermiwash and application time on Zea mays growth. *International Journal of scientific Engineering and Technology*. 2.
- Mao, H., Wang, H., Liu, S., Li, Z., Yang, X., Yan, J., Li, J., Tran, L.S., Qin, F.A. (2015).
  Transposable element in a NAC gene is associated with drought tolerance in maize seedlings. *Nature Communication*. 6:8326-8339.
- Mao, J., Oik, D.C., Fang, X., He, Z., Rohr, K.S, (2008). Influence of animal manure application on the chemical structures of soil organic matter as investigated by advanced solid-state NMR and FT–IR Spectroscopy. *Geodema*. 146:353 -362.

- Marchi, E. C. S., Zotarelli, L., Delgado, J. A., Rowland, D. L., Marchi, G. (2016). Use of the Nitrogen Index to assess nitrate leaching and water drainage from plastic-mulched horticultural cropping systems of Florida. *International Soil and Water Conservation Research*. 4:237–244 Elsevier.
- Mashilo, J., Odindo, A.O., Shimelis, H.A., Musenge, P., Tesfay, S.Z., Magwaza, L.S. (2017).
  Drought tolerance of selected bottle gourd [*Lagenaria siceraria* (Molina) Standl.]
  landraces assessed by leaf gas exchange and photosynthetic efficiency. *Plant Physiology* and Biochemistry. 120: 75–87.
- McCarthy, H.R., Pataki, D.E., Jenerette, G.D. (2011) Plant water-use efficiency as a metric of urban ecosystem services. *Ecological Application*. 21:3115–27.
- Meena, R.S., Meena, V.S., Meena,S.K., Verma, J.P. (2015). The needs of healthy soils for a healthy world. *Journal of Cleaner Production*.102:560-561
- Mendes, F.F., Guimarães, L.J.M., Souza, J.C., Guimarães, P.E.O., Magalhaes, J.V., Garcia, A.A.F. *et al.* (2014). Genetic architecture of phosphorus use efficiency in tropical maize cultivated in a low-P soil. *Crop Science*. 54: 1530-1538.
- Mensah, A. K., Frimpong, K. A. (2018). Biochar and/or compost applications improve soil properties, growth, and yield of maize grown in acidic rainforest and coastal savannah soils in Ghana. *International journal of agronomy*.
- Ministry of Agricultural Development and Food security, Geographical Information System. Gaborone, Botswana.
- Mitchell, P. J., O'grady, A. P., Tissue, D. T., White, D. A., Ottenschlaeger, M. L, Pinkard, E.A. (2013). Drought response strategies define the relative contributions of hydraulic

dysfunction and carbohydrate depletion during tree mortality. *New Phytologist*. 197:862–872.

- Mohammed, H.G., Denney M. J., Iyekar, C. (2004). Use Composted Organic Wastes as Alternative to Synthetic Fertilizers for Enhancing Crop Productivity and Agricultural Sustainability on the Tropical Island of Guam. 13<sup>th</sup> International Soil Conservation Organization Conference – Brisbane.
- Morris, M. L. 2002. Impacts of International Maize Breeding Research in Developing Countries, 1966-1998. Mexico, D.F.: CIMMYT.
- Moser, S.B., Feil, B., Jampatong, S., Stamp, P. (2006). Effects of pre-anthesis drought, nitrogen fertilizer rate, and variety on grain yield, yield components, and harvest index of tropical maize. Agric. Water Manage. 81:41-58.
- Mugwe, J., Mugendi D., Kungu J., Mucheru-Muna M.M. (2009). Maize yields response to application of organic and inorganic input under on-station and on-farm experiments in central Kenya. *Experimental Agriculture*. 45: 47- 59.
- Muhammad, Z., Jan, M.T. (2016). Evaluation of different composts composition on the yield and yield components of maize (*Zea mays L.*). *Sarhad Journal of Agriculture*. 32: 156-167.
- Naeem, M., Khan, M. M. A. (2009). Phosphorus ameliorates crop productivity, photosynthesis, nitrate reductase activity and nutrient accumulation in coffee senna (*Senna occidentalis L.*) under phosphorus-deficient soil. *Journal of Plant Interaction*. 4: 145–153.
- Nazarideljou, M.J., Heidari, Z., (2014). Effects of vermicompost on growth parameters, water use efficiency and quality of Zinnia Bedding plants (Zinnia elegance 'Dreamland Red') under

different irrigation regimes. International Journal of Horticultural Science and Technology.1:141–150.

- Ndiso, J.B., Chemining wa, G.N., Olubayo, F.M., Saha, H.M. (2017). Effect of drought stress on canopy temperature, growth and yield performance of cowpea varieties. *International Journal of Plant and soil Science*. 9: 1-12.
- Nigussie, Z., Tsunekawa, A., Haregeweyn, N., Adgo, E., Nohmi, M., Tsubo, M., *et al.* (2017). Factors influencing small-scale farmers' adoption of sustainable land management technologies in north-western Ethiopia. *Land Use Policy* 67: 57–64.
- Norton, R. M., Wachsmann, N. G. (2006). Nitrogen use and crop type affect the water use of annual crops in south-eastern Australia. *Australian Journal of Agricultural Research*. 57:257-267.
- Obata, T., Witt, S., Lisec, J., Palacios–Rojas, N., Florez–Sarasa, I., Yousfi, S., *et al.* (2015).
   Metabolite profiles of maize leaves in drought, heat, and combined stress field trials reveal the relationship between metabolism and grain yield. *Plant Physiology*.169: 2665–2683.
- Ogbonna, D.N., Isirimah, N.O., Princewill, E. (2012). Effect of organic waste compost and microbial activity on the growth of maize in the utisoils in Port Harcourt, Nigeria. *African Journal of Biotechnology*.11z: 12546-12554.
- Ogola, J.B.O., Wheeler, T.R., and Harris, P.M. (2002). Effects of nitrogen and irrigation on water use of maize crops. *Field Crops Research*. 78: 105-117.

- Okoroafor, I.B., Okelola, E.O., Edeh, O., Nemehute, V.C., Onu, C.N., Nwaneri, T.C., Chinaka, G.I. (2013). Effect of organic manure on the growth and yield performance of maize in Ishiagu, Ebonyi State, Nigeria. *Journal of Agriculture and Veterinary Science*. 5: 28-31.
- Olabode, O.S., Sola, O., Akanbi, W.B., Adesina, G.O., and Babajide, P.A. (2007). Evaluation of *Tithonia diversifolia* (Hemsl.) A Gray for Soil Improvement. *World Journal of Agricultural Sciences*. 3: 503-507.
- Olowolafe, E.A., (2008). Effects of using municipal waste as fertilizer on soil properties in Jos area, Nigeria. Resource Conservation Recycling. 52: 1015 1114.
- Omotayo, O.E., Chukwuka, K.S. (2009). Soil fertility restoration techniques in sub-Saharan Africa using organic resources. *African Journal of Agricultural Research*. 4: 144 -150.
- Oyeyiola, Y., Omueti, J. (2016). Phosphorus uptake and use efficiency by cowpea in phosphocompost and chemical fertilizer treated nutrient degraded acid soils. *Agriculture Research Technology*. 1: 1–8.
- Parentoni, S. N., Junior, C.L.S. (2008). Phosphorus acquisition and internal utilization efficiency in tropical maize genotypes. *Pesquisa Agropecuária Brasileira*. 43: 893 901.
- Pirastehc- Anoshen, H., Saed- Moucheshi, A., Pakniyat, H., Pessarakli, M. (2016). Stomatal responses to drought stress. Water Stress Crop Plants. 8: 24-40.
- Qin R.J., Stamp, P., Richner, W. (2005).Impact of tillage and banded starter fertilizer on maize root growth in the top 25 centimeters of the soil. *Agronomy Journal*.97: 674–683.

- Ragasa, C., Dankyi, A., Acheampong, P. Wiredu, A.N., Chapo-to, A., Asamoah, M., *et al.* (2013).
  Patterns of Adoption of Improved Maize Technologies in Ghana. International Food Policy
  Research Institute, Working paper 36.
- Ramegowda, V., Basu. S., Krishnan, A., Pereira, A. (2014). Rice growth under drought kinase is required for drought tolerance and grain yield under normal and drought stress conditions. *Plant Physiology.* 166:1634-1645.
- Ramesh, P. (2000). Sugarcane breeding institute, Coimbatore, India effect of different levels of drought during the formative phase on growth parameters and its relationship with dry matter accumulation in sugarcane. *Journal of Agronomy Crop Science*.185: 83–89.
- Ravi N., Basavarajappa R., Chandrashekar C., Harlapur S., Hosamani M, et al. (2012). Effect of integrated nutrient management on growth and yield of quality protein maize. Karnataka *Journal of Agricultural Sciences*. 25: 395-396.
- Reeuwijk, L. (2002). Procedures for soil analysis. International soil reference and information centre (6th ed). Food and Agriculture Organization of the United Nations.
- Rockstrom, J., Falkenmark, M., (2015). Increase water harvesting in Africa. *Nature*. 519: 283 285.
- Rockstrom, J., Karlberg, L., Wani, S.P., Barron, J., Hatibu, N., Oweis, T., Bruggeman, A., Farahani, J., Qiang, Z., (2010). Managing water in rainfed agriculture-The need for a paradigm shift. *Agricultural Water Management*. 97: 543 – 550.

- Rose, T. J., Wissuwa, M. (2012). Rethinking internal phosphorus utilization efficiency (PUE). A new approach is needed to improve PUE in grain crops. *Advanced Agronomy*. 116: 185 – 217.
- Sadras, V. O., McDonald, G., Sadras, V. O., McDonald, G. (2012). Water use efficiency of grain crops in Australia: principles, benchmarks and management. *Change*.11: 24-32.
- Sagar, K., Sharma, P. K. (2015). Effect of integration of organic and inorganic sources of nitrogen on growth, yield and nutrient uptake by Maize (*Zea mays L.*). *International Journal of Applied Sciences and Biotechnology*. 3:31-37.
- Sainju, U.M., Lenssen, A.W., Caesar-TonThat, T., Evans, R.G. (2009). Dryland crop yields and soil organic matter as influenced by long-term tillage and cropping sequence. *Agronomy Journal*.101: 243 -251.
- Salvagiotti, F., Cassman, K.G., Specht, J.E., Walters, D.T., Weiss, A., Dobermann, A. (2008). Nitrogen uptake, fixation and response to fertilizer N in soybeans: A review. *Field Crops Research*. 108: 1-13.
- Samson, H., H. Helmut. (2007). Drought effect on yield, leaf parameters and Evapotranspiration efficiency of cowpea. *Conference of International Agricultural Research for Development*. University of Kassel Witzenhause and University of Gotteingen.
- Sánchez Ó.J., Ospina D.A., Montoya S. (2017). Compost supplementation with nutrients and microorganisms in composting process. *Waste Management*. 69:136-153.

Sanchez, P.A., (2010). Tripling crop yields in tropical Africa. Nature Geoscience. 3: 299-300.

- Sanusi, S., Ch'ng, H.Y., Othman, S., (2018). Effects of incubation period and Christmas Island rock phosphate with different rate of rice straw compost on phosphorus availability in acid soil. *AIMS Agricultural Food* 3: 384-396.
- Sardans, J., Penuelas, J. (2012). The role of plants in the effects of global change on nutrient availability and stoichiometry in the plant-soil system. *Plant Physiology*. 160: 1741-1761.

SAS Institute Inc., (2004). SAS/Stat 9.2 User's Guide. SAS Institute Inc., Cary, NC.

- Sattar, A., Sher, A., Ijaz, M., Ul–Allah, S., Rizwan, M. S., Hussain, M., *et al.* (2020).Terminal drought and heat stress alter physiological and biochemical attributes in flag leaf of bread wheat.
- Schärer, M., Vollmer, T., Frossard, E., Stamm, C., Flühler, H., Sinaj, S. (2010). Effect of water composition on phosphorus concentration in runoff and water-soluble phosphate in two grassland soils. *European Journal of Soil Science*. 57:228–234.
- Schoebitz, M., Vidal G. (2016). Microbial consortium and pig slurry to improve chemical properties of degraded soil and nutrient plant uptake. *Journal of Soil Science and Plant Nutrition*. 16:226-236.
- Schröder, J.J., Smit, A.L., Cordell, D., Rosemarin, A. (2011). Improved phosphorus use efficiency in agriculture: A key requirement for its sustainable use. *Chemosphere*. 84: 822–831.
- Scotti, R., D, Ascoli, R., Bonanomi, G., Caceres, M.G., Sultana, S., Cozzolino, L., Scelza, R., Zoina, A., Rao, M.A. (2015). Combined use of compost and wood scraps to increase carbon stock and improve soil quality in intensive farming systems. *European Journal of Soil Science*.

- Sehgal, A., Sita, K., Kumar, J., Kumar, S., Singh, S., Siddique, K. H., et al. (2017). Effects of drought, heat and their interaction on the growth, yield and photosynthetic function of lentil (Lens culinaris Medikus) genotypes varying in heat and drought sensitivity. *Frontiers in Plant Sciences*. 8:1776-1198.
- Sehgal, A., Sita, K., Bhandari, K., Kumar, S., Kumar, J., Vara Prasad, P. V., *et al.* (2019). Influence of drought and heat stress, applied independently or in combination during seed development, on qualitative and quantitative aspects of seeds of lentil (Lens culinaris Medikus) genotypes, differing in droughts sensitivity. *Plant Cell and Environment*.42: 198–211.
- Setiyono, T.D., Walters, D.T., Cassman, K.G., Witt, C., Dobermann, A. (2010). Estimating maize nutrient uptake requirements. *Field Crops Research*. 118:158-168.
- Shenoy, V. V., Kalagudi, G. M. (2005). Enhancing plant phosphorus use efficiency for sustainable cropping. *Biotechnology Advanced*. 23: 501–513.
- Shrestha, R. K., Cooperband, L. R., MacGuidwin, A. E. (2010). Strategies to Reduce Nitrate Leaching into Groundwater in Potato Grown in Sandy Soils: Case Study from North Central USA. American Journal of Potato Research. 87:229–244.
- Sikuku, P.A., Onyango, J.C., Netodo, G.W. (2012). Yield components and gas exchange responses of Nerica rice varieties (*Oryza sativa L.*) to vegetative and reproductive stage water deficit. *Global journal of science. Frontier Research.* 12: 51–62.
- Simon, T., Czakó, A. (2014). Influence of long-term application of organic and inorganic fertilizers on soil properties. *Plant Soil Environment*. 60:314-319.

- Singh, C., Singh, P., Singh, R. (2017). Modern Techniques of Raising Field Crops. (2<sup>nd</sup> ed). New Delhi: Oxford and IBH Publishing Company. pp. 86.
- Soheil, R., Hossien, M. H., Gholamreza, S., Leila, H., Mozhdeh, J., Hassan, E. (2012). Effects of Composted municipal waste and its Leachate on Some Soil Chemical Properties and Corn Plant Responses. *Int. Journal of Agriculture: Research and Review*. 2: 801-814.
- Somerville, P.D., May, P.B., Livesley, S.J. (2018). Effects of deep tillage and municipal green waste compost amendments on soil properties and tree growth in compacted urban soils. *Journal of Environmental Management*. 227: 365-374.
- Souri M.K., Hatamian, M. (2019). Amino chelates in plant nutrition: a review. *Journal of Plant Nutrition*. 42:67–78.
- Souza, D.M.D., Morais, A.P.O., Matsushige, L., Rosa, L.A. (2016). Development of alternative methods for determining soil organic matter. *Revista Brasileira de Ciencia Do Solo*. 40: 1-17.
- Sperry, J. S., Love, D. M. (2015). What plant hydraulics can tell us about responses to climatechange droughts? *New Phytologist*. 207: 14–27.
- Sperry, J. S., Wang, Y., Wolfe, B. T., Mackay, D. S., Anderegg, W. R. L., Mcdowell, N. G., *et al.* (2016). Pragmatic hydraulic theory predicts stomatal responses to climatic water deficits. *New Phytologist.* 212: 577–589.
- Statistics Botswana. (2012). 2007 and 2008 Annual Agricultural Survey Report. Gaborone: Government Printers.

- Steduto, P., Hsiao, T. C., Fereres, E., Raes, D. (2012). Crop Yield Response to Water. Rome. (Irrigation and Drainage Paper 66).
- Sun, Y., Piao, S., Huang, M., Ciais, P., Zeng, Z., Cheng, L., Li, X., Zhang, X., Mao, J., Peng, S., Poulter, B., Shi, X., Wang, X., Wang, Y., Zeng, H. (2015). Global patterns and climate drivers of water-use efficiency in terrestrial ecosystems deduced from satellite-based datasets and carbon cycle models. *Global Ecology and Bio-geography*. 25:311–323.
- Suriyagoda, L.D.B., Ryan, M.H., Renton, M., Lambers, H. (2011). Above-and belowground interactions of grass and pasture legume species when grown together under drought and low phosphorus availability. *Journal of Plant and Soil*. 348: 281–297.
- Sustainable Agriculture and Rural Development. (2007). Conservation Agriculture in Africa -Policy Brief# 18. Dar es Salaam: Sustainable Agriculture and Rural Development.
- Sutton, M.A., Bleeker, A. (2013). Environmental science. The shape of nitrogen to come. *Nature*. 494: 435-437.
- Swarbreck, S.M., Wang, M., Wang, Y., Kindred, D., Sylvester-Bradley R., Shi, W., Varinderpal-Singh, Bentley A.R., Griffiths, H. (2019). A roadmap for lowering crop nitrogen requirement. *Trends in Plant Science*. 24: 892–904.
- Syers, J.K., Johnston, A.E., Curtin D. (2008). Efficiency of soil and fertilizer phosphorus use: reconciling changing concepts of soil phosphorus behaviour with agronomic information. Fertilizer and Plant Nutrition Bulletin18. Food and Agriculture Organization of the United Nations (FAO), Rome.

- Tan, K.H. (2005). *Soil Sampling, Preparation, and Analysis*, (2<sup>nd</sup> ed). CRC Press: Boca Raton, FL, USA.
- Tandzi, N.L., Ngonkeu, M.E., Nartey, E., Yeboah, M., Ngeve, J., Mafouasson, H.A., Nso ngang, A Bassi, O., Gracen, V. (2015). Farmers' adoption of improved maize varieties in the humid forest area of Cameroon. *International Journal of Scientific Engineering and Applied Science* 1: 17-28. ISSN: 2395-3470.
- Tembe, K.O., Chemining'wa, G.N., Ambuko, J., Owino, W. (2017). Effect of water stress on yield and physiological traits among selected African tomato (*Solanum lycopersicum*) land races. *International Journal of Agronomy and Agricultural Research*. 10: 78-85.
- Thalmann, M, Santelia, D. (2017). Starch as a determinant of plant fitness under abiotic stress. The New Phytologist.
- Thangarajan, R., Bolan, N.S., Tian, G., Naidu, R., Kunhikrishnan, A. (2013). Role of organic amendment application on greenhouse gas emission from soil. *Science of the Total Environment*. 465: 72-96.
- Trupiano D., Cocozza C., Baronti S. (2017). "The effects of biochar and its combination with compost on lettuce (*Lactuca sativa* L.) Growth, soil properties, and soil microbial activity and abundance," *Hindawi International Journal of Agronomy*.: Article ID 3158207, 1-12.
- USEPA (2014). Inventory of U. S. Greenhouse Gas Emissions: 1990–2013. Washington, DC: USEPA.

- Vadez, V., Deshpande, S. P., Kholova, J., Hammer, G. L., Borrell, A. K., Talwar, H. S. (2011). Stay-green quantitative trait loci's effects on water extraction, transpiration efficiency and seed yield depend on recipient parent background. *Functional Plant Biology*. 38: 553–566.
- Van- Camp, L., Bujarrabal, B., Gentile, A. R., Jones, R.J.K. (2004). Reports of the Technical Working Groups Established under the Thematic Strategy for soil protection. *European Environment Agency.* 3: 872- 882.
- Van-Schoor, L. (2009). Effect of biological amendments on soil microbial properties and performance of pome fruit trees. Dissertation presented for the Degree of Doctor of Philosophy (Agriculture) at Stellenbosch University, Cape Town, South Africa.
- Wang, F., Kang, S., Du, T., Li, F., Qiu, R. (2011). Determination of comprehensive quality index for tomato and its response to different irrigation treatments. *Agricultural Water Management*. 98: 1228-1238.
- Wang, J., Liu, W.Z., Dang, T.H. (2011). Responses of soil water balance and precipitation storage efficiency to increased fertilizer application in winter wheat. *Plant and Soil*. 347: 41-51.
- Wang, J.Z., Lu C.A., Zhang, W.J, Feng, G., Wang, X.J., Xu, M.G. (2016). Decomposition of organic materials in cropland soils across China: A meta-analysis. *Acta Pedologica Sinica*. 53: 16–27.
- Wang, X., Shen, J., Liao, H. (2010). Acquisition or utilization, which is more critical for enhancing phosphorus efficiency in modern crops. *Plant Science*. 179: 302–306.

- Wang, X.C., Deng, X.Y., Pu, T., Song, C., Yong, T.W., Yang, F., Sun, X., Liu, W.G., Liu, Y.H.
  (2017). Contribution of interspecific interactions and phosphorus application to increasing soil phosphorus availability in relay intercropping systems. *Field Crop Research*. 204: 12–22.
- White, P. J., Broadley, M. R., Greenwood, D. J., Hammond, J.P. (2005). Genetic modifications to improve phosphorus acquisition by roots. Proceedings 568 of International Fertilizer Society, New York.
- Wison, C., Hui, D., Nwaneri, E., Wang, J., Deng, Q., Duseja, D., Tegegne, F., (2012). Effects of planting dates, densities and varieties on Eco physiology of pigeon pea in the South Eastern United State. Agricultural Sciences. 3:147 – 152.
- Wortman, S.E., Galusha, T.G., Mason, S.C., Francis, C.A. (2012a). Soil fertility and crop yields in long-term organic and conventional cropping systems in eastern Nebraska. *Renewable Agriculture and Food Systems*. 27: 200-216.
- Xu, C., Mou, B. (2016). Vermicompost affects soil properties and spinach growth, physiology, and nutritional value. *Horticultural Science*. 51:847-855.
- Yada, G.L. (2011). Establishing optimum plant populations and water use of ultra-fast maize hybrid (Zea mays L.) under irrigation. PhD Thesis. University of Free State, South Africa.
- Yan, W., Zhong, Y., Shangguan, Z. (2016). A meta-analysis of leaf gas exchange and water status responses to drought. *Scientific Reports*. 6:20917- 20926.

- Yang, Y., Guo, J., Wang, G., Yang, L., Yang, Y. (2012). Effects of drought and nitrogen addition on photosynthetic characteristics and resource allocation of *Abies fabri* seedlings in eastern Tibetan plateau. *New.* 43: 505–518.
- Yang, Y., Tang, M., Sulpice, R., Chen, H., Tian, S., Ban, Y. (2014). Arbuscular mycorrhizal fungi alter fractal dimension characteristics of *Robinia pseudoacacia* L. Seedlings through regulating plant growth, leaf water status, photosynthesis, and nutrient concentration under drought stress. *Journal of Plant Growth Regulators*. 33:612-625.
- Yared, A, Y., Staggenborg, S.A., Prasad, V.P.V. (2010). Grain sorghum water requirements and responses to drought stress. *A review. Plant Management Network International*.
- Ye, Y.S., Liang, X.Q., Chen, Y.X., Liu, J., Gu, J.T., Guo, R., Li, L. (2013). Alternate wetting and drying irrigation and controlled -release nitrogen fertilizer in late-season rice. Effects on dry matter accumulation, yield, water and nitrogen use. *Field Crop Research*.144: 212-224.
- Zaccardelli, M., Villecco, D., Celano, G., Scotti, R. (2013b). Soil amendment with seed meals:
  Short term effects on soil respiration and biochemical properties. *Applied Soil Ecology*.
  72: 225-231.
- Zaidi, P.H., Jat, M.L., Jat, H.S., Lenka, D., Swain, D. (2017). Package of Practices for Profitable Maize Cultivation- a Field Manual. Hyderabad, India: CIMMYT.
- Zamir, S.I. (1998). Effect of plant spacing on yield and yield components of maize. M.Sc. Agri. Thesis, pp.98-101. Department of Agronomy. University of Agriculture. Faisalabad, Pakistan.

- Zandalinas, S. I., Balfagón, D., Arbona, V., Gómez–Cadenas, A. (2017). Modulation of antioxidant defence system is associated with combined drought and heat stress tolerance in citrus. *Frontiers in Plant Science*.8: 953-963.
- Zhang, H. M., Wang, B, R., Xu, M. G., Fan, T. L. (2009). Crop yield and soil responses to long-term fertilization on a red soil in southern China. *Pedosphere*. 19:199–207.
- Zhang, H., Chen, C., Gray, E.M., Boyd, S.E., Yang, H., Zhang, D. (2016). Roles of biochar in improving phosphorus availability in soils: a phosphate absorbent and a source of available phosphorus. *Geoderma*: 276: 1-6
- Zhang, S.Y., Zhang, G.C., Gu, S.Y., Xia, J.B., Zhao, J.K. (2010). Critical responses of photosynthetic efficiency of goldspur apple tree to soil water variation in semiarid loess hilly area. *Photosynthetica* .48: 589–595.
- Zhang, W., Tian, Z., Pan, X., Zhao, X., Wang, F. (2013). Oxidative stress and non-enzymatic antioxidants in leaves of three edible canna cultivars under drought stress. *Horticulture, Environment and. Biotechnology.* 54: 1-8.
- Zhang, X., Lei, L., Lai, J., Zhao, H., Song, W. (2018). Effects of drought stress and water recovery on physiological responses and gene expression in maize seedlings. BMC *Plant Biology*. 18: 1–16.
- Zhang, Z., Dong, X., Wang, S., Pu, X. (2020). Benefits of organic manure combined with biochar amendments to cotton root growth and yield under continuous cropping systems in Xinjiang. *Scientific Reports*.10: 4718-4728.

- Zhang, Z.P., Hua, Q.I., Zhang, Y., Sun, S.X., Yang, G.H. (2009). Effects of water stress on photosynthetic rate and water use efficiency of maize. *Acta Agriculture Boreali Sinica*. 24:155-158.
- Zhao, D., Li, Y. (2015). Climate change and sugarcane production: Potential impact and mitigation strategies. *International Journal of Agronomy*.2:1-10.
- Zhao, F., Zhang, D., Zhao, Y., Wang, W., Yang, H., Tai, F., et al. (2016). The difference of physiological and proteomic changes in maize leaves adaptation to drought, heat, and combined both stresses. *Frontiers in Plant Science*. 7:1471-1490.
- Zotarelli, L., Scholberg, J. M., Dukes, M. D., Mun, R. (2007). Monitoring of nitrate leaching in sandy soils: comparison of three methods. *Journal of Environment Quality*. 36:953–962.